

COLUMBIA LIBRARIES OFFSITE
HEALTH SCIENCES STANDARD



HX64169693

RG586 B22 1903 Uterine und tubal ge

RECAP

UTERINE
AND
TUBAL GESTATION

RG586 B22

RG586

B22

Columbia University
in the City of New York

Copy

College of Physicians and Surgeons



Reference Library



Digitized by the Internet Archive
in 2010 with funding from
Open Knowledge Commons

<http://www.archive.org/details/uterinetubalgest00band>

UTERINE AND TUBAL GESTATION

A STUDY OF THE EMBEDDING AND DEVELOPMENT OF THE HUMAN
OVUM, THE EARLY GROWTH OF THE EMBRYO, AND
THE DEVELOPMENT OF THE SYNCYTIUM
AND PLACENTAL GLAND

BY

SAMUEL WYLLIS BANDLER, M.D.
Instructor in Gynecology, N. Y. Post-Graduate Medical School

ILLUSTRATED BY NINETY-THREE DRAWINGS

Dr. Francis Huber
109 E. 17th St.
N. Y. City

NEW YORK
WILLIAM WOOD & COMPANY
MDCCCCIII

COPYRIGHTED, 1903,
BY
SAMUEL WYLIS BANDLER.

PRESS OF
STETTNER BROTHERS,
52-58 DUANE ST.,
NEW YORK.

DEDICATED TO
FERDINAND GRAF von SPEE,
PROFESSOR OF ANATOMY IN THE UNIVERSITY OF KIEL,
IN ADMIRATION OF HIS MOST VALUABLE CONTRIBUTIONS TO OUR SCIENTIFIC
KNOWLEDGE AND IN GRATITUDE FOR PERSONAL KINDNESSES.

PREFACE.

Many of these pages appeared in THE AMERICAN JOURNAL OF OBSTETRICS AND GYNECOLOGY under the title, "On the Etiology, Histology, and Usual Course of Ectopic Gestation." Enough has been added to make the processes antedating and accompanying uterine gestation fairly complete and up to date. The essential features in this channel, as regards the earliest stages, are to be credited to Spee. Attempt has been made to aid in the decision concerning several doubtful problems, particularly as to the origin of the syncytium. Attention has been paid to the decided probability that the placenta is a gland with potentials of great importance from the standpoint of secretion. Some personal views concerning the formation of villi and the blood-forming function of the trophoblast have been brought forward with a full realization that criticism and further observations are to prove them correct or otherwise. In spite of the great labors of gifted investigators, final decision is yet to be given on many points, so changeable are the processes at various stages and so probable is it that many ova are pathological.

To Minot, Mall, v. Spee, and others we are greatly indebted for pioneer work on allied questions. The subject of chorioma, or chorio-epithelioma, has been introduced, because in its microscopical character it so closely reproduces many normal conditions.

I am under great personal obligation to Dr. A. Brothers for the gross specimens which furnished the material for Part II and a portion of Part III. He placed all of his large material and histories at my disposal, for which kindnesses I here express my sincerest thanks.

SAMUEL WYLLIS BANDLER.

CONTENTS.

PART I.

THE ESSENTIALS OF UTERINE GESTATION.

| CHAP. | | PAGE. |
|-------|---|-------|
| I. | The Processes Antedating Uterine Gestation..... | 1 |
| | A. The Trophic Influence of the Ovary..... | 1 |
| | B. Constitutional Changes Dependent on the Ovary..... | 6 |
| | C. Menstruation | 9 |
| | D. The Action of Ovarian Secretion on the Endometrium..... | 12 |
| | E. The Relation of Ovulation and Menstruation..... | 13 |
| | F. Ovulation | 15 |
| II. | The Embedding of the Ovum in the Guinea-Pig..... | 18 |
| | A. Uterus of the Guinea-Pig..... | 18 |
| | B. The Embedding of the Guinea-Pig's Ovum..... | 20 |
| III. | The Embedding of the Human Ovum..... | 32 |
| | A. The Uterus | 32 |
| | Decidua Menstrualis | 33 |
| | Decidua Graviditatis in the First Week..... | 33 |
| | Decidua Graviditatis | 35 |
| | B. The Embedding of the Human Ovum..... | 37 |
| | Ovum in the Earliest Stages..... | 38 |
| | Capsularis | 39 |
| | The Enveloping Zone..... | 39 |
| IV. | The Early Development of the Human Ovum..... | 40 |
| | Division into Embryonal and Extra-Embryonal Areas..... | 45 |
| V. | The Trophoblast in the Ova of Animals..... | 46 |
| | The Earliest Development of the Ectoblastic Extra-Embryonal Area | 46 |
| VI. | The Trophoblast of the Human Ovum..... | 50 |
| | The Earliest Development of the Ectoblastic Extra-Embryonal Area of the Ovum..... | 50 |
| | Trophoblast | 50 |
| | The Primary Intervillous Space..... | 53 |
| VII. | The Further Development of the Human Ovum..... | 57 |
| | The Early Development of the Embryonal Area..... | 57 |
| VIII. | The Chorionic Villi..... | 66 |
| | A. Early Development | 66 |
| | B. In the Fourth Week of Uterine Gestation..... | 68 |
| IX. | The Membrana Chorii..... | 75 |
| X. | The Blood-Forming Function of the Trophoblast..... | 82 |
| XI. | The Further Development of the Uterine Placenta..... | 89 |

| CHAP. | PAGE. |
|---|-------|
| XII. The Placenta | 93 |
| XIII. The Umbilical Vessels and Cord..... | 99 |
| A. The Umbilical Vessels..... | 99 |
| B. The Umbilical Cord..... | 101 |
| C. The Amnion | 102 |
| XIV. Gross Anatomy of the Placenta..... | 103 |

PART II.

THE ESSENTIALS OF TUBAL GESTATION.

| | |
|--|-----|
| I. Processes Antedating Gestation in the Tube..... | 109 |
| Etiology | 109 |
| II. Varying Views Concerning the Histology of Tubal Gestation. | 114 |
| The Decidua | 114 |
| Embedding of the Ovum, the Reflexa or Capsularis..... | 115 |
| Intervillous Space | 115 |
| Villi | 116 |
| Deportation | 117 |
| Conclusions | 119 |
| III. Embedding of the Ovum and the Development of Extra-Embryonal structures | 121 |
| I. The Columnar Type of Tubal Gestation..... | 121 |
| II. The Intercolumnar Type of Tubal Gestation..... | 127 |
| III. The Centrifugal Type of Tubal Gestation..... | 131 |
| Conclusions | 135 |
| IV. The Usual Course of Tubal Gestation..... | 137 |

PART III.

OVARIAN AND PLACENTAL SECRETION.

| | |
|--|-----|
| The Relation of the Chorionic Epithelium to Chorio-Epithelioma | 145 |
| Chorio-Epithelioma or Chorioma..... | 151 |

LIST OF ILLUSTRATIONS

EMBEDDING OF GUINEA-PIG'S OVUM.

| FIG. | PAGE. |
|---|-------|
| 1. Uterine horn (Spee)..... | 18 |
| 2. Compact zone of uterine lining (Spee)..... | 19 |
| 3. Ovum free in uterine cavity (Spee)..... | 20 |
| 3a. Ovum adherent to uterine epithelium (Spee)..... | 21 |
| 4. Ovum partially embedded (Spee)..... | 22 |
| 5. Further stage of embedding (Spee)..... | 23 |
| 6. Partially embedded ovum (Spee)..... | 24 |
| 7. Ovum surrounded by symplasma (Spee)..... | 25 |
| 8. Almost embedded ovum (Spee)..... | 26 |
| 9. Ovum entirely under uterine epithelium (Spee)..... | 27 |
| 10. Demarkation of symplasma (Spee)..... | 28 |
| 11. Embedded ovum (Spee)..... | 29 |
| 12. Completely embedded, growing ovum (Spee)..... | 30 |
| 13. Rapidly-growing ovum (Spee)..... | 32 |

EMBEDDING OF HUMAN OVUM.

| | |
|---|----|
| 14. Menstrual decidua (Abel)..... | 34 |
| 14a. Decidua in the second month (Abel)..... | 35 |
| 14b. Gland in the decidua graviditatis..... | 36 |
| 15. Schematic; embedding human ovum (Peters)..... | 37 |
| 16. Schematic; embedded human ovum (Peters)..... | 38 |

THE EARLY DEVELOPMENT OF THE HUMAN OVUM.

| | |
|---|----|
| 17. Schematic; ovum with ectoblast and entoblast..... | 40 |
| 17a. Schematic; ovum with amnion..... | 40 |
| 17b. Schematic; ovum with separating amnion..... | 41 |
| 17c. Schematic; ovum with ectoblast, entoblast, amnion, germinal plate, but no mesoblast..... | 41 |
| 18. Schematic; ovum with beginning growth of mesoblast..... | 42 |
| 18a. Schematic; ovum with three germinal layers..... | 42 |
| 19. Ovum with three germinal layers (Peters)..... | 43 |
| 19a. Schematic; ovum with mesoblastic periembryonal slit..... | 45 |

HUMAN TROPHOBlast.

| | |
|---|----|
| 20. Trophoblast, etc., of three-day ovum (Peters)..... | 50 |
| 21. Central portion of trophoblast layer (Peters)..... | 51 |
| 22. Trophoblast infiltrated with blood lacunæ (Peters)..... | 52 |
| 23. Change of trophoblast to syncytium (Peters)..... | 53 |
| 24. Scheme of earliest stage of placenta (Peters)..... | 54 |

LIST OF ILLUSTRATIONS.

EARLY DEVELOPMENT OF THE EMBRYONAL AREA.

| | PAGE |
|--|------|
| 25. Ovum <i>v. H.</i> of Spee..... | 57 |
| 26. Longitudinal section through 25..... | 58 |
| 27. Fig. 26 enlarged..... | 58 |
| 28. Germinal plate of ovum <i>v. H.</i> of Spee..... | 59 |
| 29. Ventral curve in the germinal plate..... | 69 |
| 29a. Embryo <i>Gle</i> of Spee..... | 60 |
| 29b. Embryo <i>Gle</i> of Spee..... | 61 |
| 30. Three layers of a human embryo (Keibel)..... | 62 |
| 30a. The forming of the intestine (Kollman)..... | 63 |
| 30b. Human embryo 2-4 mm. long (His)..... | 63 |
| 30c. Caudal end of embryo 3 mm. long (Keibel)..... | 64 |
| 30a. Schematic; embryo with ventral surface toward the abdominal pedicle | 64 |
| 30e. Schematic; embryonal formation (Waldeyer)..... | 65 |

THE CHORIONIC VILLI.

| | |
|---|----|
| 31. Schematic; later stage of placental development (Peters)..... | 67 |
| 31a. A well-developed villus..... | 68 |
| 31b. An outgrowth on the membrana chorii..... | 69 |
| 32. Villus composed of trophoblast cells..... | 70 |
| 32a. Villus with protoplasmatic trophoblast cells..... | 70 |
| 33. Villus with beginning centre of mesoderm..... | 71 |
| 34. Older villus | 71 |
| 35. Villus in various stages..... | 72 |
| 35a. Cell group with invading syncytium..... | 72 |
| 35b. Further stage of 35a..... | 73 |
| 35c. Further stage of 35b..... | 73 |

MEMBRANA CHORII.

| | |
|---|----|
| 36. Membrana chorii of five-weeks ovum..... | 78 |
| 37. Vacuoles in membrana chorii..... | 79 |
| 38. Membrana chorii of tubal ovum..... | 85 |

FURTHER DEVELOPMENT OF UTERINE PLACENTA.

| | |
|--|----|
| 39. Uterus and fetal sac of seventh week..... | 89 |
| 40. Decidua vera | 90 |
| 41. Cell groups invading the serotina..... | 91 |
| 42. Syncytial cells invading the serotina..... | 91 |

THE PLACENTA.

| | |
|---|----|
| 43. Uterus and placenta at full term..... | 95 |
| 44. Utero-placenta junction at full term..... | 95 |
| 45. Invading fetal cells..... | 96 |
| 46. Placental villi | 96 |

THE UMBILICAL VESSELS AND CORD.

| | PAGE. |
|-------------------------------------|-------|
| 46a. Young embryo, with amnion..... | 101 |
| 46b. Schematic; umbilical cord..... | 102 |
| 46c. Scheme of placenta..... | 104 |

PART II.

| | |
|--|-----|
| 47. Columnar type of tubal gestation..... | 121 |
| 47a. Ovum 47, magnified..... | 122 |
| 48. Tubal ovum 47..... | 123 |
| 49. A more peripheral section of 47..... | 123 |
| 50. A still more peripheral section of 47..... | 124 |
| 51. Most peripheral section of 47..... | 125 |
| 52. Change of trophoblast to syncytium..... | 126 |
| 53. Intercolumnar Type of Tubal Gestation..... | 127 |
| 54. Serotinal area of 53..... | 128 |
| 54a. Typical area of 54..... | 128 |
| 55. Serotinal area of 53..... | 129 |
| 55a. Area of Fig. 55..... | 130 |
| 56. Tubal gestation sac with fetus— <i>Centrifugal Type of Tubal Gestation</i> | 131 |
| 56a. Tubal ovum with capsularis..... | 132 |
| 57. Capsularis and tube wall of 56a..... | 133 |
| 58. Advancing trophoblast cells of 57..... | 134 |
| 59. Serotinal area of 56a..... | 135 |

PART III.

CHORIOMA.

| | |
|------------------------------------|-----|
| 60. Typical form of chorioma..... | 152 |
| 61. Fig. 60 magnified..... | 153 |
| 62. Fig. 61 magnified..... | 154 |
| 63. Fig. 60 magnified..... | 155 |
| 64. Atypical form of chorioma..... | 156 |



PART I.

THE ESSENTIALS OF UTERINE GESTATION.

CHAPTER I.

THE PROCESSES ANTEDATING UTERINE GESTATION.

A. THE TROPHIC INFLUENCE OF THE OVARY.

Halban transplanted the ovaries in newly born guinea-pigs to see what effect transplantation and castration would exert upon the development of the genitalia and the breasts. The comparisons with non-castrated animals are very interesting. In the *castrated guinea-pig* the breasts were later found to be one-fourth the normal size; the genitalia were small, the vulva was one-third smaller than normal; the uterus was as small as at birth, showing very slight development of muscle and endometrium, and containing no ciliated cells. The breasts showed no glandular tissue, the mammillæ were hypoplastic, the vaginal mucous membrane showed both squamous and cylindrical epithelium—a novel condition. When the ovaries were transplanted under the skin, the uterus was well developed, the tubes were normal, the vulva was large, as were the mammillæ likewise. In the transplantation of one ovary a part of the uterus and a piece of the tube had likewise been taken, and these grew in a normal manner, and increased in size. The transplanted piece of uterus contained well-developed muscle and normally secreting glands. The portion of tube showed muscle, epithelium, and ciliated cells. Halban thus showed the *influence of the ovary, as a secreting organ, upon the uterus, vagina, vulva, and mamma*. He considers it probable that the ovarian secretion acts upon the uterus; the uterine secretion acting in turn upon the mammary gland.

Ribbert implanted the mamma of a young guinea-pig, with its covering of skin, into a cut near the ear. The wound healed, and five months after the operation, the animal having borne two young, this mamma secreted milk normally, a proof that the *connection between the breasts and the ovary and uterus is to be found in no other channels than those of the circulation*.

Hegar and Kehrer showed that in newly-born animals, after castration, the uterus does not develop, but remains at the same stage, or even undergoes atrophy.

Sokoloff castrated rabbits and dogs and found that, a short time after, they became fat and apathetic, and grew constantly worse. Subsequent examination of the uterus divulged muscular atrophy, especially of the circular layer; the vessels were fewer, the walls thicker, the lumen was small and even obliterated. The mucosa, however, remained unchanged. *When only one ovary had been removed, there was no change in the sexual development of these animals, the young ones evidencing a normal and constant development of the genitalia.*

Jentzer and Beuttner examined the organs of cows within one year to twenty-two months after castration, and found *all the layers of the uterus atrophied and substituted by connective tissue.* The stratum vasculare had greatly changed; the atrophy of the mucosa, while not constant, was in most cases acute. They castrated rabbits, gave subsequently hypodermatic injections of ovarin, and yet the uterus atrophied. The objections to considering this last procedure conclusive are: 1. That the ovarin of cows was used. 2. That it was given hypodermically. 3. That there was no sufficient increase in the amounts used. The castration of other rabbits showed that the cylindrical epithelium became lower, the protoplasm degenerated, these processes becoming more defined the longer after castration the animals were examined. The muscularis was found to be atrophied; the larger vessels were gone. Although glands were present, the mucosa showed decided atrophy. In dogs castration produced atrophy of the muscle, but only after three or four months; the epithelium of the uterus was lower, the protoplasma cloudy, the glands were degenerated, and the vessel walls thickened.

Knauer transplanted the ovaries of rabbits and dogs between the fasciae of the abdominal wall and into the mesometrium, being careful to remove absolutely every bit of ovarian structure. In the mesentery he fastened the ovary with two sutures between two folds of peritoneum, the ovaries being then nourished through endosmosis or through plasmatic circulation. New vessels grew into the ovarian tissue and furnished its subsequent support; this change began as early as the fourth day. Examination at various periods showed that a small part of each ovary usually degenerated, and new connective tissue appeared in place of the lost cells. *In all cases in which a complete degeneration of the ovaries was found, atrophy of the breasts and of the genitalia had occurred.* The muscle of the uterus was atrophied, the

intermuscular connective tissue was increased, the mucous membrane was atrophied—changes like those which occurred after double castration. *Retention of function* on the part of the transplanted ovaries was always evidenced by the growth of follicles in a normal manner, by the ripening of the follicles, and by the discharge of the ova. *In all such cases the normal character of the breasts, of the uterus, and of the genitalia was preserved*, and in the younger animals all these organs underwent a natural development. In one dog with well-developed breasts, thirteen months after transplantation an opening of the abdomen showed the right ovary to be of normal size and to contain three follicles. The left ovary was small. Two months later coitus took place, and after two more months two well-developed young were born. Three years after transplantation had been done, this animal on examination showed externally a normal condition of development of the breasts and the genitalia. The right ovary was the size of a pea and contained follicles; the left ovary was larger than on the previous examination. The right ovary was covered with germinal epithelium; many follicles but no primary follicles being present. A like condition was found in the left ovary. The uterus was normal in every way; the mucous lining contained many glands and ciliated epithelium. The breasts were of normal character, with normal secretion and glands. Knauer's results proved that the preservation to the organism of the functioning ovaries preserves the breasts, the genital organs, and the sexual instinct, *a result possible only through the absorption into the circulation of ovarian secretion, and that the function which the ovary exerts upon the body stands in closest relation to its ability to form ripe ova*. Ovarian tissue which has ceased to develop ova has lost its other function, that is, secretion. Therefore the numerous nerves of the ovary are in all probability only vessel nerves. Knauer transplanted in sixteen cases the ovaries of animals into each other, obtaining the above good results in only two cases, since in the others the ovaries degenerated. This shows that a certain relation exists between the cells of one and the same body. Although difficult, transplantation of ovaries from one animal to another is possible.

Morris removed the adnexa in a human patient and sewed a piece of one ovary into the stump of the right tube. One month later the patient became gravid, aborting in the third month. Into a twenty-year-old girl with uterus infantilis and rudimen-

tary adnexa, with suppressio mensium, he transplanted a piece of the ovary of a thirty-year-old patient, fixing it in the fundus of the uterus. Eight weeks later a profuse menstruation, lasting ten days, occurred; after six weeks a normal one lasting five days; the third one five days later. The fourth and fifth menstruations were normal, at intervals of four weeks.

Glass transplanted an ovary of a seventeen-year-old girl into a twenty-nine-year-old patient from whom both ovaries had been removed two years before. The ovary was fixed in its normal situation through an opening in the sac of Douglas, per vaginam. Sixteen days later occurred a bleeding lasting two days; six months afterward a menstruation of three days; eight months after operation the patient showed good color, was of good appearance, with a return to former mental and body conditions, after two years of artificial climax.

Dudley implanted an ovary into the fundus of the uterus after removal of a pyosalpinx duplex, and regular menstruation continued.

An examination of the uterus after castration in women shows an atrophy of the cervix, an immediate atrophy of the corpus, a sclerosis of the vessels which show a growth of the intima, and an endarteritis obliterans, especially in the larger vessels. Few glands are present, and the connective tissue is increased.

Gottschalk found an exceptional condition in one case, in which the muscularis was well preserved, although the mucous lining had almost entirely disappeared.

Cholmogoroff found the uteri at climacterium to contain a large amount of connective tissue, the vessels being greatly sclerosed. I have found the same condition in a high degree in two cases of my own.

Normal menstruation is absolute evidence of the presence of a normally functioning ovary. The absence of ovaries, a poor development of these glands, an insufficient secretion of ovarian substance, or a diversion of ovarian secretion to other organs of the body, always causes a failure of uterine development or uterine atrophy. It is possible that, with partial or total absence or atrophy of the uterus or other genital organs, normal ovaries may be present. It is likewise possible for poorly developed ovaries to be the result of a failure of development of the body in general or to be simply a failure of development of the ovary itself, through embryonal disturbances.

Winckel has shown that, in the development of the uterus and

tubes, the situation of the Wolffian body close to the ducts of Müller may influence, to a very great degree, their growth and is a frequent cause of malformation. The early completion of the Wolffian bodies, their opening into the sinus urogenitalis, the growth of the Müller's ducts along the Wolffian, and their crossing at that spot where the union of Müller's ducts finds its upper limit, are anatomical embryological factors easily recognized as causes of uterine maldevelopment and hypoplasia. Further, the origin of the ligamentum ileo-genitale rotundum at this upper limit, its close union with the ducts of Müller, the fact that its line of development in a measure opposes the union of the ducts, in addition to tension, pressure, and torsion exerted by the neighboring organs, such as the Wolffian bodies, the bladder, the ureters, the vessels and nerves of the uterus and rectum, are important factors influencing the development of the uterus. In addition, Winckel recognizes the occurrence of abnormal cells in the septum between the ducts of Müller, and general hypoplasia of the vessel system, as additional causes of maldevelopment. Aside from secondary atrophy, the result of constitutional diseases, the embryological cause here mentioned, and the forms associated with general hypoplasia, *we recognize in the ovary and its secretion the factor which governs the development of the uterus, the genitalia, and the breasts, and the factor which is concerned in the preservation of these organs and the regulation of menstruation.*

Eberlin¹ describes a patient with vaginal defects, who, in her eighteenth year, had vicarious menstruation from the nose at regular intervals for six months, after which no recurrence was noted. Her mammae and external genitalia were small. She suffered from molimina menstrualia every three weeks, at first for periods of three to four days. Eventually severe pain was experienced constantly, associated with sickness at the stomach and vomiting. Laparotomy divulged a uterus rudimentarius, with absence of the adnexa of the left side. The right tube and ovary were almost normal. The uterus was 3.5 centimetres in length, differing microscopically in nowise from the normal as regards muscle, but possessing no cavity. The ovary showed a thickening and a hyaline degeneration of the vessel walls. In addition to the corpus luteum, very few follicles but no Graafian follicles were present. *After castration all the annoying symptoms disappeared.*

Fritsch operated in a like case in which severe molimina men-

¹Vineberg, AM. JOUR. OF OBSTET., No. 4, 1898.

stralia, sickness at the stomach, and rectal bleedings were the indications.

In all, Eberlin finds twenty-one cases of this sort associated with vaginal defect to have been operated upon. The ovary in the above-cited case of Eberlin, though containing few follicles, nevertheless secreted sufficiently to cause the severe symptoms. Since no endometrium was present, and no excretion by that channel took place, the ovarian secretion, through cumulative action, was responsible for the severity of the pain. *Whatever part the uterus took in these attacks must have been the result of uterine contraction*, since congestion alone is not responsible for so much pain, as is evidenced by the absence of such suffering in pure congestions of the uterus, associated with metritis, etc.

B. CONSTITUTIONAL CHANGES DEPENDENT ON THE OVARY.

Were we to consider the relation between the ovary and the uterus, and between the genitalia and other organs to exist through the medium of the cerebro-spinal or sympathetic systems, why should we find the walls of the tubes thin and containing more connective tissue in cases of poorly developed ovaries and at the menopause, when no primary follicles and no Graafian follicles are present in the ovaries? *The uterus then shrinks, the walls become thin and flabby, the mucous membrane atrophies, the connective-tissue elements are more prominent, and the ciliated epithelium disappears.* Is it not extraordinary to make the occurrence of these decided changes dependent upon a cessation of ovarian congestion? That these differences are not due to the lack simply of reflex stimulation on the part of the ovary is sufficiently evidenced by the constitutional changes occurring at puberty, at menstruation, during pregnancy, and at the menopause. The relation between the changes occurring at puberty and at the menopause, before menstruation and after menstruation, during pregnancy and after pregnancy, show a decided resemblance. Until shortly before each menstrual period, temperature, pulse, muscular activity, lung capacity, and the excretion of urea increase, and reach their maximum two to three days before the appearance of blood. During this period we find hyperemia, edema, increased activity of the ovary, changes in all the mucous membranes, and increased function of the glandular apparatus. The occurrence of swelling of the breasts, tenderness of the abdomen, even pain and the passage from the vagina of increased mucus, sometimes mixed with blood,

prove at the beginning of each menstrual period a wave movement and an increased blood tension by no possibility due to reflex causes. During and after menstruation regressive changes are evident, and the excretion of nitrogenous elements is diminished.

During pregnancy we have an increased amount of the watery elements of the blood, an increased proportion of fibrin, a diminished amount of albumin, an increase in the white blood cells, a genuine increase in the number of red blood cells and in the amount of hemoglobin.

Before labor the temperature is highest in the last three months of pregnancy, and there is an increase in the elements of the body, equal to one-thirteenth of the body weight. This increase is due to serous infiltration, and to the increased ability of the body to form organized tissue. Post partum, after a temporarily short rise, the temperature is lower, the blood pressure sinks, and becomes normal on the ninth day. The loss in body weight is equal to one-ninth of the weight of the pregnant person. For labor and post partum together, there is a loss of weight nearly equal to one-fifth of the body weight at full term. After labor there is a diminution of tissue change and a diminution in the amount of urine.

Therefore as regards temperature, blood pressure, body weight, the amount of urine secreted, etc., there is always a similar increase before menstruation, and a like decrease in intensity during and after menstruation, as during and after parturition, so that Virchow has well characterized menstruation as being a labor *en miniature*.

According to Loewenthal, who believes that every ovum is embedded in the mucous membrane of the uterus, we may say that *labor is a menstruation in which a fully developed ovum is expelled*. The coincidence of labor with a menstrual period seems therefore natural. The resemblance of the mechanical processes concerned in menstruation and in labor is remarkable, and these changes can be due only to the stimulative effect of the ovarian secretion, acting not alone upon the uterus and the genital organs, but likewise upon the breasts, upon the blood elements, and upon blood tension. This secretion is a stimulus likewise to uterine contractions, and is the probable cause of the contractions normally occurring during pregnancy. The action of this accumulated secretion upon the uterus at the end of pregnancy is probably the cause of labor pains.

The action of the ovarian secretion upon pulse tension, and its effect upon the mucous membranes of the body generally, are likewise evidenced by the congestion of the vocal cords during menstruation, so that during this time the singing voice is poor. The secretion of intestinal mucus is also greater, there is increased perspiration, the lower turbinate bones are swollen, and the eye suffers limitations in power. The best evidence of the constitutional elements involved in the process of menstruation is the occurrence of vicarious menstruation. *Under this designation we consider bleedings occurring at regular intervals in a patient suffering from amenorrhea.* The most frequent spot for this bleeding is the nose, usually the lower turbinate bones.

Fleiss described a two-year-old child with well-developed pubes and breasts who had periodical bleedings from the nose. In a second case, a girl fourteen years old, well developed, suffered from regular bleedings from the nose, which stopped, however, when real menstruation began. In another case the regular nose bleedings at intervals of twenty-nine days stopped during pregnancy, only to begin afterward, and after a continuation of eight months ceased again on the occurrence of a second pregnancy. Bleedings of the same kind have been described as occurring regularly from other mucous membranes, the trachea, the lungs, and the stomach. In the latter instance the bleedings were not always associated with vomiting, the blood being usually found in the feces. In other cases there were bleedings into the thyroid gland. In cases with poorly-developed uteri, these bleedings disappeared only when the uterus began to functionate properly.

The secretion of the ovary exerts through the medium of the blood a stimulating effect upon the breasts, noticeable before menstruation, during pregnancy, and during the period of lactation.

Goltz cut through the cord of a dog at the level of the first lumbar vertebra, and later saw the signs of rut appear. After coitus one dead and two living young were born. The breasts were well developed, and lactation and nursing followed the normal course. Since these changes, the sexual tendency and the process of labor could not have been excited through the cord, it must be that a certain secretion of the ovary, *acting through the medium of the circulation*, gives the stimulus for the exercise of those functions.

Ovarian secretion exerts a most decided effect upon the development of the uterus and the genitalia, as well as upon the

breasts. It is absolutely necessary for the preservation of the developed uterus, the other genitalia, and the breasts. An over-production of ovarian secretion or a disturbance in the function of the ovary causes pathological changes; an under-production of ovarian secretion is likewise the cause of pathological conditions. The production of ovarian secretion and its action for a certain period of time are the probable causes of the pains of labor.

C. MENSTRUATION.

Menstruation is the periodical loss of blood from the uterus or from any mucous membrane, occurring for the reason that *no fecundated ovum is present in uterus or tube*, and may be divided into three periods: the *premenstrual*, the *menstrual*, and the *postmenstrual*. During the premenstrual period, the ten days immediately preceding the appearance of blood, the following changes take place:

The superficial capillaries become greatly dilated, and serous infiltration of the endometrium takes place, which separates the meshes of the stroma, accompanied by a gradual but decided dilatation of all the blood vessels and lymph channels. There occur a growth of round cells in the interglandular tissue, and an entrance of leucocytes into the mucous membrane. The glands become larger and wider, being often filled with secretion.

This swelling of the mucous membrane, the dilatation of the blood vessels, the production of round cells, and the growth of the superficial layer of the endometrium produce the so-called *decidua menstrualis*. Although in the connective tissue large cells, not to be distinguished from young stages of decidua cells, are found, it is to be noted that typical decidua cells do not, as a rule, develop at this time in the superficial layer. The endometrium is at this period from 6 to 7 millimetres in thickness.

The period during which blood is thrown out is the next, or *menstrual* period. The superficial capillaries are greatly dilated, and an exit of blood elements, not dependent on a bursting of the capillaries, goes on for several days. The bleeding occurs partly through diapedesis, and, in strong bleedings, through rhesis. There is little or no destruction of the mucosa, only a very slight fatty degeneration of the epithelium of the uppermost layer, so that in the excreted blood relatively few epithelial cells are found.

After menstruation, the uterus shows an almost continuous covering of epithelium, interrupted in spots, especially if patho-

logical processes be present. Most of the ciliated epithelium is preserved. The first stimulus to bleeding is due to contraction of the uterus, which at the height of congestion is possibly accompanied by contractions of the tube. During menstruation the uterus is larger and in the first few days following likewise soft and flabby. The flabbiness lasts longer than the bleeding. A spontaneous dilatation of the cervix canal takes place, and reaches its height on the third or fourth day. This dilatation takes place without regard to the amount of blood discharged, whether the menstruation be painful or painless. The cervix is hyperemic, the glands showing an increased secretion of mucus. The blood thrown off is mixed with the mucus of the uterus and cervix, and later with the acid secretion of the vagina, for this reason coagulating less easily than other blood.

The secretion from the sebaceous glands of the external genitalia is responsible for the peculiar odor present in some cases, and in nowise differs from that observed in the axilla. On the inverted uterus the blood has been observed to appear in single drops from the region of the openings of the glands. The endometrium is then covered with white and bloody mucus, the openings of the glands being clearly seen. There is no degeneration or destruction of the mucous membrane. The changes in the tubes, if any, are slight (Gusserow).

The next, or *postmenstrual* period comprises fourteen days, during which the mucous membrane returns to a thickness of 3 millimetres. During this time all cells not capable of further growth are thrown off, and the epithelium, only partially denuded, is regenerated. The epithelial cells found singly or in groups in the lumen of the glands disappear by resorption, and partially by phagocytosis. From the sixth day after the beginning of menstruation, cell division is prominent. It is probable that mitosis takes place, even during the exudation of blood. The presence of mitosis in the leucocytes is an evidence of growth of the interstitial tissue during menstruation.

Regeneration concerns the *stroma*, the *glands*, and *surface epithelium*, reaching its height on the fourteenth or fifteenth day after the beginning of menstruation. The mucous membrane becomes pale, the glands and vessels have returned to their normal state, the lost epithelium has been restored. From the middle of the third week on there is a diminution in mitosis.

Menstruation is not a process by which the mucous lining of the uterus is thrown off, with subsequent regeneration previous

to the next menstruation. It is simply the excretion of blood from the decidua menstrualis occurring for the simple and sole reason that there is in the uterus no fecundated ovum.

Ovarian secretion is the direct cause of this periodical swelling of the mucosa, and it continues, if fecundation has taken place, to exert, not alone a local, but likewise a stimulating influence on the general and sexual organs. This stimulation occurs to a heightened degree during pregnancy, and on removal of the uterus, since the secretion is ordinarily excreted during menstruation. *On the occurrence of pregnancy, this secretion causes a further development of the uterus and the decidua, and plays an important part in the process of labor; it stimulates the functions of the breasts, exerts a decided constitutional stimulation, and is the cause of many of the pathological conditions occurring before and during labor.*

The generally accepted view as to the relation between ovulation and menstruation is that of Pflüger. The increase of the contents of the Graafian follicle, which results through the secreting activity of the cells of the membrana granulosa, is said to stimulate the nerves running in the stroma and ending in the cells. The increasing tension reflexly irritates the vasomotor centre, with resulting dilatation and congestion of the uterine vessels. At the height of this congestion the follicle is supposed to burst, through pressure. As congestion and tension in the ovary are supposedly the reflex causes of the congestion of the uterus and pelvic organs, menstrual bleeding is considered a result of periodical ovulation. This theory obtained support through the experiments of Strassman, who showed that increased tension in the ovaries of animals, produced by the injection into them of fluid, caused swelling of the mucous membrane of the uterus. In answer it may be asked why the same condition does not result in all cases of ovarian tumors and ovarian cysts, which may develop entirely beneath the albuginea.

This theory of Pflüger and the experiments of Strassman have been generally quoted to prove the causal relation of ovulation to menstruation. The congestion in the ovary, and the swelling of the Graafian follicle, must then, through the nervous system, stimulate the uterus to congestion, with a resulting periodical bleeding. The cessation of menstruation after castration has likewise been cited as proof of the fact that without ovulation there could be no menstruation, since the periodical hyperemia reflexly caused by ovulation no longer takes place. By others it

was considered that the distension of the ovary, occurring periodically, exerted an action upon the vasomotor system, then causing congestion. The atrophy of the uterus after castration was likewise pointed out as a proof of the resulting diminished blood supply, ordinarily furnished by this reflex action of the ovary. On the other hand, it has been frequently stated that the result of castration was due to the tying off of the arteriæ spermaticæ internæ, and that the resulting anemia was the cause of atrophy of the uterus, and of the diminution in the size of the uterine myomata.

Rein cut all the sympathetic and all the spinal nerves running to the uterus in a dog, and yet the animal bore young. Inasmuch as the uterus, freed from the central nervous system, performed its functions as before, he concluded that a regulating nerve centre exists in the ovaries. All this was taken to prove that menstruation was a result of reflex action originating in the ovaries.

D. THE ACTION OF OVARIAN SECRETION ON THE ENDOMETRIUM.

The periodical swelling of the mucous membrane is due to the secretion given off by the ovary, and the experiment of Strassman proves that the forcing of an additional amount of this secretion into the circulation only enhances this effect. That ovulation and menstruation, or at least the latter, occur with a certain regularity is a fact which we are not able to explain any more than we can say why in malaria the plasmodia are thrown into the circulation at regular intervals, or why the menopause usually occurs at a certain period. At any rate, were menstruation the direct result of periodical ovulation, we should still be in need of a satisfactory explanation for the latter phenomenon. No other theory is satisfactory than that of secretion. *Ovulation and menstruation are evidences of the functional capability of the ovary.* Since ovulation may occur without menstruation, but the latter never without the former, we have here an evidence that a certain functional activity of the ovary is necessary to stimulate the mucous membrane to its periodical changes. That after double castration regular bleedings may occur for a certain period is proof of the independent rôle which the endometrium, to a certain degree, plays in the process of menstruation, for sufficient of the secretion of the ovaries may still remain in the circulation to produce the normal processes after removal of the ovaries. The part which the ovaries play in the

development of the body, the effect of their presence upon the breasts and the genital tract at puberty, before each menstrual period, at the menopause, and after castration, are sufficient evidence of their secreting power. The experiments of Knauer and others prove that it is simply the presence of the ovaries, and the preservation of their secretion, which are of importance to the body, and that their action upon the uterus is in nowise reflex in character, since when removed and implanted elsewhere, and in this way loosened from their connection with nerve plexuses and the nervous system, every sexual peculiarity is absolutely preserved.

Menstruation is simply an evidence that a fecundated ovum is not present in the tube or in the uterus. Practically no part of the endometrium is thrown off. It seems as if the blood thus lost simply gave an exit to the secretion of the ovary, which, if retained too long in the body, produces in pregnancy certain normal changes, and in a large number of cases abnormal processes. In pregnancy the ovarian secretion stimulates the uterus to enlargement and growth, it stimulates the formation of blood, produces tension and congestion in the vascular system, and stimulates the function of the breasts. After labor, lactation is stimulated by this secretion, so that little or no effect is exerted upon the uterus; therefore it rarely undergoes the periodical stimulation and has a tendency to atrophy. The changes occurring in the decidua menstrualis and the decidua graviditatis are certainly evidences of the action of a secreted substance. The method in which the ovum is embedded in the decidua, and the processes occurring in its immediate vicinity, as well as the fact that in extrauterine gravidity a decidua graviditatis is formed in the uterus, with decided enlargement of this organ, are likewise proofs of this fact.

E. THE RELATION OF OVULATION AND MENSTRUATION.

As a matter of fact, ovulation and menstruation are related only in that both are the result of the secreting function of the ovary, and are in nowise connected as regards cause and effect. *The ovum is the external secretion, the internal secretion entering the blood through the lymph channels.* The ripening and expulsion of an ovum may occur at any time. It is, however, a fact that *menstruation occurs only after the ovary is capable of producing ripe ova*, and whether the egg be expelled, or occasionally degenerate in the follicle, is immaterial. The inde-

pendence of ovulation and menstruation is evidenced by the fact that the former takes place before menstruation has appeared, and likewise after the menopause, as is proven by cases, by no means rare, of pregnancy in girls who have not yet menstruated, as well as by the occurrence of pregnancy at variable periods after the amenorrhea of the climacterium. In children the primary follicles develop fully before puberty, and the same development occurs even in the fetus and in the newly born. Those ova and follicles go through the same stages of development and ripening as in the case of adults. That they are not capable of fecundation is shown by the fact that the ova are only one-half as large as in adults. In those cases, however, of young children with well-developed breasts and genitalia, when menstruation begins there is an unusually strong development of the body, and the ova, as well as the follicles, differ in no way from those found in menstruating adults.

Leopold found that in forty-two pairs of ovaries, thirty pairs showed a corpus luteum belonging to the last menstruation. In thirteen pairs no follicles were found which, on account of their size or the swelling of the follicle, justified a belief that a bursting of any follicle would have occurred at the next menstrual period. In twelve cases no corpus luteum was found belonging to the last menstruation. One case showed a follicle which had burst between menstrual epochs.

Arnold found that in fifty-four cases, only thirty-nine showed the presence of fresh corpora lutea after the last menstrual period.

Williams found this to be the case in twelve cases out of sixteen. Therefore, ovulation, although in the majority of cases occurring at or near the menstrual period, is not the cause of the same.

Leopold has shown experimentally that in the inter-menstrual period follicles ready to burst are present, and that through certain causes, such as coitus, the exit of an ovulum may result at any time. That ovulation occurs during pregnancy is proved in the case mentioned by Kroenig, in which conception occurred as a result of coitus four days post partum. The relatively frequent occurrence of pregnancy during the temporary amenorrhea of lactation is a proof of ovulation during this time.

Consentino finds that during pregnancy and lactation ovulation continues. Although the ovum of a menstrual period may be fecundated immediately thereafter, the ovum usually fecundated

is the one given off between four and eight days before menstruation, the egg thus belonging to the period, so to say, first omitted. The ovum retains its vitality for an average period of twelve days, and yet out of a collection of two hundred and fourteen pregnant cases, in sixty-five coitus had taken place after the twelfth day following menstruation. Therefore, in these pregnancies at least, the ovum must have belonged to what may be called the next awaited menstrual period. Even if the expulsion of the ovum occur only a few days before that time, its fecundation is not difficult to explain, nor are the cases puzzling in which one coitus directly after menstruation produces pregnancy, as the spermatozoa retain their vitality for a long time, and have been preserved, in proper temperature, for a period of eight days. *Dührssen has found spermatozoa in the tube three and one-half weeks after the last coitus.*

It is therefore evident that ovulation, as a rule, occurs from four to eight days before menstruation, but it may occur at other periods, as ripe ova, practically speaking, may be present at almost any time.

F. OVULATION.

The other of the functions of the normal ovary is the production and expulsion of ova capable of being fecundated. It is probable that after birth no new ova are formed from the germinal epithelium. At and after puberty we judge the vitality of the ovary by its ability to bring these ova to a stage which may be called ripe. For the expulsion of an ovum from the Graafian follicle, a gradual increase in size of the follicles takes place, depending partly on an increase in the amount of liquor folliculi. The cells of the follicle epithelium undergo fatty degeneration, and the internal layer, the tunica interna, shows an increase in the size of the cells, and a decided development of the blood vessels. The protoplasm enlarges greatly, begins to take on a yellow color, and these cells, now lutein cells, are arranged in several layers, forming an irregular surface. As a result of the fatty degeneration of the follicle epithelium, the ovum is freed from the cumulus oophorus. The most prominent point of the follicle is poor in blood supply, and furnishes the so-called *stigma folliculi*. It is here that the opening takes place which serves as an outlet for the ovum. This opening is probably the result of the reaction or chemical effect produced by the ripe ovum, since in the newly born, and in children, follicles of the same size and even

larger ones exist without bursting—the so-called atresic follicles. The facts that large follicle cysts occur in the ovary without opening, and also that in women in whom the unopened follicles degenerate, disturbances of menstruation occur—so-called missed ovulation—speak for a chemical reaction as probably one of the functions of the ripe ovum. That the orgasmus venereus does play a part in hastening the expulsion of the ovum is not to be questioned. In most cases the opened follicle is then filled with blood which likewise empties into the peritoneal cavity, and the so-called corpus luteum spurium results. If pregnaney takes place, this body develops deecidedly, forming the corpus luteum verum (Nagel).

After ovulation, the ovum is thrown out into the abdominal cavity, and then, influenced by the wave movement of the ciliated epithelium of the tubes, the fimbriæ of the ampulla, and the fimbriæ ovaricæ, finds its way into the uterus. It is not necessary that the tube should grasp or surround the ovary, since it would embrace, even under favorable circumstances, only part of the ovary. *This wave movement of the ciliated epithelium causes a current in the peritoneal plasma, which directs the ovum into one or the other of the tubes.*

Lode injected, with a needle pointed toward the diaphragm, the eggs of ascarides into the abdominal cavity of rabbits, in the region of the umbilicus. After ten hours, without the excitement of coitus, he found these eggs in the tube. An interesting proof of the existence of this current is given by Knauer, who removed the ovaries of dogs, and sewed them into the mesentery. In one case, on opening the abdomen he saw in one of the ovaries three ripe Graafian follicles. Shortly after, coitus took place, and in due time two young were born. A further proof is found in the experiments of Leopold, who showed that an ovum given off by one ovary may enter the tube of the other side. The cases are not rare in which the tube of one side was closed or absent, and although the corpus luteum verum was found in the ovary of the same side, yet the ovum was found in the uterus. The same is true of those cases in which the corpus luteum verum is on one side, and the ovum develops in the other tube, or in the horn of a uterus unicornis of the opposite side. Therefore it is certain that, normally, a wave current exists on either side of the uterus, and that the ovum is attracted by the stronger current, usually that of the side from which it came, possibly, however, by the current of the other side.

It is therefore evident that an ovum, by natural means, may at any time enter the tubes, and that no congestion of the tube, no erection of the tube, and no reflex spasmodic grasping of the ovary by the tube are necessary.

Lode has shown that an ovum finds its way through the tube into the uterus in thirty hours.

CHAPTER II.

THE EMBEDDING OF THE OVUM IN THE GUINEA-PIG.

A. UTERUS OF THE GUINEA-PIG.

The uterus of the guinea-pig is six centimetres long and consists of two muscular layers, an external longitudinal one closely connected with the peritoneum, and an internal circular one. Sometimes, within the latter, are found isolated bands of longitudinal fibres.

Between the two muscular layers is a broad zone of loose con-

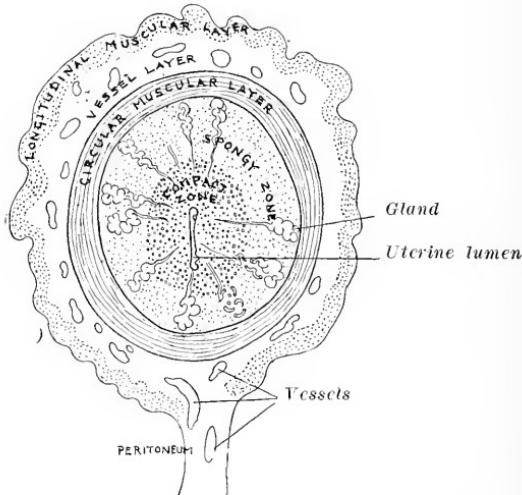


FIG. 1.—Half-schematic section of the uterine horn of a guinea-pig. (v. Spee.)

nective tissue, and in this are found the continuations of the vessels which enter through the longitudinal layers at the mesometral area. All the larger vessels are in the circular layer. Their walls lose the muscularis and their continuations then enter the specific parenchyma of the uterus, where they appear as *capillaries or simple endothelial channels*.

The uterine lumen is lined with a single uninterrupted layer of cylindrical epithelial cells; no cilia are clearly seen. This epithelial layer descends into the connective tissue at numerous places, into glands lined with a low epithelium. The peripheral

ends of the glands are twisted and extend up to the circular layer. An active increase of epithelial cells through mitosis takes place *only in the end areas of the glands*, so that here the regenerative stations for the uterine epithelium are to be sought. In other areas of the uterine lining the epithelial cells have lost the power to produce new cells. *This is the case in animals in whom the epithelium plays no part in the formation of the placenta.*

The connective tissue consists of two zones separated by a transition zone. (1) A peripheral zone surrounding the twisted gland ends and consisting of spindle or branching cells with intervening spaces, resembling a loose reticular connective tissue. (2) A central compact zone situated between the straight excretory ducts of the glands. *In the implantation of the ovum only this area is invaded.* Extremely numerous mitoses, and the



FIG. 2.—Cells of the compact zone with the epithelial lining of the uterine lumen of the guinea-pig. (v. Spee.)

varying appearance and thickness of their tissue, are proofs of active changes and an increased tissue change. Since at points not important the strongest cell increase may be found, the conclusion is justified that the life of the connective-tissue cells of the uterus is short. There are no spindle-shaped or branching cells here as are found in the peripheral zone.

The cells of the central zone (Fig. 2) lie like the cells of stratified epithelium. They are polygonal, close together, generally separated by sharp contours, which are probably intercellular spaces. The most firmly uniting basis is furnished by the capillary network, for a real connecting factor among the elements of the connective tissue, especially of the compacta, is not present. The normal relation is preserved by the firmly united epithelium and the circular muscular layer, between which lie the two layers of connective tissue.

B. THE EMBEDDING OF THE GUINEA-PIG'S OVUM.

In 1883 Spee stated that on the sixth day post coitus the ovum of the guinea-pig is an oval germinal vesicle one-tenth of a millimetre in diameter, surrounded by a zona pellucida and lying free in the uterine cavity (Fig. 3). The cell wall of the vesicle, the germinal layer, consists, in the region of the ovum's equator, of a single layer of very flat cells. At the two poles the wall seems thicker. At one pole the cell wall of the vesicle is stratified and forms a prominence into the cavity of the vesicle, the Germinal Prominence of Hensen or the Placental Pole (P. P.). At the other pole (the Opposite Pole of Spee) the germinal layer consists of a single layer of cubical cells, constituting the Implantation Pole (I. P.). It forms after implantation that summit of the ovum into which the embryonal sphere enters and in which the embryo develops.

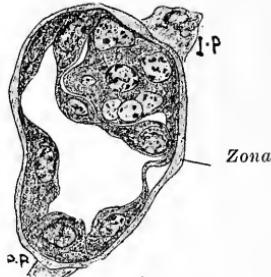


FIG. 3.—Ovum of guinea-pig free in the uterine cavity. Ovum with zona.
Note.—The lettering of Fig. 3 is reversed. (v. Spee.)

The zona is preserved up to the beginning of the seventh day post coitus and can be readily seen on the free germinal vesicle. The cubical cells of the Implantation Pole send prolongations of their cell bodies through the zona and can enter into direct relation with the epithelial lining of the uterus before the zona is lost finally to the ovum, *and may so furnish the first connection which leads to implantation*. These prolongations have been seen only in the region of the Implantation Pole. The short period, during which the prolongations of the Implantation Pole hold the ovum fixed to the uterine epithelium, however, has not been observed.

When the ovum is first attached to the uterine epithelium the zona is present (Fig. 3a), but disappears in a very short time, for it is not seen in embedding ova. Probably the Implantation Pole prolongations perforate it and cause its disappearance at these

points. The rest is probably rubbed off. In irrigating a uterus for ova, Spee once found the cell body of an ovum and an empty zona with a hole large enough to have permitted the former to slip out. Acids dissolve the zona, and, as these were often used in fixing the specimens, the fact that no zona was found in the youngest ova which had entered the uterine wall does not prove that the ova were embedded after removal of the zona. The ova during implantation, however, are so closely surrounded by uterine tissue that between them there is no room for a zona, and so in all probability the ovum does not take the zona with it into the uterine wall.

The Embedding or Implantation of an Ovum includes the group of processes which result in the ovum entering from the

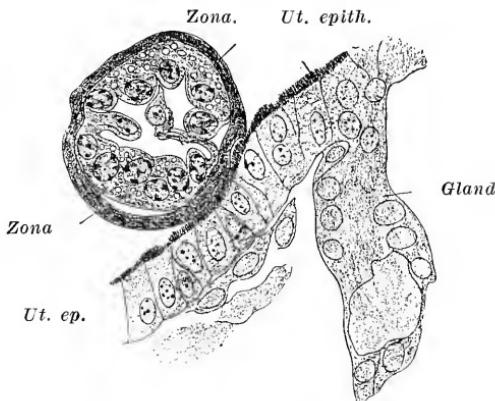


FIG. 3a.—Ovum of guinea-pig in its first adhesion to the uterine epithelium. Ovum still possesses its zona. (v. Spee.)

uterine cavity into the connective tissue of the uterine wall. The processes antedate the formation of the placenta and are distinct from the processes of placental formation, but the first evidences of the latter follow immediately upon implantation. In the guinea-pig, ova are never implanted at a point at which or in whose neighborhood signs of circulatory disturbances (excessive collection of tissue exudation in the interstices of the subepithelial connective tissue, or a grouping of red blood cells in these interstices), or evidences of a throwing off of tissue into the uterine lumen, are noted. Only such areas are selected as are normal and whose tension, dependent on tissue sap, is even, which fact coincides with the observation that the uterine lumen at the selected point always shows a surprisingly smooth epithelial lining without folds.

The ovum is almost always embedded on the anti-mesometral side, corresponding to the fundus uteri in the human being, and beings about 6 days, 8 to 12 hours post coitus, seldom later at the end of the seventh day. The time left for embedding is 4 to 8 hours. Its beginning is known with a possible variation of about 6 hours. The ovum does not increase in size before its embedding, and measures with the zona only 0.1 millimetre, without the zona 0.08 millimetre, while the uterus is 6 centimetres long. As often only one or two ova are present and as their location is not macroscopically evident, only series sections divulge the various points. If the ovum is already through the uterine epithelium, it is seen with difficulty because of its small size. One may readily imagine the labor involved in obtaining the present un-

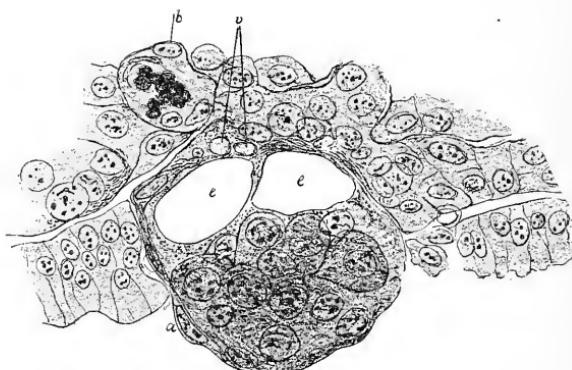


FIG. 4.—*b*, capillary; *v*, vacuole; *c*, cavity in ovum. Ovum of guinea-pig partially embedded, showing disappearance of the uterine epithelium. (v. Spee.)

broken series illustrative of the various stages, a work on which Spee has been engaged for the last ten years.

The cells of that pole which leads all other parts in its entrance into the uterus (I. P.) send prolongations through the zona shortly before the implantation period, and render contact and exchange with the uterine epithelial cells possible, even before the zona is gone. These prolongations cause the first adhesion of the ovum to the uterine epithelium (Fig. 3a).

The uterus plays a passive rôle in implantation of the ovum. The ovum enters the uterine wall and a correspondingly large space of uterine tissue disappears.

The uterine epithelium shows no sign of growth.

The uterine epithelium of just that area in contact with the ovum disappears (Fig. 4).

In Fig. 4 the ovum is 6 days and 10 hours post coitus, and evidences no zona. A thin cell layer (*a*) surrounds for a distance the cavity (*e*) of the ovum on one side, but is not distinct on the other side. The solid portion of the ovum consists of large round cells which are not surrounded by a cell covering. Only at *a* is there a flat cell which stands in continuity with the cell membrane enclosing the cavity of the ovum (*e*). The membranous portion of the ovum contains two small vacuoles (*v*).

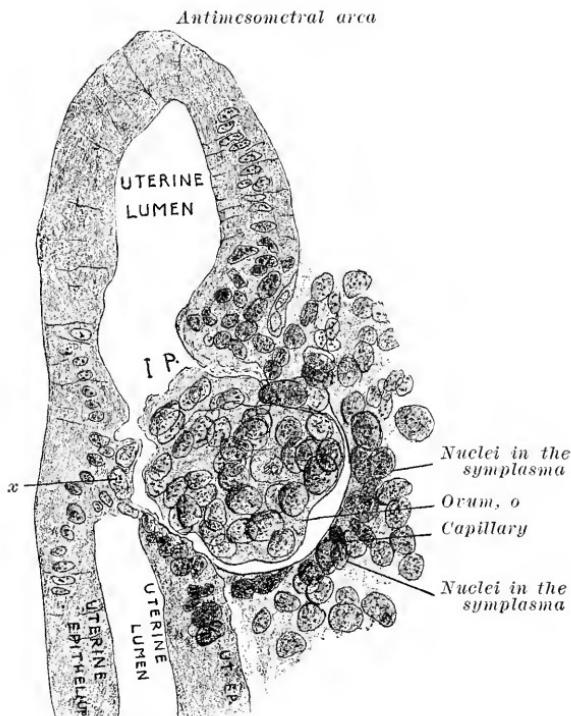


FIG. 5.—Further stage in embedding of ovum, showing changes wrought in the connective tissue compacta. (v. Spee.) *x*, two cells of the ovum which are embedded on the opposite side.

The central cavity is lined with a non-celled coagulation substance, which has divided the cavity into two parts, probably a postmortem change. The external surface of the solid area is not covered by a specially differentiated layer of flat cells. The solid group of cells represents the placental pole. The cavity *e* represents the germinal cavity. The part of the ovum near the connective tissue, at the Implantation Pole, is the cell wall of the cavity, the germinal membrane. The space between the cells,

and the thin layer near the connective tissue, later becomes filled with cells, forming a solid ovum.

Decided changes occur in the connective tissue which makes room for the ovum. These changes for a long time are such as may be considered a necrosis and a paralysis of the life processes of the connective-tissue cells (Fig. 5).

Fig. 5 shows a double embedding at two points vis-à-vis. The ovum evidences no cavity. The ovum is already deep in the wall and the uterine epithelium is interrupted in the circumference of the ovum. The surrounding connective-tissue cells have changed

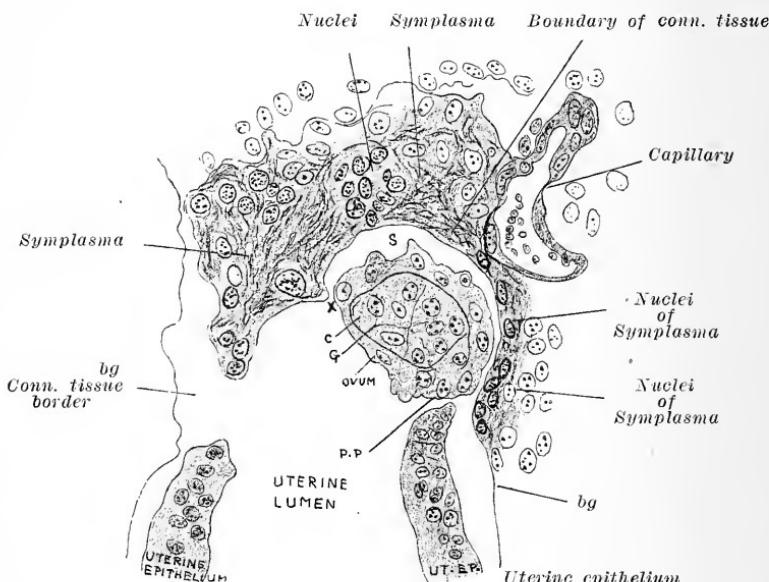


FIG. 6.—Further stage of embedding of guinea-pig's ovum, showing the connective tissue compacta about the ovum destroyed by the bio-chemical influence of the ovum. (v. Spee.) c. centre of ovum; G. embryonal sphere.

in that a half-moon arranged row of connective-tissue cells of a stronger stain has been formed. The cells of the ovum are much larger than the connective-tissue cells. To the left at *x* two cells of the ovum are united to the uterine epithelium, and two others, not distinctly seen, have perforated the uterine epithelium. The single layer about the ovum at *I. P.* is the germinal layer plus the implantation pole. The cell group at the placental pole has grown into, and filled out, the germinal vesicle, but the ovum is not enlarged. Therefore the ovum now evidences no cavity. The sharp contour at *o* bounds the connective

tissue sharply and is independent of the cell body of the ovum. That the cells of the ovum dissolve the epithelium with which they come in contact, is seen in its early stages at *x*, where four cells are present. The connective-tissue cells about the ovum are large and polygonal with sharp contours and of epithelioid form. This form also is found in certain areas away from the ovum, and is there dependent on other influences than the ovum. This change in the cells extends, and this area is called the Implantation Area. During this peripheral extension the cells nearest to the ovum show other changes in the nuclei. They stain darker. This is due to a process of dissolution, and as a matter of fact

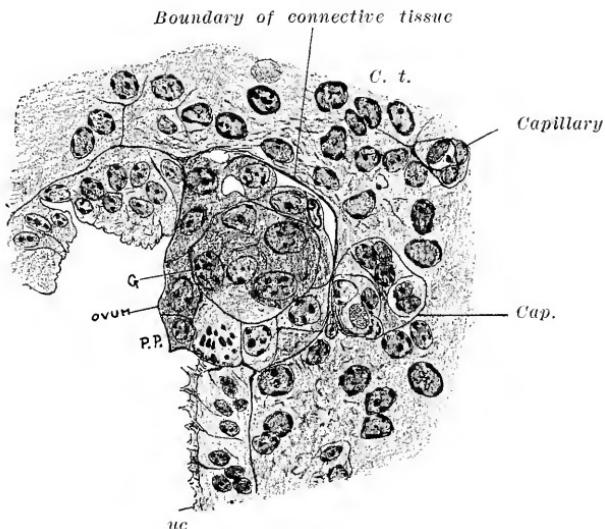


FIG. 7.—A stage of embedding one-half hour later than Fig. 6 (v. Spee), more highly magnified. *G*, embryonal sphere. The ovum is surrounded by a symplasmatic zone.

the nuclei nearest to the ovum are already smaller and mitosis has ceased in the immediate circumference of the ovum.

The connective-tissue cells are fluidified by a form of digestion. The disappearance of mitosis for a considerable distance about the ovum, the dissolving of the connective-tissue cells immediately about the ovum, while at a distance normal cell division takes place, show that the destruction of uterine tissue is due to a biochemical process dependent on the ovum (Fig. 6).

Fig. 6 is still further embedded, and in four-fifths of its circumference is a single layer of cells, the germinal layer. This is connected at the placental pole with a solid mass, the germinal

prominence, which fills the space surrounded by the germinal layer. The Implantation Pole is situated on either side of *S*.

In Fig. 7 the ovum is situated half in the epithelium and half in the subepithelial connective tissue. The ovum is sharply bounded from the connective tissue. It is surrounded by a *symplesmotic* zone in which a histolytic process causes the contours and nuclei of the connective-tissue cells to disappear, as if the ovum were a poison. The ovum consists of a covering layer and a contained mass (*G*) of large cells containing fat. A small

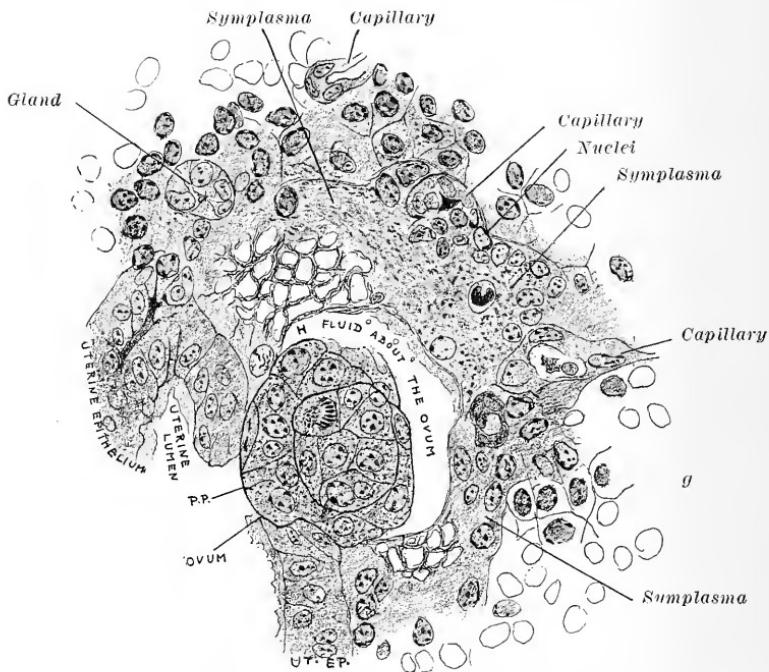


FIG. 8.—Almost embedded ovum surrounded by fluid resulting from fluidified connective tissue cells. (v. Spee.)

space separates the ovum, in part, from the connective tissue (*C. t.*).

The ova 6, 7, and 8, though differing only about half an hour in age, produce great changes in the connective tissue. Ovum No. 6 is about three-fourths embedded. Ova 7 and 8 were found in the same uterus. Only in 8 is a growth of the ovum, as compared with 5, evident. Important are the connective-tissue changes, first in the cells of the Implantation Area, and, second, in the boundary line which faces the ovum. Ovum No. 8 is al-

most embedded and several of its cells are on a line with the uterine epithelium. A comparison of ova 6 and 8 evidences two differences, which show the two varying functions in two different parts of the ovum, (1) the part consisting of the external layer, a derivative of the germinal layer, and the Implantation Pole; (2) the cell mass at the Placental Pole. In the circumference of the Implantation Pole the connective-tissue cells beneath the epithelium disappear or degenerate. The cells at the Placental Pole do not destroy the uterine tissue, therefore the hole in the

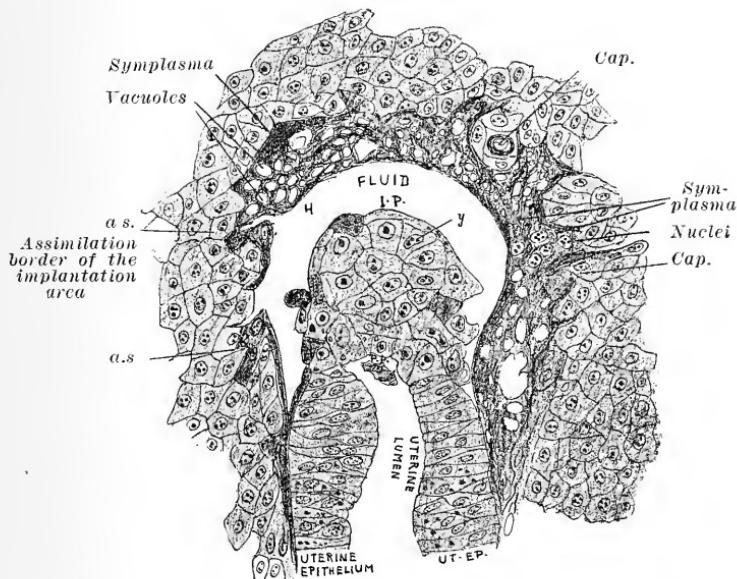


FIG. 9.—Ovum entirely under the uterine epithelium. *a.s.*, assimilation border of the implantation area; *y*, embryonal sphere; *H*, fluid space about ovum. (v. Spee.)

uterine epithelium at the Placental Pole of ovum 8 is not enlarged.

In the Implantation Area the connective-tissue cells and the nuclei become large. The chromatin of the nuclei becomes grouped on the inner surface of the nuclear membrane, but the nuclear centre is pale. Peripheral to the implantation zone numerous mitoses are observed and are also found in the vessel endothelia. A dense tissue results and the cell interstices disappear, as may be seen in Fig. 9. This very peripheral area has no meaning with regard to implantation or to placental formation.

In Fig. 5 the cells nearest the ovum stain darker and are smaller. In Fig. 6 a contour divides the connective tissue near

the ovum from the ovum, but instead of numerous cells and nuclei we see a fibred, granular mass, a group of closely gathered nuclei without cell contours. In Fig. 7 some nuclei are still well stained, but lie in a fibred, granular mass in which no cell contours are evident. *A symplasma has been formed through the dissolution of the connective-tissue cells of the implantation zone which is sharply marked off from the ovum.*

The ovum is then almost surrounded by fluid resulting from these changed cells (Figs. 8, 9, 10).

In Fig. 8 the symplasma is separated from the ovum by the

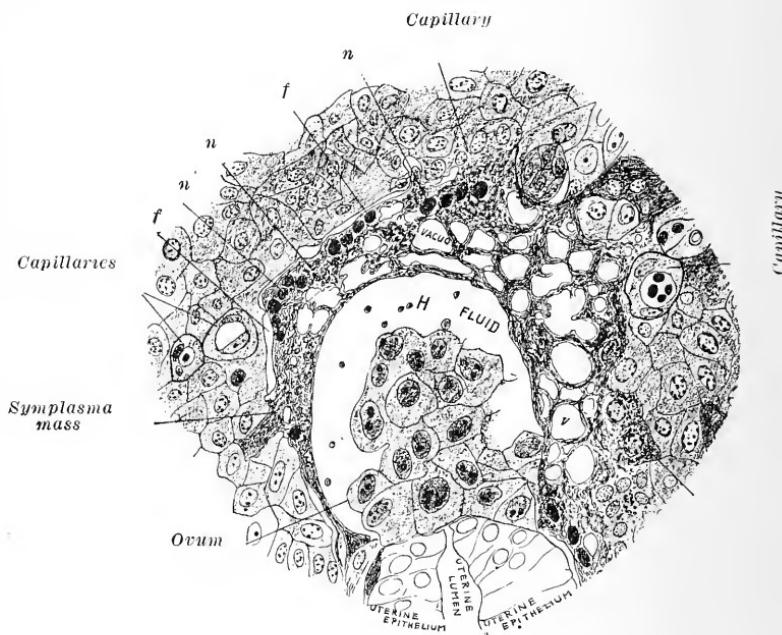


FIG. 10.—A further stage, showing demarcation of the symplasma, thus limiting the assimilation of future areas. *n*, free nuclei in the symplasma. (v Spee.)

space *H*, which is wider than in Fig. 7. In the periphery are smaller, darker cells ready to join the symplasma. Here, then, is the *assimilation boundary* of the symplasma. The space *H* is filled with a fluid into which the edge of the symplasma passes over gradually. This constitutes the dissolution boundary of the symplasma. The contents of the space *H* are a thin emulsion which is under pressure, causing the circular contour of the symplasma in Fig. 9.

The fluidifying of the symplasma occurs not only at its edge but in its substance, and becomes porous and filled with vacuoles which contain fluid (Figs. 8 and 9). In Fig. 9 the vacuoles are present in large number. At the assimilation boundary at *a. s.* are seen darker, smaller cells. At *f* (Fig. 10) is seen a sharp demarkation of the symplasma of the implantation zone which limits the assimilation of future areas. The nuclei of the last assimilated cells lie at this area of separation, in short simple rows. They furnish almost the same picture as the nuclei in the syncytial or plasmoidal formations in the placenta of other animals. The symplasma degenerates and the original boundary of *H* formed by it disappears.

Sympl.

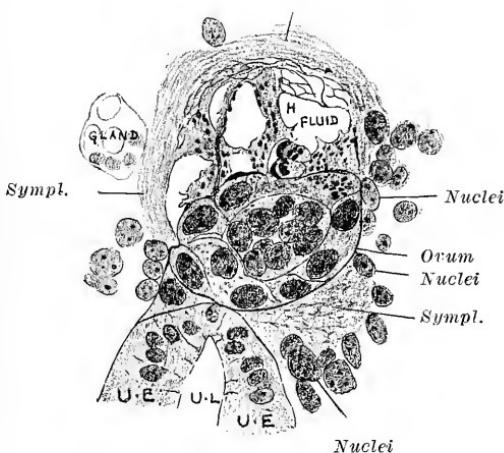


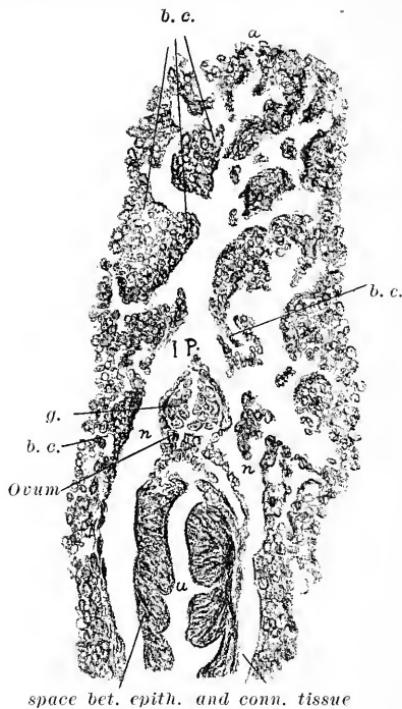
FIG. 11.—Ovum entirely embedded in connective tissue compacta, showing spaces (*H*) in the fibred tissue about the ovum. The fibred tissue about the ovum contains numerous granules. This is a middle stage between the normal compacta and the final fluid symplasma. (v. Spee.)

Along the epithelial lining the subepithelial cells become smaller, and darker areas of separation appear between them and the epithelium leading up to complete loosening of the latter. The subepithelial cells, fluidifying, form lacunæ in the subepithelial connective tissue (Fig. 12).

This fluid, through diffusion or possible absorption by the ovum, makes place for the ovum, which only now begins to grow quickly. Later, at the boundary between necrotic and healthy tissue, a real granulation tissue is formed as an active reaction to the entering ovum (Fig. 12).

In the transition zone appear granular cells, which increase

from Fig. 9 on, and eventually forming a characteristic zone in the far periphery, in which the cells also seem clouded. Soon a separation of the implantation area from the connective tissue occurs at the transition zone through great growth of cells in the peripheral areas, and through a stoppage of growth and degeneration in the implantation area. The capillaries pass through this boundary and on their walls are grouped connective-tissue cells. They are numerous at the point of separation, so that the



space bet. epith. and conn. tissue

FIG. 12.—*g.*, embryonal sphere; *n.*, free nuclei of the symplasma; *b.c.*, capillaries. The ovum has grown. Ovum has a cavity, except at implantation pole, where the embryonal sphere is in contact with the external layer of the ovum. (v. Spee.) Completely embedded, growing ovum.

wall looks like a granulating wound surface of endothelial channels. The space about the ovum is gradually filled with granulation tissue in the next twelve hours. Seven days post coitus the ovum has grown considerably (Fig. 12). The Placental Pole evidences a papillation of the germinal layer which gives this end a half-cylindrical groove, due to two lateral folds. The ovum now evidences a cavity, except at the Implantation Pole, where the embryonal sphere is in close contact with the external membrane of the ovum.

Seven days thirteen and one-half hours post coitus the ovum has grown large (Fig. 13). The space between the embryonal sphere and the Placental Pole has grown decidedly, so that the ovum is a long, cylindrical vesicle, with its Placental Pole in contact with the uterine epithelium.

The investigations of Spee show that the ovum of the guinea-pig, not grown since its fecundation, destroys like an injurious parasite the uterine epithelium and the subepithelial connective tissue to make place for itself in the uterine wall in which it is embedded, and that only after this does it enter into a symbiotic connection with the uterus, which connection finds its expression in the formation of a placenta. The same occurs with the human ovum.

CHAPTER III.

THE EMBEDDING OF THE HUMAN OVUM.

A. THE UTERUS.

For many years it had been taught that the human ovum in the uterus developed on the mucosa (decidua serotina) and was enveloped by mucosa growing up about it (decidua reflexa).

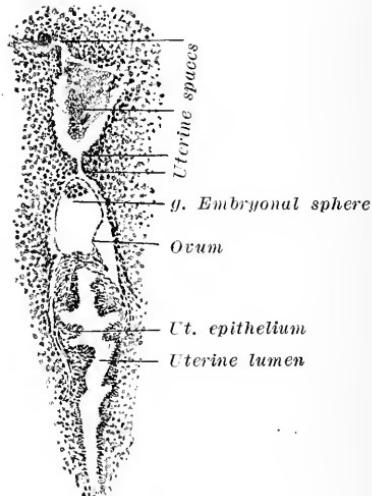


FIG. 13.—Cavity in ovum between embryonal sphere and placental pole has grown greatly. *g*, embryonal sphere. (v. Spee.) Rapidly growing, embedded guinea-pig ovum.

In the examination of young human ova Spee was never able to find uterine epithelium lining the bed in which the ovum rested, and he doubted the theory that the uterine mucosa and its epithelium grew up and surrounded the ovum. Hensen had made the observation that the ovum of the guinea-pig, on the seventh day post coitus, was no longer to be found free within the uterus, and that on the eighth day it was located in the uterine wall *underneath the epithelial lining* (Fig. 13). Selenka, on the other hand, supported the view that the ovum of the guinea-pig entered into a uterine gland whose epithelium was later destroyed.

The relation between ovum and uterus varies in different animals. There are forms in which the uterine epithelium is preserved at the point at which the ovum is attached. In all those cases in which the ovum *grows before its attachment*, it remains during its entire development within the lumen of the uterus or in a portion of the same. The closer relation between ovum and uterine wall develops later. In the case of ova *which do not grow at that stage*, the epithelium disappears. The smaller the ovum, and the smaller the area of uterine wall necessary for its attachment, the more intensely is the uterine tissue affected by the ovum, so that the area affected at this point becomes rapidly very large in proportion to the size of the ovum, and naturally the development of close connection between ovum and uterus occurs earlier than in the first class. Early attachment and small ovum are therefore favorable to the nourishment of the latter. A most favorable feature for the ovum is the form in which the fecundated germinal vesicle, *without any increase in growth*, is completely embedded in the subepithelial connective tissue of the uterus. *Such is the case with ova of the rat, mouse, guinea-pig, and human being.*

Decidua Menstrualis.—In the menstrual decidua the superficial capillaries become greatly dilated, and serous infiltration of the endometrium takes place, which separates the meshes of the stroma, accompanied by a gradual but decided dilatation of all the blood vessels and lymph channels. There occur a growth of round cells in the inter glandular tissue and an entrance of leucocytes into the mucous membrane. The glands become larger and wider, being often filled with secretion (Fig. 14).

This swelling of the mucous membrane, the dilatation of the blood vessels, the production of round cells, and the growth of the superficial layer of the endometrium produce the so-called decidua menstrualis. Although in the connective tissue large cells, not to be distinguished from young stages of decidua cells, are found, it is to be noted that typical decidua cells do not, as a rule, develop at this time in the superficial layer. These round cells are the beginnings of the decidua cells. The endometrium is at this period from 6 to 7 millimetres in thickness.

Decidua Graviditatis in the First Week.—On the occurrence of pregnancy, as will be noted below, the superficial portion of the mucosa is later composed mainly of cells and is called the *compacta*. The deeper layer is composed mainly of glands and is called *spongiosa*. In the first week, however, as may be

seen from the description of Peters, there is a decided division into compacta and spongiosa only near the ovum. There is at this time no real decidua elsewhere and no difference exists between the connective-tissue cells of the superficial and deep layers. The tissue between the glands, however, is thicker

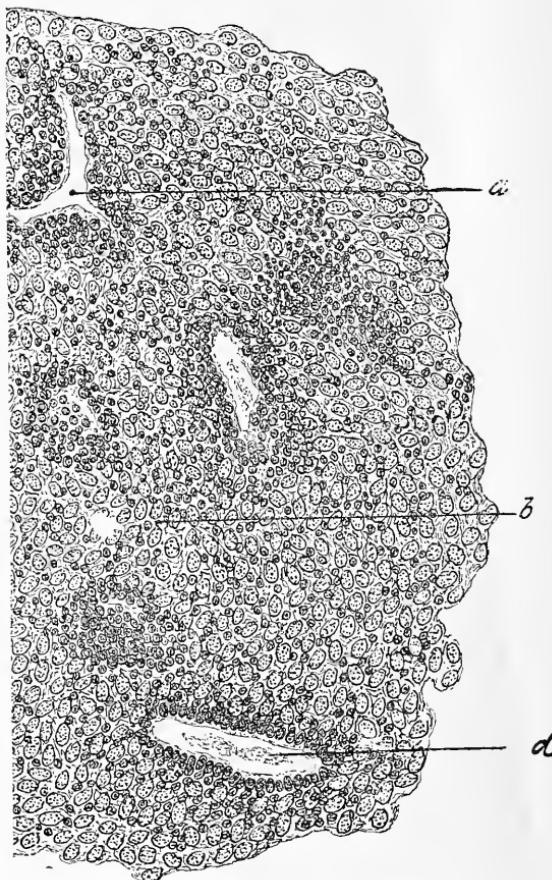


FIG. 14.—Menstrual decidua. *a*, section through vessels surrounded by groups of round cells; *b*, interglandular tissue consisting of normal cells with scattered round cells; *d*, section of a gland somewhat dilated—its epithelia are somewhat smaller than normal. (Abel.)

toward the surface. We find spindle-shaped connective-tissue cells mainly. The spaces between the cells are filled with a pale homogeneous plasma, and the tissue looks like reticular embryonal connective tissue. The nuclei in the more superficial cells are somewhat larger, and numerous small round cells are pres-

ent which represent the early stages of decidua cells. The epithelium of the superficial glands and of the uterine surface is somewhat flattened. An hypertrophy of the glands, especially in the deeper layers, is present, and in transverse section they are lined with *papillary projections on which are long,*

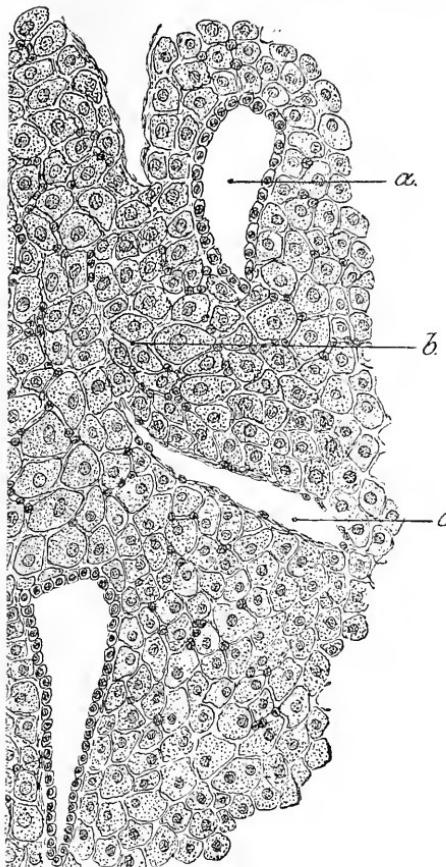


FIG. 14a.—Decidua in intrauterine pregnancy (abortion) at the second month (curetting). *a*, section of a gland with flattened epithelia; *b*, interstitial tissue consisting of the so-called decidua cells, between which at certain points irregularly-scattered round cells are seen; *c*, section of a vessel—in the wall are endothelia. (Abel.)

high, cylindrical cells, often looking like beaker cells. Numerous epithelia are found free in the glands. These glandular changes are marked mainly near the ovum.

Decidua Graviditatis.—These typical changes occur, chronologically, at a later period in the entire uterus. Since, however,

they are marked in the first two weeks *immediately about the ovum*, they are mentioned now. It is especially necessary to consider the base in which the ovum develops, as it is difficult at all times, even as early as the first week, to distinguish in many areas between fetal and maternal cells. The mucosa of the pregnant uterus, at its thickest, increases to ten times its original depth by the growth of all its elements. At the end of the first month it is one centimetre thick. Its surface is uneven, deep grooves dividing into irregular fields. These changes are greatest at the fundus. In the region of the internal os the changes are less marked, the lining of the cervix, except for a high degree of hyperemia, having no part in these changes. The increase in the volume of the decidua depends greatly upon the growth of round cells in the endometrium and upon their change into decidua cells. The round cells of the connective tissue, sit-



FIG. 14b.—A gland in the decidua graviditatis, showing the erroneously named "syncytial change" of the lining epithelium. Taken from the curetted mucosa of a uterus two weeks gravid.

uated in a meshwork of spindle- and star-shaped cells forming the stroma, enlarge. The protoplasm increases greatly in amount and the nucleus becomes larger. *The cells are closely grouped, pale and epithelial in appearance* (Fig. 14a). They are, like the connective-tissue cells of the normal uterus, infiltrated by capillaries and larger vessels whose endothelium borders immediately on the individual cells. The nuclei are round and long, 8 to 10 μ long and 10 to 12 μ wide. They stain lightly because their chromatin network is very fine. There are nucleoli. Mitosis is rarely observed, but two nuclei may be present. They generally retain their spindle-shaped form or are round polygonal. There is often a fine space between the individual cells, filled with a homogeneous intercellular substance. These cells appear at first in the upper layer, and so prominently that this

layer of the gravid decidua is called the compacta. Later, as a result of pressure, the cells resemble squamous epithelium.

The external layer near the muscle contains so many gland lumina that the interglandular tissue is in the background. This interglandular tissue disappears more and more, so that finally a tissue honeycombed by glands results. This layer, in contrast to the superficial, is known as the spongiosa. The compacta is thrown off with the ovum; the spongiosa remains to regenerate the glands.

The gland cells lose their cylindrical form and become cubical and spherical. The protoplasm grows and the nuclei are therefore further apart. Into the gland extend prominences covered with groups of epithelium (Fig. 14b). Sometimes the cell boundaries become indistinct, giving the so-called "syneytial change," which has nothing to do, however, with real syncytium.

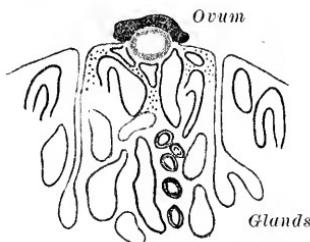


FIG. 15.—Schematic representation of the earliest stage in the embedding of the human ovum. (Peters.)

The epithelium of the surface is subjected to pressure and is flattened, becoming later thin as endothelium.

B. THE EMBEDDING OF THE HUMAN OVUM.

Spee and von Herff have shown that in the guinea-pig the ovum forms an opening through the epithelial lining of the uterus, and that, after having passed entirely through the mucous membrane, it develops further in the region of the subepithelial connective tissue. The opening thus formed is closed by lymph exudation, and there results about the ovum a so-called lymph space, showing that an exchange between the ovum and the surrounding tissue exerts upon the latter a certain reaction. That the same process of *centrifugal descent* of the ovum into its bed occurs in the human being is to be expected, for the following reasons: 1. By analogy with the above process. 2. In human beings there is no epithelium, nor

are there any gland openings on the surface of the so-called reflexa, facing the ovum. 3. The youngest human ova found in the uterus show no prominence into the uterine cavity, which would be the case if they developed upon the decidua vera and were surrounded by a decidua reflexa.

Ovum in the Earliest Stages.—Peters examined a growing ovum supposedly three days old. The uterus was slightly enlarged and soft. The decidua at the fundus and on the posterior wall was 8 millimetres thick; on the anterior, 5 millimetres. The lining of the cervix was hyperemic. On the posterior wall was a slight prominence, which proved to be the ovum, whose largest diameter was found to be 1.6 millimetres. The ovum was embedded in the compact part of the decidua, which portion, as

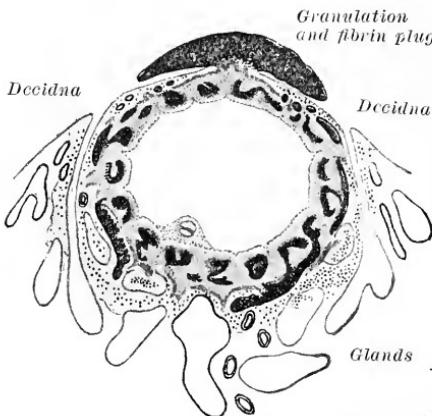


FIG. 16.—Schematic representation of a centrifugally embedded human ovum. (Peters.) The summit of the capsularis is closed by a fibrin plug.

before stated, showed only a slight prominence. The summit of the ovum was *not covered by a capsule*, but showed a spot of blood granulation.

Only in the immediate circumference of the ovum was there a clear division into compacta and spongiosa. The nearer to the ovum, the more frequently were free red and white blood cells found between the cells of the stroma and in the plasma. Nowhere was there an evidence of fatty changes in the superficial layers. The mucosa was everywhere extremely vascular. The venous channels showed, superficially, often only a simple canal of thin endothelium, without adventitia. The arteries preserved their adventitia up to the covering epithelium. The nearer to the ovum, the greater was the vascularization. In this case fecundation took place shortly before the expected men-

strual period. The fact that this ovum had made its way into the decidua, wherein it was firmly embedded, proves, analogous to the observation of Spee, that *an ovum can develop only on a spot free of epithelium*, and that, through reaction upon the surrounding tissue, an ovum sinks into the decidua, and that this reaction causes a dilatation of the surrounding lymph spaces, with a resulting localized edema (Fig. 20).

In the edges of the groove in which the ovum was embedded, and also deeper down, a more decided *hemorrhagic edema* took place and minute blood extravasations forced their way up to the epithelium and lifted it off. The resulting coagulation, containing likewise the remains of these cells, *serves to close the opening and to cover the ovum* (Fig. 16).

Capsularis.—In place of the old view of a reflexa growing up and above the ovum supposedly situated on the epithelium lining the uterus, a *centrifugal descent* of the ovum must be taken for granted. In v. Spee's young ovum the capsularis had not yet united at the summit. In Leopold's case (ovum seven to eight days) union had already occurred. In Peters' case the summit was closed by fibrin. Leopold and almost all investigators find the structure at this point atypical. The same changes are found as Sänger remarked in the organization of hematocele capsules, *i.e.*, fibrin, connective tissue, and capillaries. Keibel, Kollman, and Reichert say that at the summit of the capsule union results in the formation of what may be called a cicatrix, to which is given the name cicatrix of Reichert.

As proof of the ready and *early connection between maternal blood and fetal tissues*, it may be mentioned that the tissue which separates the capillaries from the uterine epithelium becomes extremely thin in the decidua menstrualis. The capillaries which run a twisted course between the glands of the spongy layer extend up to the epithelium under the mucosa. In the compacta or superficial layer they often possess only an endothelial wall. When the ovum sinks into the decidua the decidua basalis presents in addition a huge dilatation of the capillaries. *The zone which surrounds the ovum is formed by the compacta.* As the ovum descends, gradually more compacta surrounds it, forming the so-called *enveloping zone*.

The Enveloping Zone is the layer of the compacta immediately about the ovum. It furnishes the blood supply to the growing blastocyst. Through its edematous infiltration it renders the centrifugal descent of the ovum easy.

CHAPTER IV.

THE EARLY DEVELOPMENT OF THE HUMAN OVUM.

The early cell division taking place in an ovum is a more or less even increase, which later gives way to a development along specific lines in certain and well-distinguished areas, so that larger or smaller cell complexes pass through varying phases leading ultimately to the production of *embryonal* and *extra-embryonal* structures. Preliminary to this stage the cells of the ovum, which lie loosely together in the earliest period of cell division, become more closely united, so that an epithelium is formed. The epithelial membranes resulting from this early cell



FIG. 17.

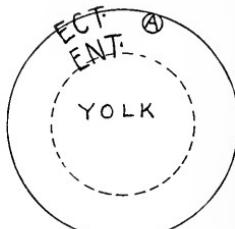


FIG. 17a.

FIG. 17.—Scheme of ovum at stage of simple chorion ectoderm and entoderm.

FIG. 17a.—A, amnion. Scheme of ovum with formation of amnion from the ectoderm.

division are called germinal layers (Fig. 17). It is in them that the various specific areas of development subsequently appear. On the two germinal layers (ectoderm and entoderm) lining the germinal vesicle or ovum and on the subsequent complexes of these layers, two surfaces are to be distinguished: (1) An inner surface or base facing the cavity of the ovum, and (2) an external or free surface. The various cells of a germinal layer may increase or advance in either direction. If the cells advance from the basal side this constitutes an invagination; if from the free surface, an outgrowth or papillation.

The germinal vesicle increases in size through an increase of fluid in its centre, and is composed of two original boundary

layers lying together: an outer, the *chorion ectoderm*, and an inner, lining the cavity, the *entoderm*. This combination is called the germinal vesicle and probably represents the stage immediately after embedding. At a certain point is an oval area, the *area embryonalis*, in which appears an additional very small cavity, the *amniotic cavity*, in the wall of which the *embryo* first appears. The exact mode of this production cannot be positively stated, but it follows one or two closely allied forms. It may be formed through an invagination of the ectoderm (Fig. 17a), the folds in the area of invagination lying close together and uniting quickly, thus leaving within the ectoderm a small enclosed cavity. The very smallest amniotic cavities (flying-dog, guinea-pig) are formed by a solid group of cells passing out from the ectoderm and finally becoming separated from it. In the

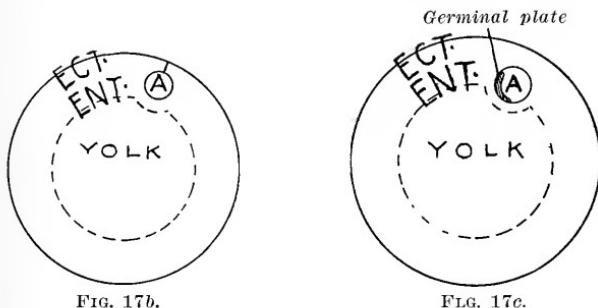


FIG. 17b.—Ovum with ectodermal amnion separated from the chorion ectoderm. A, amnion.

FIG. 17c.—Ovum with ectoderm, entoderm, amnion, germinal plate, but no mesoderm. (Schematic.)

centre of this solid group the subsequent amniotic cavity develops (Fig. 17b).

The early amniotic cavity of the human ovum is extremely small. In no animals where an amniotic cavity is formed through the production of reflected and approaching germinal folds does such a small amniotic cavity result. Therefore the amniotic cavity of the human ovum follows either of the two aforementioned processes. By either of these two methods, then, an amniotic cavity passes out from the ectoderm and pushes the entoderm toward the central space of the germinal vesicle. (It is possible that the amnion develops in the embryonal sphere of Fig. 13. We know little about the origin of the entodermal space, and it is described as the lining of the germinal vesicle, although it is possible that it develops in a manner resembling

the amniotic cavity.) In the human ovum, in all probability, the early stage consists of small ectodermal amniotic cavity sunk into a depression in the closed yolk vesicle lined with entoderm, these being surrounded by the single layered ectoblast vesicle, the later chorionic ectoderm (Fig. 17c). At this stage no mesoderm is present.

On a portion of the ectodermal lining of the amniotic cavity appears an area, the *primitive streak*. The cells about it become differentiated into the ectodermal *germinal plate* (the future embryo). In the primitive streak is a furrow—the *primitive furrow*—which lies, therefore, in the middle of the later germinal plate. By a growth of cells of the germinal plate on either side

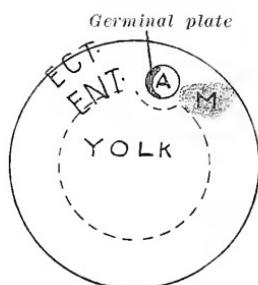


FIG. 18.

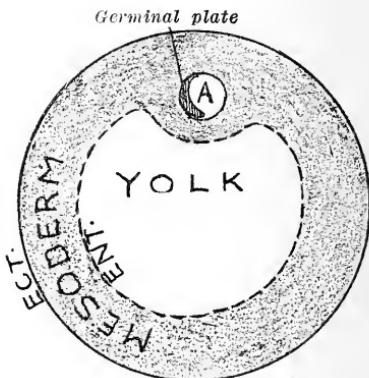


FIG. 18a.

FIG. 18.—Ovum showing the beginning of a mesodermal growth at the caudal end of the germinal plate. (Schematic.)

FIG. 18a.—Ovum with completed growth of mesoderm. (Schematic.)

and over the primitive furrow (*primitive medullary plates*) a canal results—the medullary canal (Fig. 30a).

There begins, with the appearance of the amniotic cavity and the differentiation of a portion of its lining into the ectodermal germinal plate, the development of mesoderm at the caudal end of the latter (Fig. 18). This growth of mesoderm begins close to the inner surface of the chorion ectoderm and forms (1) a thick mesodermal mass, the *caudal knot* of the primitive streak, extending about the caudal side of the amnion up to the chorion (Fig. 18); (2) thin mesodermal layers which grow between the chorion ectoderm and the ectodermal amniotic cavity, and the entodermal yolk vesicle.

In the schematic drawings the entodermal yolk vesicle has been drawn large to make the relations clear. Whatever its size may have been at the beginning, and whether it originates as a very small vesicle in an embryonal nodule or not, it is certain that on growth of the mesoderm the amniotic and yolk cavities are found in the embryonal area of the ovum and take up a relatively small portion of it.

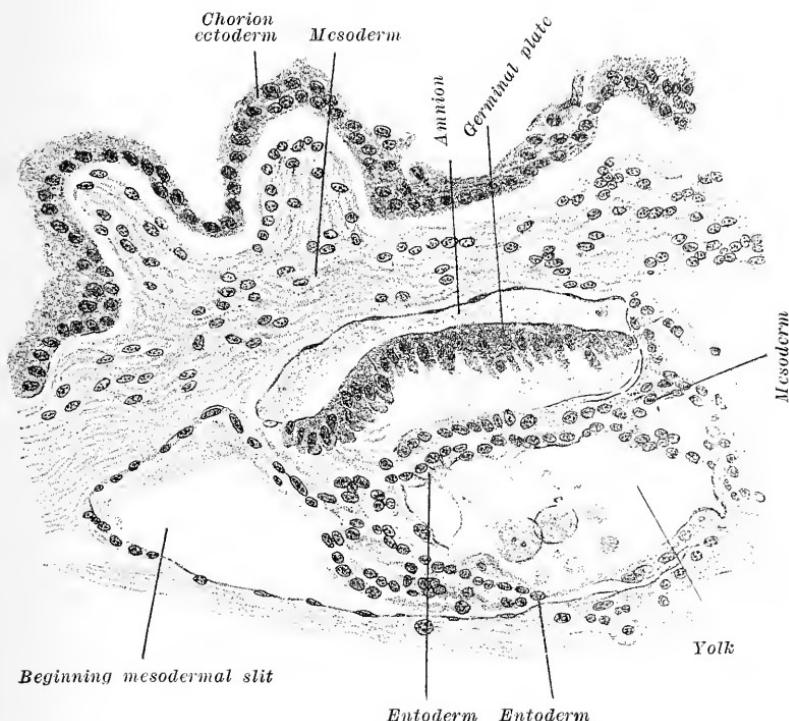


FIG. 19.—A part of the periphery of the ovum of Peters, showing actual conditions pictured schematically in Fig. 18a. Yolk sac is very small compared with Fig. 18a, and naturally the amount of mesoderm below the germinal plate and the yolk sac is much larger than in Fig. 18a. Fig. 19 is the E.F. of Fig. 20.

The ectoderm of the ovum develops hugely, forming the chorionic ectodermal cover or trophoblast.

This stage is well represented in the embryonal formation of Peters (Fig. 19). The ovum is surrounded by chorion ectoblast formation. Cells are more plentiful in the mesoderm near the chorion, but the mesoderm is thick only in the region of the embryonal formation. The internal part of the oval cavity is poor in cells, between which is a weakly staining, fibred, granular

mass. The embryonal formation consists of two small epithelial cavities, the ectodermal amniotic and the entoderm yolk vesicle surrounded by mesoderm and embedded in a thickening of the mesoblast near the chorion. The amniotic cavity is entirely closed. Its wall is differentiated into the very thin amniotic membrane and into the germinal plate, composed of high cylindrical cells. Between these and the entoderm cells of the yolk, the future umbilical vesicle, is a layer of mesoderm cells, which are separated from the ectoblast area of the amnion by a membrana prima which always develops at the border between ectoderm and mesoderm.

DIVISION INTO EMBRYONAL AND EXTRA-EMBRYONAL AREAS.

Through the appearance of a slit in the entire circumference of the mesoderm beginning at the caudal end of the germinal plate, but not dividing the dense mass of mesoderm at the caudal end, the amnion with its germinal plate and the future umbilical vesicle are separated from the chorion ectoderm up to the dense mass of mesoderm, which then constitutes the point of union between the embryonal formation and the chorion ectoderm. This

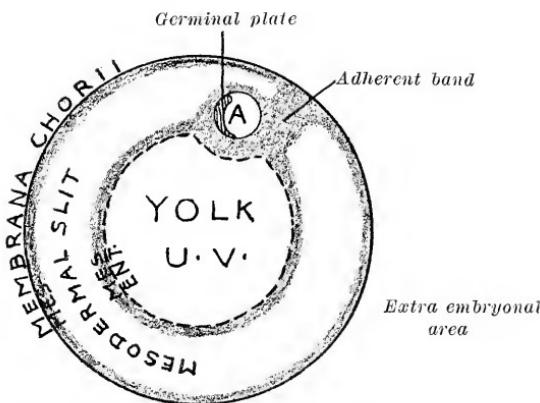


FIG. 19a.—Ovum. Fig. 18a, after the development of the mesodermal peri-embryonal slit. (Schematic.) The adherent band of mesoderm connects the embryonal and extra-embryonal divisions.

point of union contains the adherent band (future abdominal pedicle) and the caudal knot of the primitive streak (Fig. 19a).

The amnion and embryo develop from a small area in the germinal covering, and with the umbilical vesicle are separated from the EXTRA-EMBRYONAL AREA of the ovum, with which they are connected through the adherent band of mesoderm alone. *The embryo develops from a portion of the ectodermal lining of the amnion.*

The extra-embryonal area of the ovum forms the trophoblast, villi, the chorionic membrane, and the placenta.

CHAPTER V.

THE TROPHOBlast IN THE OVA OF ANIMALS.

THE EARLIEST DEVELOPMENT OF THE ECTODERMAL EXTRA-EMBRYONAL AREA OF ANIMAL OVA.

In discussing the histology of gestation it is important to note that in this field, as well as in that of embryology, many of the most important and valuable points have been learned through the study of like processes in animals. The results obtained in investigations in the latter have been applied more or less closely to the various developmental changes in the human being. Though oftentimes erroneously applied, they have been the source through which many vexed questions have been settled. More recent investigations, however, prove that it is not so much the erroneous applications of the results gained in the study of animals as it is errors made in these observations which have so long delayed a satisfactory conclusion concerning the various processes involved in human placentation. It is necessary, for that reason, to consider first the more recent studies concerning animal placentation, as reviewed by Strahl.

Marchand examined the placentæ of rabbits gravid from eight to sixteen days. In the earliest specimens he finds on the ovum a growth of ectoderm elements consisting of two layers: (1) a deeper layer of separated cells and (2) a superficial plasmodial. Since the latter is still covered in part by the zona, it is necessarily of fetal origin. In addition he observes *transitions from the cell layer of the ectoderm into the plasmodial*. The deeper cells become larger and are arranged in groups. Their cell boundaries disappear, and, through a union of these cells, a plasmodial layer is formed, covering the deeper ectoderm, but, as a rule, distinctly outlined from it. This ectodermal plasmodium unites with the syncytially-changed uterine epithelium, forming the first connection between the ovum and the uterine wall. In the placenta of nine to ten days Marchand finds blood (1) in the maternal vessels possessing an endothelial lining and (2) *in ectodermal spaces in the cell layer*. Into these spaces, which possess no endothelium, the blood from the maternal vessels enters.

In the placenta of eleven days few of these unlined ectodermal spaces containing blood remain. Most of them are now lined with a continuous layer of elements rich in protoplasm, which Marchand considers originate from the maternal vessel endothelium (?), which has undergone a "syncytial change."

Maximow, in his investigations on rabbit placentæ, denies, in contrast to Marchand, the existence of a fetal ectodermal plasmodium as early as the eighth day. He believes that, when the ovum attaches itself to the uterine wall, the ectoderm consists of only *separated cells, which come in contact with the epithelium of the uterus and cause it to degenerate.* When the ectoderm, after disappearance of the uterine epithelium, reaches the maternal vessels, the glycogen-containing cells of the uterine connective tissue change to large polynuclear structures. In these cells he finds elements resembling red blood cells in form and color. Only later, when the ectoderm enters into the decidua, does it divide into two layers: (1) one with cell boundaries, the cytotblast, and (2) one without cell boundaries, the plasmoblast. The latter is found only where the villi come in contact with the glycogen-containing cells, *at the walls of the maternal vessels.* It is not found where the villi grow into the connective tissue between the vessels. From the tenth day on, spaces filled with maternal blood are found in the fetal plasmodium and there develops an ecto-placenta, *into whose ectodermal villous formations the mesodermal elements bringing the allantoic vessels enter.*

Opitz finds in the rabbit a double ectodermal layer during the process of attachment of the ovum in the uterine wall: (1) an internal layer formed of separated cells, and (2) an external plasmoidal layer. *The latter causes the uterine syncytium to disappear and aids the attachment of the ovum to the uterine wall freed of epithelium.* In the plasmodium spaces occur into which maternal blood empties from the eroded maternal vessels. From the ovum, ectodermal cell groups enter into this ectodermal plasmodium, *are vascularized, and later become also plasmoidal.*

Those syncytial cells which Marchand considers as resulting from maternal endothelium, Maximow, however, views as ectodermal plasmodium when found in the fetal portion of the vessels, but considers them in the maternal portion to be endothelial. Opitz considers these cells *to be entirely of ectodermal origin.* It is to be noted that Maximow and Opitz mention the destruction of the uterine epithelium. Later fetal mesoderm with the allantoic

vessels enters into the placenta and on its external surface the remnants of the ectoderm are found as flat covering cells. The epithelium of the uterus plays no part in the development of the placenta, *but forms a syncytium which is absorbed by the ectoderm cells.*

In Tarsius, Hubrecht finds that the uterine wall, through a change of the connective tissue, forms a trophospongia, *between which the glands degenerate.* On the growth of the embryonal ectoderm or trophoblast, a mixture of trophoblast and trophospongia occurs, with a decided increase in the volume of this early placental formation. Then the development of the mesodermal villous elements takes place, and the mesoderm villi surround themselves with a trophoblast covering. Between the trophoblast cells lacunæ form, into which maternal blood passes. Finally, the trophospongia becomes a thin layer which represents the boundary toward the deeper maternal tissue.

In Tupaja, when the ovum rests upon the uterine wall *its ectoderm causes the uterine epithelium to disappear.* The trophoblast grows decidedly, with the formation of giant cells, and occasionally forms a combined layer with the uterine epithelium in which maternal and embryonal nuclei lie in a common plasmodium. *The maternal portions degenerate,* but the ectodermal tissue is divided into (1) a deeper layer, the cytотrophoblast, and (2) a superficial syncytial layer, the plasmодитrophобласт, which constitutes only a temporary differentiation. In the uterine wall, in the meantime, the connective-tissue trophospongia is formed; the border between ectoderm and maternal connective tissue disappears, for these tissues infiltrate each other. In this resulting union the trophospongia is overcome by the trophoblast and the maternal blood passes from the maternal capillaries into the embryonal trophoblast spaces. The mesodermal fetal cells then enter with the allantoic vessels and receive on their external surface a covering of trophoblast.

Although various investigations have shown that the variations in the development of the placenta in different animals are quite unexpected, and that among the individual mammalia, even when closely related, decided differences are found, yet the trend is more toward the view that, as a rule, the syncytium originates from fetal ectoderm. According to Strahl and Selenka, in quite a group of placenta the uterine epithelium plays no unimportant rôle. On the other hand, Fränkel, as a result of extensive investigations, comes to the conclusion that *the*

higher the organization of the placenta and the more firm the connection between maternal and fetal tissues, so much the less is the maternal epithelium preserved.

He finds that in the rodents and in the insectivora, whose placentæ, of all the animals he examined, stand nearest to the human placenta, *the uterine epithelium ceases at the border of the placenta.* Only in the pig does it remain. In the cow and sheep the epithelium in the cotyledo shows a tendency to degeneration. In the cat, rabbit, squirrel, guinea-pig, rat, mouse, and mole the uterine epithelium takes no part in the formation of the placenta. A so-called syncytial formation, however, he finds to occur in the most varying tissues. *The higher in the animal plane the animal stands, the more does the chorionic epithelium grow into the maternal connective tissue robbed of its epithelium.*

Opitz finds that in the guinea-pig the foundation for the placenta is the fetal ectodermal plasmodium vascularized by maternal vessels. The placentæ of the rabbit, the guinea-pig, and the human being agree in that earlier or later the ectodermal surface of the ovum comes in contact with the connective tissue of the uterine mucosa. Finally, between maternal blood and the fetal vessels found in the mesoderm of the villi only one layer of the ectodermal syncytium is left. The apparently great differences are to be explained by the fact that in the rabbit and guinea-pig the projections of ectoderm and mesoderm (the villi) come into union with each other, while in the human being they are always separated.

CHAPTER VI.

THE TROPHOBLAST OF THE HUMAN OVUM.

THE EARLY DEVELOPMENT OF THE ECTODERMAL EXTRA-EMBRYONAL AREA.

Trophoblast.—The outer layer of the ovum consists of trophoblast. *It is a product of the ectoderm* and from it develop the future syncytium and, according to most authors, the cells of

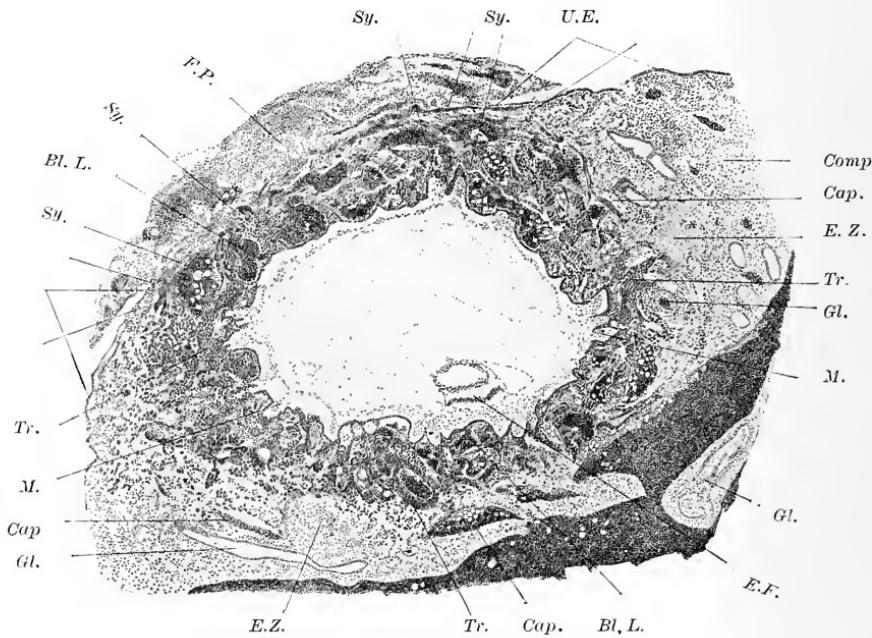


FIG. 20.—Low-power drawing of the trophoblast, enveloping zone, etc., of an ovum three days old, in transverse section. (Peters.) *Sy.*, syncytium; *F.P.*, plug of fibrin at the summit of the capsularis; *Bl. L.*, blood lacunæ; *Tr.*, trophoblast; *M.*, mesoderm; *Cap.*, capillaries; *Gl.*, glands; *E.Z.*, enveloping zone; *E.F.*, embryo formation; *Comp.*, compacta; *U.E.*, uterine epithelium; *Gl.*, gland.

Langhans. The “trophoblast,” according to Hubrecht, “is the epiblast of the blastocyst, as far as it has a direct nutritive significance as indicated by proliferating processes, by immediate contact with maternal tissues, maternal blood, or secreted material.” It consists of small and large cells of various forms.

Shortly after the ovum descends into the mucosa a connection

between the trophoblast and the maternal blood takes place. As all the capillaries near the trophoblast consist of endothelium and evidence many blood sinuses, the trophoblast is infiltrated with blood lacunæ which are separated from each other by these trophoblast cell groups. As a result of the growth of these cells, of congestion and of pressure, the blood takes its exit from the capillaries and bathes the cells. The lacunæ, then, represent a united system fed by peripheral capillaries and are separated externally from the decidua by a thin layer of ectodermal trophoblast cells, but the *trophoblast at various points may extend much further into the compacta*.

The ectoblast cells are of varying form: 1. Cubical, with large, strongly stained round or oval nucleus containing granules and

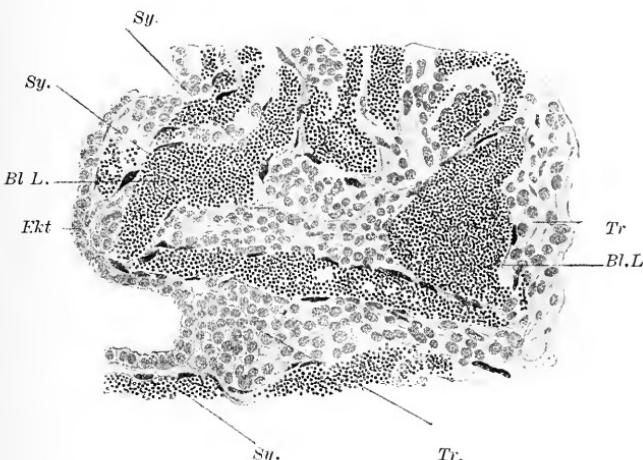


FIG. 21.—Section through the central portion of the trophoblast layer. The trophoblast (*Tr.*) is covered with syncytium (*Sy.*) at all points in contact with the blood lacunæ (*Bl.L.*). *Ekt.*, ectoderm. (Peters.)

nucleolus. 2. These become changed. The nucleus becomes pale and swollen. Vacuoles appear in the cells and strongly stained fragments of the nucleus are evident. The cells are irregular in form. 3. The nuclei are larger, especially in the external layers, and more swollen. The chromatin network is irregular and dark lumps appear in the nucleus. The nuclei are of various forms, oval, spindle-shaped, and contain vacuoles. 4. Swollen groups of nuclei in degeneration are found. These nuclei contain one or more nucleoli. 5. The protoplasm of these cells unites, forming protoplasmic masses in the shape of bands or irregular groups with extensions.

From the simple cubical cells of the central layer of the trophoblast we find, in going toward the periphery, gradual changes from 1 to 5. The trophoblast cells grow through direct cell division. These changes represent the results of the *corrosive action of the blood* on these cells, whereby the future syncytium is formed. The syncytium then lines the lacunæ (Fig. 22). The endothelium of the lacunæ and of the capillaries degenerates wherever it is not lined by syncytium. The syncytium is found also under the endothelium of the most peripheral trophoblast

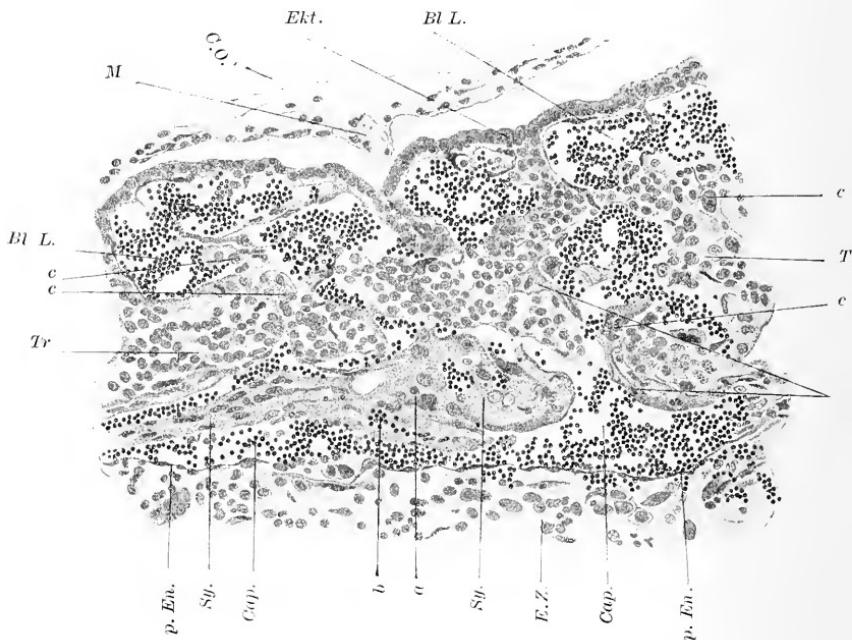


FIG. 22.—A capillary (*Cap.*) in the serotinal portion of the trophoblast infiltrated with blood lacunæ (*Bl.L.*). The peripheral wall of the capillary is preserved and forms the serotinal wall of the intervillous space at this point. The capillary at many points communicates with the lacunæ (*Bl.L.*). In the capillary is a large mass of syncytium (*Sy.*) containing changed trophoblast nuclei (*a*) and red blood cells (*b*). At *c* changes in the trophoblast nuclei are evident. (Peters.) *Tr.*, trophoblast; *Ekt.*, ectoderm; *M.*, mesoderm; *C.O.*, central cavity of ovum; *p. En.*, peripheral endothelium.

masses, and proves that the *endothelium plays no part in the formation of the syncytium*.

It is to be noted that the opening of the maternal vessels occurs at this early stage, *before villi are formed*. The endothelium of the capillaries degenerates and the blood makes its exit from

the capillaries. It ought to coagulate, but does not. It circulates against the fetal cells, which have the power to prevent coagulation, and yet the blood exerts a deleterious influence on these cells. Thus the resulting syncytium lines the lacunæ, prevents coagulation in this primary intervillous space and later about the villi, separating the blood at all times from the cells of the chorionic centre. Free syncytial cells are present, even in the vessels of the periphery in the compacta. At later periods they are found free in the veins and arteries. It is thus seen that *the cells of the trophoblast of the ovum enter the veins at an early period.*

The Primary Intervillous Space is intravascular and is bounded by maternal and fetal structures. That the developed inter-

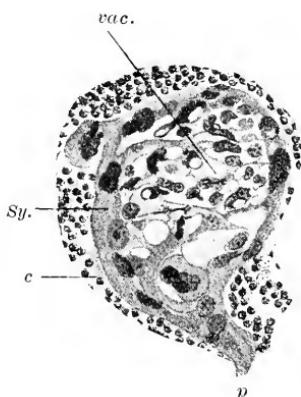


FIG. 23.—Change of trophoblast to syncytium (*Sy.*) The syncytially changed trophoblast is infiltrated with numerous vacuoles (*vac.*). *c*, red blood cells of the capillary in which this syncytial mass was floating attached by a pedicle (*p*). (Peters.)

villous space contains free blood has been granted by Virchow, Kölliker, Langhans, Waldeyer, etc. The drawings of Peters indicate a slowing and probable stagnation of the blood current. The maternal endothelium on the side toward the compacta is at first intact and forms the serotinal wall of the space (Fig. 22). In it are found isolated cells and groups of trophoblast. Trophoblast cells are also present on the serotinal wall, so that this wall consists of trophoblast, maternal endothelium, and decidual tissue. Later the endothelium of the serotinal side is found only in the areas corresponding to the openings of the maternal vessels. Where the trophoblast cells extend into the enveloping zone in the form of bands, uniting with each other in a network, it is

often difficult to distinguish between maternal and fetal cells. Merttens says the same. *Thus the earliest ectoderm cells enter actively into the maternal tissues.*

The primary intervillous space is formed through the gradual consumption of the trophoblast. Through its growth more and more layers of the compacta are included in the enveloping zone, until the corrosive action of the blood has furnished the villous mantle of the ovum. 1. The trophoblast is reduced to a single layer. 2. The union of the lacunæ at first separated by large trophoblast cell masses takes place. The origin and growth of the intervillous space thus goes hand in hand with the various steps leading to the formation of villi. In the ovum of Peters no villi are yet present. The inner surface of the trophoblast shows irregular, finger-like depressions into which mesoderm is beginning to enter. The lacunæ have enlarged and the cell

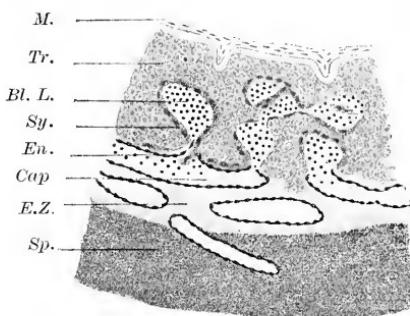


FIG. 24.—Schematic representation of the earliest stage of the development of the placenta. (Peters.) *M.*, mesoderm; *Tr.*, trophoblast; *Bl. L.*, blood lacunæ; *Sy.*, syncytium; *En.*, endothelium; *Cap.*, capillary; *E.Z.*, enveloping zone; *Sp.*, spongiosa.

masses between them have become smaller. Through the entrance of the growing mesoderm into these divided cell masses, which are finally reduced by the blood to the syncytial layer, villi are formed and the primary intervillous space becomes larger. *The primary enveloping zone becomes the subsequent intervillous space.*

The maternal endothelium breaks down and is found floating in the lacunæ. It is never seen in a proliferating stage. Syncytium is found under it. It is impossible for the endothelium to fill out all the spaces of the trophoblast, and, if it did do so, we ought to see it somewhere in the process of growth. On the contrary, it is always seen to degenerate. Therefore *the syncytium does not originate from maternal endothelium.*

The syncytium is found over the surface of the ovum, in the trophoblast, in the lacunæ, toward the enveloping zone, and also in the periphery of the ectodermal trophoblast. At the summit of the ovum syncytium is also present. In numerous points the direct transition from trophoblast cells to syncytium is distinctly seen. Therefore *it does not originate from the uterine epithelium.*

The glands in the decidua about the ovum become filled with blood. They are pushed aside by the growing ovum and are broken into by the trophoblast and syncytium. Their epithelium degenerates and disappears. *The uterine epithelium plays no part in the formation of the syncytium.*

The syncytium consists of a shining, granular protoplasm containing numerous nuclei. The nuclei are round, oval, or flattened. There are also vacuoles in the protoplasm, which make the latter appear like septa in which are flat and half-moon-shaped nuclei (Fig. 23).

Van Siegenbeck finds syncytial giant cells especially near the union between maternal and fetal tissues, in and about the lacunæ, between the ectoblast and the compacta, covering the free compacta, about the maternal capillaries, and even more externally in the vessels. He finds a gradual transition from trophoblast to syncytium. In one place he observed a syncytial giant cell between the mesoblast, and the ectoblast of the fetal sac, which he considers as a proof of the ability of these cells to wander into surrounding tissues.

The processes, so far, are well represented in the schematic drawing, Fig. 24.

Merttens found in the particles obtained by curetting sixteen days after the last menstruation a few sections of an ovum with the enveloping decidua. He describes (1) the chorion and the villi; on the latter are cell groups, and between them are found spaces constituting the intervillous space; (2) the spongiosa, containing numerous glands, and (3) the compacta, a pale zone lying between 1 and 2, and containing large pale cells between which are numerous darkly-stained nuclei.

In the compacta he finds large cells and large spaces. Between the large cells syncytial masses are irregularly distributed. The cell spaces are round, long, or irregular, and often empty into the intervillous space. They contain blood and are all lined with syncytial masses, which at some points are very thin, and at other points quite thick. Where these spaces empty into the

intervillous space the syncytial masses continue into the syncytium of the villi. Merttens does not know whether these spaces are glands, vessels, or lymph spaces. *They are possibly, however, the lacunæ of Hubrecht and Peters.*

The epithelia of the glands are like beaker cells, and on their free tips is a half-moon of glycogen. The glands evidence papillary projections (compare Fig. 14b). Merttens believes that the syncytium results through a change of the surface and gland epithelium (?). In his Fig. 8 Merttens represents, in the upper portion, the epithelium of such a gland, and the lower half represents syncytial masses. He believes that the latter results from these epithelial cells, which lose their cylindrical form and unite. From his description and his drawings it is neither clear nor probable that the epithelium goes over into these syncytial masses. Besides, this area is taken from decidua particles not connected with the ovum, and at no point can he find the epithelium of the glands contributing in any way to the syncytium of the chorion.

On the contrary, *this ovum is at the stage where the trophoblast is almost consumed by the growth of the intervillous space*, and the cell groups at the end of the villi, which Merttens calls points of union between the villi and the decidua, are really the remains of the trophoblast (Fig. 31).

CHAPTER VII.

THE FURTHER DEVELOPMENT OF THE HUMAN OVUM.

THE EARLY DEVELOPMENT OF THE EMBRYONAL AREA.

1. The ovum *v. H.*, described by Spee, was thrown off after five weeks menopause. The ovum contains an oval germinal plate entirely in the stage of the primitive streak and situated in the wall of the amniotic cavity. The entire embryonal formation is 0.4 millimetre long. The greatest diameter of the ovum including the villi is $5\frac{1}{2}$ to 7 millimetres. Under the chorion was found a thin layer of connective tissue poor in cells and bounding directly the mesodermal slit. From this layer the chorion could be lifted off. The inner part of the ovum contained a bubble of

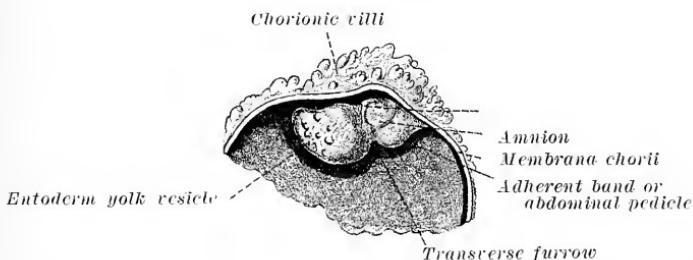


FIG. 25.—Ovum *v.H.* (Spee), showing embryonal and extra-embryonal areas. The extra-embryonal villi and membrana chori are connected with the embryonal formation by the adherent band of mesoderm.

air and a non-celled fibred coagulation substance in which, on the serotinal side, was seen the embryonal formation, which included the amniotic cavity with the germinal plate and the umbilical vesicle (Fig. 25).

The primitive streak region is small, and yet the development of mesoderm within the chorion, in the abdominal pedicle and about the amniotic cavity and about the umbilical vesicle, is so great that considerable time must have been needed for its development. At the time when the mesoderm began to develop, the ovum was probably of a diameter of 0.5 millimetre. At the first beginning of the mesoderm slit the ovum was probably of a diameter of 1 millimetre.

The embryonal formation is a long, thick projection connected

at one end with the inner side of the chorion, but otherwise projecting free into the cavity of the vesicle, that is, the periembryonal mesoderm slit (Fig. 25). A superficial transverse furrow divides it into two elliptical parts. The larger is at the free end and contains the entoderm vesicle. The smaller part contains the amniotic cavity lined with ectoderm and consists, in

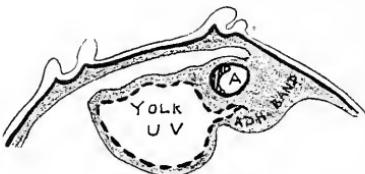


FIG. 26.—Longitudinal section through Fig. 25, ovum *v.H.* of Spee. *U.V.*, the future umbilical vesicle. The adherent band of mesoderm connects the embryonal formation with the extra-embryonal membranes.

addition, of the compact band of mesoderm which covers three-fourths of the amnion, from the mesodermal covering of the entoderm vesicle up to the chorion (Fig. 26). This compact band is the *adherent pedicle* and corresponds to the caudal end of the primitive streak. The entoderm vesicle sends a blind duct, lined with entoderm, into the adherent pedicle. The entoderm vesicle

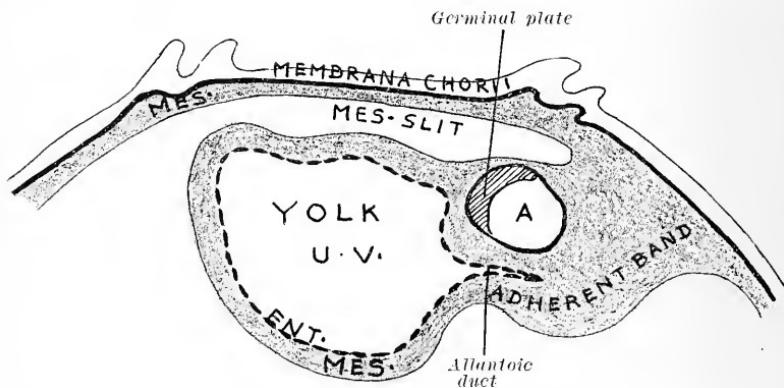


FIG. 27.—Fig. 26 enlarged, showing actual conditions in the ovum *v.H.* pictured schematically in Fig. 19a. *U.V.*, the future umbilical vesicle; *ENT.*, entoderm; *MES.*, mesoderm.

lined with a single row of cubical cells, and covered with mesoderm, is the *umbilical vesicle*. The amniotic cavity is lined with ectoderm and is likewise covered with mesoderm. The single layer of flat cells lining the amniotic cavity begins to thicken at a certain point; the cells become high cylindrical and constitute the *germinal plate*. That part of the ectoderm of the

amniotic cavity lying on the umbilical vesicle enters, then, into the formation of this thick germinal plate. The mesoderm which covers the umbilical vesicle and the amnion unites in the narrow zone which separates the two. *Here where the walls of the umbilical vesicle and amnion run parallel, the germinal plate is found on the inner surface of the amniotic cavity.* From the caudal knot up to the chorion extends the adherent pedicle, the future *abdominal pedicle*, and here the mesoderm lining the

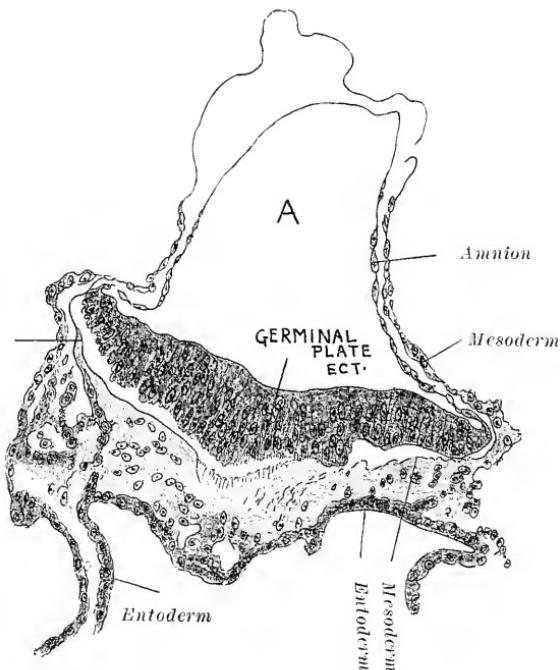


FIG. 28.—Transverse section through germinal plate of Figs. 26 and 27 (Spee.)

chorion and that found in the embryonal formation unite (Figs. 26, 27).

Posterior to the caudal knot of the germinal plate, the umbilical vesicle sends a prolongation, the allantoic duct, into the abdominal pedicle, which duct is separated from the caudal side of the amnion by mesoderm. It is relatively long and probably formed early. It is *through the adherent band of mesoderm that the vessels of the fetus pass to the chorionic membrane and into the villi.*

The caudal knot of the primitive streak region forms the ab-

abdominal pedicle. The germinal plate is only a portion of the completed primitive streak region. There is as yet no differentiation into medullary plates or chorda. The germinal plate has its dorsal surface toward the point of union of the abdominal

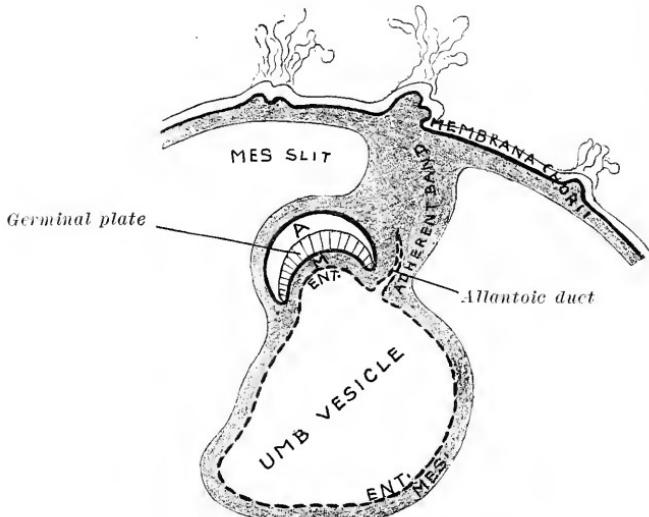


FIG. 29.—Change from Fig. 27, showing ventral curve due to growth in length of germinal plate. (Schematic.) *M*, mesoderm.

pedicle to the chorion and its long axis runs radially to the chorion. It is of oval form and has a median furrow on its dorsal surface. Its dorsal surface follows, in addition, the curve

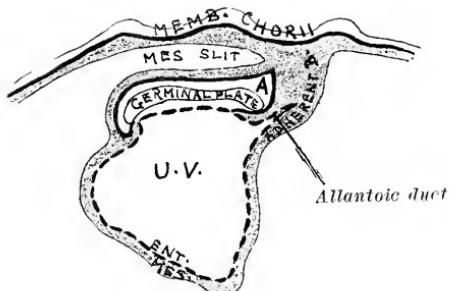


FIG. 29a.—Stage of still further growth of cephalic end of germinal plate. Embryo *Gle* of Spee.

of the amniotic cavity and is therefore concave. Its dorsal furrow lies in the same plane as the lumen of the allantoic duct, and its caudal portion is at right angles to the duct. It consists of high cylindrical cells, highest at the middle (Fig. 28). Laterally

it extends on into the amniotic lining. Laterally, ectoderm and mesoderm are separated by a membrana prima, which is always present between parallel layers of mesoderm and ectoderm. In the median line, however, is a fibred, granular substance, and here possibly a connection between ectoderm and mesoderm existed. A characteristic connection at the region of the primitive streak is present in mammalia and in older human embryos. There is probably no connection between mesoderm and entoderm, and in the next embryo there is no layer continuity be-

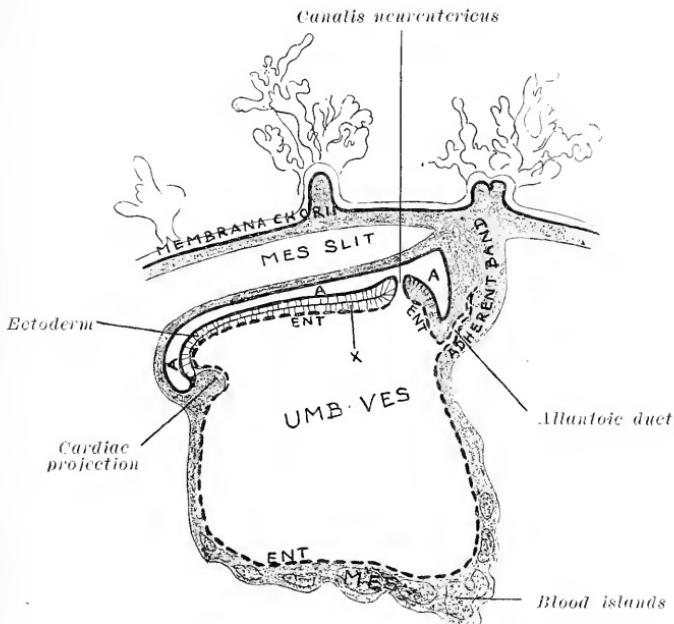


FIG. 29b.—Embryo Gle of Spee. A, amnion cavity. X, between ectoderm and entoderm mesoderm comes. All anterior to the canalis neuretericus is new as compared with embryo v.H., Fig. 27.

tween mesoderm and entoderm posterior to the canalis neuretericus.

It is to be noted that in Fig. 27 the germinal plate has a dorsal curve. The next stage is represented by a ventral curve (Fig. 29). As a result of this (1) the mesoderm between the germinal plate and the entodermal vesicle, and (2) *that part of the entodermal vesicle lying parallel to the germinal plate become included in the ventrum of the curved germinal plate and form the mesoderm and entoderm of the fetus.* The actual change is seen in Fig. 29 a and b.

2. The next older embryo (*Gle*) shows that the new growth concerns the *addition of new elements to the germinal plate*, plus an increase in the size of the amniotic cavity and the umbilical vesicle (Fig. 29b). The primitive streak region in *v. H.* has grown a little, but keeps its position approximately. The mesoderm of the abdominal pedicle, the allantoic duct, and the caudal part of the primitive streak region remain about the same. *In their growth a pause ensued*, which pause continues until the embryo has seven primary vertebrae. Therefore the *added growth concerns the cranial end* of the germinal plate, while the caudal end remains a fixed point. The anterior part of the germinal plate has grown to four times the size of that in *v. H.*, with the result that it extends almost at right angles to the original germinal plate, because the caudal end remained a fixed point and thus caused a transposition of the cranial area of the germinal plate. Thus

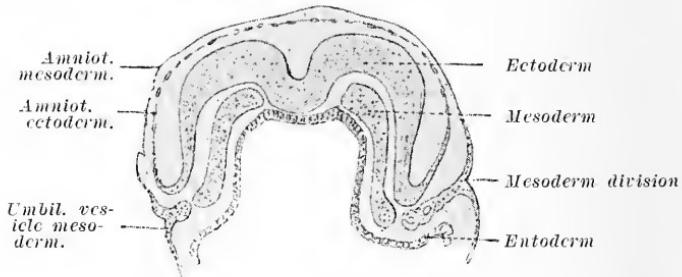


FIG. 30.—The three layers of a human embryo, still without primary vertebrae. Transverse section. (Keibel.) Showing growth of lateral walls, which thus enclose the entoderm within them.

the umbilical vesicle and amniotic cavity are pushed away from their position near the chorion into the deeper part of the mesodermal slit. *A continuation of this process causes the embryo eventually to lie with its ventral surface turned toward the point of adhesion of the abdominal pedicle.* At the same time the umbilical vesicle has grown, too, so that the embryonal formation of *Gle* has added a growth almost as large as *v. H.* itself. In *v. H.* all those areas of the germinal plate are absent which in *Gle* turn their dorsal side to the chorion, that is, *all cranial to a line drawn transversely through the canalis neurentericus* (Fig. 29b).

The germinal plate, as a result of the rapid growth of the medullary plates, grows at right angles to the long axis of the primitive streak region of *v. H.* The primitive furrow has an S shape on the dorsal surface. The cranial end of the chorda formation bends ventrally and ends together with the anterior end of the medullary plate. Anterior to the canalis neurenter-

cus the chorda entoderm and the medullary plate lie close together. At the anterior boundary of the canalis neurentericus

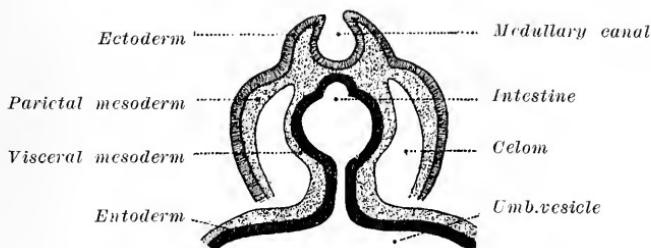


FIG. 30a.—The forming of the intestine. Schematic. (Kollmann.) The mesoderm is divided into a parietal and a visceral layer. The celom, or space between these layers, is the future peritoneum. Intestine and umbilical vesicle are connected by a narrow duct.

the ectoderm bends into the area of the chorda entoderm and the latter possesses elements of the ectoderm.

With the continued growth of the cephalic end and the conse-

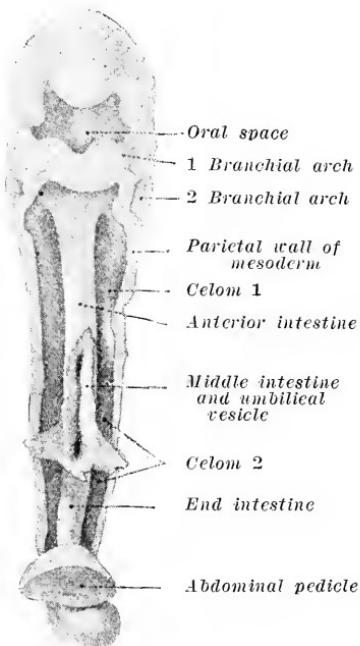


FIG. 30b.—Human embryo, 2-4 millimetres long, with heart and abdominal vesicle removed and umbilical pedicle cut. (After His.) Showing connection between intestine and umbilical vesicle.

quently decided ventral curve of the embryo, as well as through the growth of the lateral walls, a portion of the entodermal sac lies within the embryo. The ectodermal germinal plate has

on its ventral surface then a strip of mesoderm and upon this mesoderm a part of the very endoderm which lines the endodermal

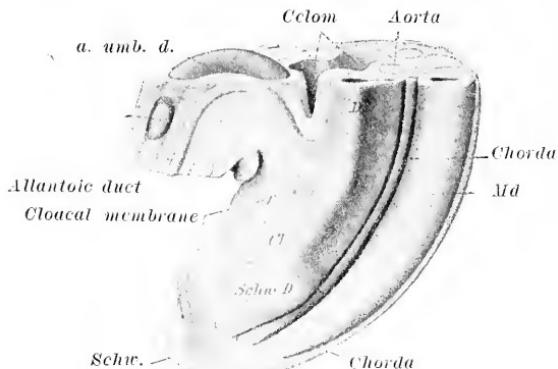


FIG. 30c.—Caudal end of embryo, 3 mm. long. Reconstruction. (Keibel.) *Cl.*, cloaca; *Md.*, medullary canal; *Schw.*, tail; *Schw. D.*, caudal intestine. Showing relation of allantoic duct to the cloaca.

abdominal vesicle (Fig. 30). Anterior to the canalis neurenertericus is found most of the embryo and the medullary furrow and plates. The embryo consists here of three layers.

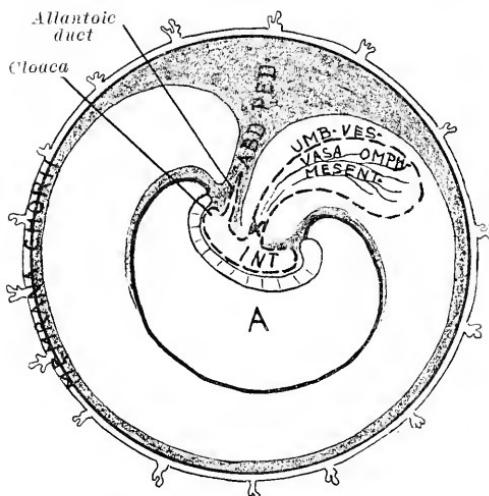


FIG. 30d.—Embryo 29b, with decided ventral curve, has turned so that the abdominal surface faces the adherent band. *X*, omphalo-enteric duct. The heavy black line about *A*, passing off from either end of the embryo, is the amnion. *Int.*, intestine. *Umb. ves.*, umbilical vesicle. (Schematic.) The embryo now has its ventral surface turned toward the abdominal pedicle.

Posterior to the canalis neurenertericus is the original primitive furrow with the primitive lateral plates. The embryo here con-

sists, too, of three layers. The mesoderm of the posterior part goes over into the adherent mesodermal band, the future abdominal pedicle. At the anterior and posterior ends the ectodermal layer of the embryo goes over into the single-layered *amnion*.

The original cephalic and caudal ends of the embryo (the points where ectodermal germinal plate goes over into single-layered amnion) develop decided ventral curves. So do the lateral walls or sides of the germinal plate. As a result, the abdominal vesicle becomes constricted more and more, and the wide connection of the abdominal vesicle with the entodermal area of the embryo is narrowed so that a constantly narrower portion connects the abdominal vesicle with the interior of the embryo (Fig. 30a). The portion *in* the embryo, the entoderm of the embryo, forms the intestine and the *cloaca* (Fig. 30 a and b. The cloaca is

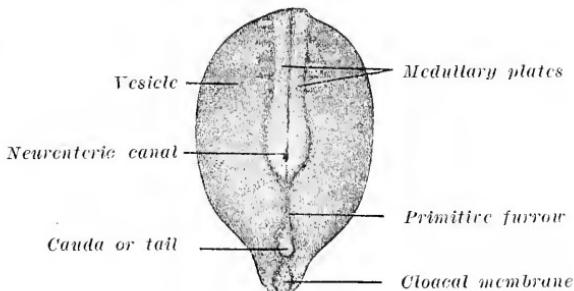


FIG. 30c.—External scheme of embryonal formation, caudal end, with amnion removed, showing relative situation of parts. A combination of Figs. 29b and 30c. (Waldeyer.)

a sac into which intestine and allantoic duct enter (Fig. 30c). As a result the abdominal vesicle is connected with the intestine by what has become a narrow duct, the omphalo-enteric duct, while the allantoic duct *situated in the adherent band* enters the cloaca (Fig. 30d). It may be said, for the sake of a readier understanding, that the cloacal portion of the entoderm sends a prolongation, the allantoic duct, into the adherent band. (On the umbilical vesicle develop vessels, the omphalo-mesenteric vessels, which enter the mesentery of the intestines, while the omphalo-enteric duct enters the intestine near the future cecum.) Later the cloaca becomes divided into a posterior part continuous with the intestine, *the rectum*, and an anterior part, *or bladder*, continuous with the allantoic duct, *the future urachus*.

CHAPTER VIII.

THE CHORIONIC VILLI.

EARLY DEVELOPMENT.

The ectodermal cover of the ovum has the name *chorion-ectoderm* to distinguish it from the *ectoderm in the amniotic cavity*, a part of which takes part in the embryonal formation.

This ectodermal cover evidences a great growth of cells in the entire circumference of the ovum, forming what is called the TROPHOBlast. These trophoblast cells invade the decidual compacta. The capillaries of the compacta dilate into *lacunæ* or wide spaces. The blood breaks through the lacunæ and now, AT THIS EARLY TIME, maternal BLOOD comes in contact with fetal ectodermal trophoblast cells. The maternal blood when it comes into contact with the trophoblast cells changes them to large poly-nuclear cells, consisting of the protoplasm of several cells in which the trophoblast nuclei of these cells form a polynuclear group. Such groups and likewise large mononuclear cells result from the action of the blood on the trophoblast. Such resulting cells are called *syncytial* cells, mononuclear or polynuclear. They line, too, the lacunæ in the circumference of the trophoblast.

The trophoblast cells are constantly spreading in the periphery of the ovum, entering further and further into the decidua. They have a *destructive action* on the decidual cells; they reach capillaries and vessels and enter the latter. Wherever they open lacunæ or vessels the outpoured blood changes the trophoblast cells with which it comes in contact into syncytium. As a result the trophoblast is divided and subdivided into small islands consisting of groups and masses of trophoblast cells. The cells on the outside of these groups are changed by blood to syncytium. Such groups of trophoblast cells surrounded by syncytium are the future villi. Subsequently mesoderm develops in the interior of these trophoblast islands or "cell groups" and we have young villi (Fig. 31). Later most of the trophoblast cells are displaced and disappear with the exception of a single row situated immediately beneath the syncytium. This single row, composed of large distinct cells with pale protoplasm and large nuclei, is called

the cell-layer of Langhans. In the centre of such groups is the developed mesoderm, and the so-called villi are now formed, but as yet no capillaries are present. Such extensions, composed of a centre of mesoderm and a periphery of Langhans cells, with the latter surrounded by syncytium, are the final villi in whose connective-tissue centre capillaries finally develop which are connected with the capillaries of the fetus.

When the trophoblast first develops and extends into the maternal compacta its margin is irregular and serrated (Fig. 20). As capillaries on the compacta are opened syncytium is formed wherever blood touches the trophoblast cells. The points of trophoblast furthest advanced extend continually into new compacta, destroy it, and open more capillaries. In this way the

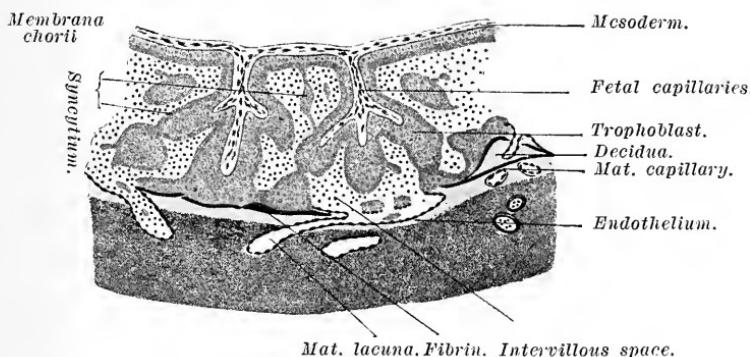


FIG. 31.—Schematic representation of a later stage of placental development than Fig. 24, showing young villi. (Peters.)

process is carried on through a constantly increasing periphery. As a result long, irregular, branching, finger-like extensions of trophoblast project out from the trophoblast layers situated immediately on the external layer of mesoderm produced by the mesoderm slit. These layers are, too, changed to syncytium and cell layer of Langhans. Under them the mesoderm also grows and displaces most of the trophoblast cells. The resulting continuous outer wall of the ovum from which the finger-like extensions (villi) project is called the *membrana chorii* (Figs. 25, 27, 29, 29b, 30d).

To the inner surface of the *membrana chorii* is attached the adherent band, or abdominal pedicle. It is a band of mesoderm which has grown continually longer. In it is the allantoic duct. Through it pass the vessels of the fetus to enter the mesoderm of the *membrana chorii* and the mesoderm of the villi.

The capillaries which develop in the villi unite with the vessels of the fetus in the membrana chorii. Thus, through the medium of the adherent band, fetal blood is brought into the villi. The villi are surrounded by maternal blood and so an exchange between fetal and maternal blood is carried on through the two layers of epithelium covering the villi (Fig. 31).

In the Fourth Week of Uterine Gestation, when the original trophoblast capsule is consumed, *the ovum is loosely connected with the decidua by only a few adherent villi.* The following remarks concern such an ovum, covered with chorionic villi,

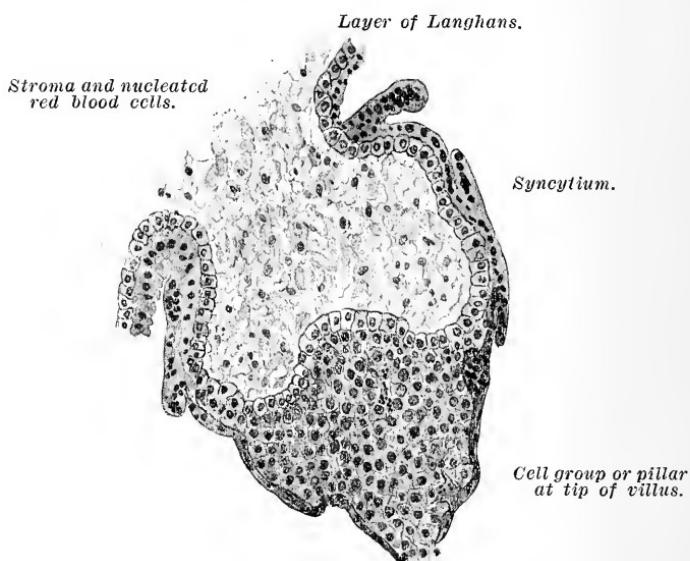


FIG. 31a.—A well-developed villus showing cell layer of Langhans with "cell group or pillar" at its tip. These cell groups are trophoblast cells. The villus is covered with syncytium which is very thin over the "cell group."

aborted in the fourth week. Its attachment in the uterus must have been exceedingly loose, for the numerous and countless villi connected with the covering of the ovum carried with them no decidua cells. It was hardened in alcohol, cut in series sections, and stained with hematoxylin and eosin. Examination showed a point of rupture in the wall through which the fetus had been expelled.

The important points with regard to the villi are as follows: In a well-developed villus (Fig. 31a) we find a connective-tissue centre made up of long, thin, and branching cells situated in a

basis composed of a finely granular, frothy, and often red-staining substance like that found in the fetal sac. On this connective-tissue centre is a single layer of large, sharply outlined mononuclear cells forming a continuous covering of the same. This layer frequently forms groups at the tips of the villi (Fig. 31a). This layer, the so-called layer of Langhans, is composed of large round or polygonal cells with a pale, almost water-clear, protoplasm poor in granules. The nucleus is large, round, and homogeneous and generally contains one nucleolus. The

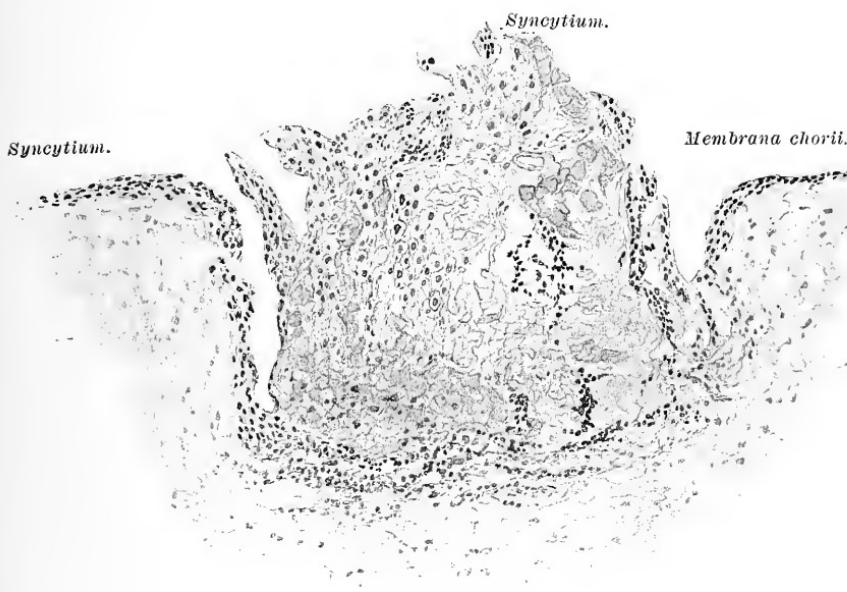


FIG. 31b.—An outgrowth on the membrana chorii containing trophoblast cells in the lower portion, and showing the various changes to distinct Langhans cells in the centre, and especially to syncytium and syncytial masses on contact with maternal blood in the periphery.

layer of Langhans is separated from the stroma of the villus by a sharp outline. The side away from the mesoderm is smooth and covered by a distinct structureless membrane. The outer syncytial covering is a thin layer of protoplasm containing a single row of dark nuclei. In the stroma of these villi may be seen round spheroidal trophoblast cells, and their change into nucleated red blood cells can be readily observed.

The younger villi and the large cell groups which repre-

sent the early formative stages of villi possess the same structure and cells as the outgrowths found on the external surface of the membrana chorii (Fig. 31b). We find villi composed entirely of spheroidal trophoblast cells surrounded by a covering of syncy-



FIG. 32.—Villus composed of trophoblast cells showing transitions to the syncytial covering and to syncytial groups on the left.

tium. The gradual transition from these trophoblast cells into syncytium, and the change of the trophoblast nuclei into the nuclei of the syncytium, can be distinctly observed (Figs. 31b and 32). Other villi are composed of trophoblast cells with a distinct



FIG. 32a.—Villus composed of trophoblast cells with distinct large protoplasm.

protoplasmatic body (Fig. 32a). The trophoblast nuclei preserve their pronounced spheroidal form. The syncytial covering is at numerous points possessed of more than a single layer of nuclei and sends extensions into the trophoblast centre. Other villi composed of trophoblast nuclei possess a beginning centre of

mesoderm (Fig. 33). The syncytial covering contains vacuoles in the wall of which are crescent nuclei. A further stage is represented in villi whose mesodermal stroma contains but few trophoblast nuclei and where the syncytium, thin as endothelium, rests on no layer of Langhans (Fig. 34). In numerous villi, at one end is found the typical structure of a complete villus, the

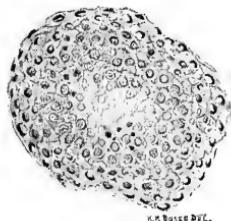


FIG. 33.—Villus with beginning centre of mesoderm. The syncytial covering contains vacuoles and crescent-shaped nuclei.

syncytial covering evidencing sprouts of a polynuclear character (Fig. 35). The other end of the villus is of a younger stage. The syncytium is there at the period characterized by the frothy, finely granular character of the protoplasm, while the nuclei are irregular in form and do not stain deeply (Fig. 35). Here, too, the change of trophoblast nuclei into syncytial ele-

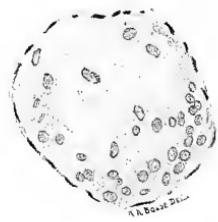


FIG. 34.—Older villus with thin syncytium and no layer of Langhans.

ments is evident. In the cell groups (Fig. 35*a*) we observe a covering of syncytium which at numerous points extends into the substance of the cell group, forming protoplasmatic masses of a thin endothelial character or of a polynuclear nature, and which divide the cell groups into irregular fields (Fig. 35*b*) wherein may be recognized the various early stages of subsequent villi (Fig. 35*c*). This invasion of syncytial cells is not alone an active

growth, but is due to the infiltration of a cell group by blood, which transforms the cells with which it comes into contact into syncytium. Here most distinctly can the gradual transition

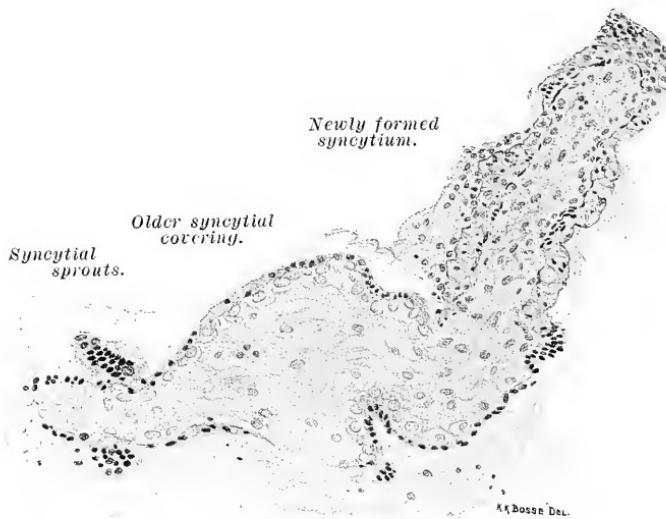


FIG. 35.—Villus showing an older stage in the left half and a younger stage in the right half.



FIG. 35a.—Cell group with syncytial elements invading it at various points.

from trophoblast to syncytium be observed. The large, poly-nuclear groups and knobs of syncytium, the so-called giant cells, are found at all points in the intervillous space. The nuclei



FIG. 35b.—Further stage of Fig. 35a. Cell group divided by syncytial elements, with the formation of fields which represent future villi.



FIG. 35c.—A further stage of Fig. 35b. Cell group divided into syncytial masses and villi. V_2 , villus in whose stroma are nucleated red blood cells; S , a syncytial mass; T , syncytial nuclei resulting from changes in the trophoblast cells; $Tr.$, trophoblast cells with beginning change to syncytial nuclei; V_1 , villus with whose syncytial covering numerous bridges of syncytium are connected.

are round, flat, and crescent shape. They are very frequently attached to the villi by larger or thinner pedicles. They are frequently found at the tips of the villi, where they represent the change of the small so-called cell groups, or pillars, to syncytium. *The villi are, then, in their various stages simply reproductions of what is found in the membrana chorii.*

CHAPTER IX.

THE MEMBRANA CHORII.

The great growth of the ectodermal cover in the ova of animals has been made known to us by Van Beneden, Duval, Nolf, and especially by Hubrecht, who gave it the name trophoblast.

In 1889 Hubrecht, in examining the placenta of the hedgehog, found a decided resemblance in numerous points to the human placenta. He asked then that the following questions with regard to the human ova should be studied:

1. Can, in the earliest stages of human ova, evidences be found of a trophospheral tissue between the villous covered ovum and the inner surface of the decidua reflexa?
2. Are there in this tissue blood vessels which receive their supply from those blood vessels which run in the reflexa?
3. Can evidences of a trophospheral tissue be found in the serotinal region?

The discussion of the histology of gestation in both uterus and tube shows that the various processes in the development of the human placenta resemble these observed in animal placentation.

It must be noted that these processes follow the steps observed by Hubrecht, Maximow, Opitz, etc. Yet there are still many diverging views as to the origin of the syncytial covering of the villi, as has been also observed in this regard in the investigations in animals.

Selenka and Strahl, in their examination of young human ova, find their views, gained in the examination of animal placentæ, sustained—namely, that the syncytial layer originates from the uterine epithelium. With this view Kossmann and Merttens agree. Eckardt finds that the syncytium originates from the maternal endothelium. Spee states that the syncytium develops from the connective-tissue cells of the decidua. Langhans considers the syncytial layer ectodermal, and the inner layer of Langhans mesodermal. Leopold and Gaisser consider the layer of Langhans to be of mesodermal origin *because they find these cells in the mesoderm of the villi*. Fränkel is uncertain as to the origin of the syncytium in human ova, and *finds transitions between*

the cell of Langhans and cells situated in the mesoderm of the villi. Kastschenko, Minot, Van Heukelom, and Ulesko-Stroganova consider both layers to be of fetal origin.

Eckardt, who holds that the syncytium originates from the maternal endothelium (?), states that the capillaries of the decidua which form lacunæ under the surface have a varying character. Many possess a distinct growth of the wall elements, which is often so great that he would consider these areas to be sections of glands, were it not for the blood therein contained. We know that, in the decidua menstrualis and graviditatis, blood may enter the glands between the epithelial cells or may break into the glands. The epithelium, which, especially in the neighborhood of the ovum, grows distinctly and shows an enlarged elongated protoplasm, *resembles to a certain degree the syncytium.* These cells, however, degenerate through the action of the blood and the glands are destroyed or pushed aside. From his description it is quite evident that Eckardt was describing such areas, which he mistook for a growth of maternal endothelium. With these remarks the views of Eckardt may be dismissed.

Merttens found in the particles obtained by curetting sixteen days after the last menstruation a few sections of an ovum with the enveloping decidua. He describes (1) the chorion and the villi. On the latter are cell groups, and between them are found spaces constituting the intervillous space. (2) The spongiosa, containing numerous glands, and (3) the compacta, a pale zone lying between 1 and 2 and containing large pale cells, between which are numerous darkly stained nuclei.

In the compacta he finds *large cells and large spaces.* Between the large cells *syncytial masses are irregularly distributed.* The cell spaces are round, long, or irregular, and often empty into the intervillous space. They contain blood and are lined with syncytial masses, which at some points are very thin and at other points quite thick. Where these spaces empty into the intervillous space the syncytial masses continue into the syncytium of the villi. Merttens does not know whether these spaces are glands, vessels, or lymph spaces. They are possibly, however, the lacunæ of Hubrecht, Peters, and Opitz.

The epithelia of the glands are like beaker cells, and on their free tips is a half-moon of glycogen. The glands show papillary projections (compare Fig. 14b). Merttens believes that the syncytium results through a change of the surface and gland epithelium (?). In his Fig. 8 Merttens represents, in the upper

portion, the epithelium of such a gland, and the lower half represents syncytial masses. He believes that the latter result from these epithelial cells, which lose their cylindrical form and unite. From his descriptions and his drawings it is neither clear nor probable that the epithelium changes into final syncytial masses. Besides, this area is taken from decidua particles not connected with the ovum, and at no point can he find the *epithelium of the glands contributing in any way to the syncytium of the chorion.*

On the contrary, this ovum is at the stage where the trophoblast is almost consumed by the growth of the intervillous space, and the cell groups at the end of the villi, which Merttens calls points of union between the villi and the decidua, are really the remains of the trophoblast.

In a very early ovum described by Spee the villi are covered with two layers. The inner, the cell layer of Langhans, he attributes to the ectoderm. The outer layer, *the syncytium or adventitia of the ovum*, he describes as follows: "This layer is often lifted off from the chorion ectoderm. It appears as a continuous thin streak of protoplasm with more or less closely grouped nuclei arranged in a single row. In the layers of the enveloping zone lying near the ovum are found cells which, through a diffuse staining of the nucleus and through the vacuolar character of the protoplasm, resemble the cell elements of the syncytium very much. In the space between the serotina and the ovum (intervillous space) we find them often close together and not distinctly separated, so that here the early stages of the polynuclear protoplasmatic groups are present. Such groups are also found united to the membrana chorii. Concerning their origin opinions differ. I have never observed a mitosis or an amitosis in their nuclei. The idea that the chorion ectoderm furnishes these masses may be set aside because of the sharp division between these two layers by a cuticula. These conditions prove that the syncytial formation does not increase through a division of the nucleus or the cell in the intervillous space. Since, however, an increase in the amount of the syncytial substance occurs within the intervillous space, this can only be explained through a wandering of already finished masses from the connective tissue of the compacta into the ovum.¹ Epithelial cells cannot be considered as the source, since the maternal cavity surrounding the ovum, even in the earliest stages, shows

¹Symplasma.

no epithelial lining, although in the later stages closely pressed glands are present all about the ovum and their epithelium is preserved at least up to the end of the first month.” It may be seen that Spee left out of consideration any possible action of the blood upon the ectoderm cells as an aid to the formation of syncytium. His decidua cells, resembling syncytium, are possibly changed trophoblast cells¹. *Cells from without* probably form the syncytial covering of the villi and of the membrana chorii, in his opinion. We think they are trophoblast cells of

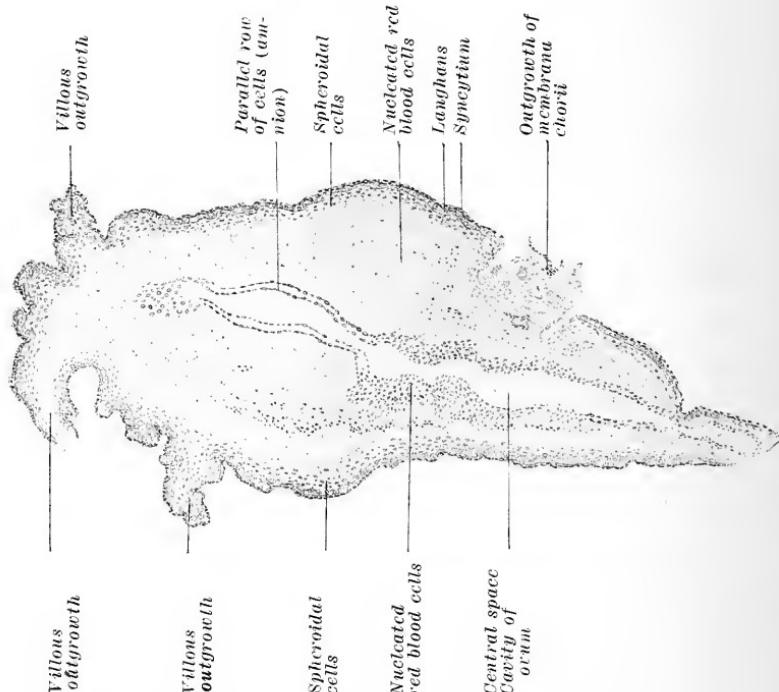


FIG. 36.—Transverse section through the fetal sac or membrana chorii of a four- to five-weeks ovum.

fetal ectodermal origin, acted on by the blood—a possibility which he left out of consideration.

Fig. 36 represents a transverse section through the greatest diameter of the ovum loosely connected with the decidua in the fourth or fifth week of uterine gestation. The centre is a space lined with a double parallel layer of cells arranged in single file. This also loses its parallel arrangement at certain points, where cells of the same character are found in groups several layers deep.

¹The resemblance of changed decidua cells to the future syncytium is often striking. See foot note, p. 87.

External to these cells, in the entire circumference of the sac, is a substance staining a deeper red and composed of extremely fine, granular, and thin-fibred, cotton-like material. In it, at numerous points, are isolated and grouped cells which look like nucleated red blood cells.

The next stratum is formed of round and oval cells, of the same character as the lining of the central space, but arranged in several somewhat parallel layers. Between these cells is a fine fibre-like substance, like the one just described, but taking a lighter red stain. The outer cells are rounder and more spherical than the inner, and the outermost cells, with a clear protoplasm and distinct nucleus, form a single layer of distinct cells with distinct cell boundaries (36).

The extreme external covering of the membrana chorii is of a

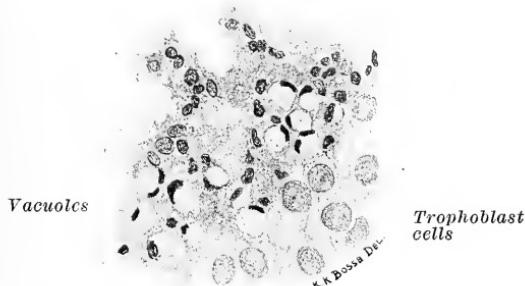


FIG. 37.—Vacuoles representing body of trophoblast cells, while the nucleus is a crescent in the circumference. From the membrana chorii of Fig. 36.

plasmodial character, and contains nuclei of various forms and evidences many vacuoles (Fig. 37). Each vacuole represents the space filled by a trophoblast cell, the nucleus being compressed at one point into a small crescent. At certain points outgrowths are present which contain larger and smaller areas of granular and plasmodial character, the aforementioned distinct cells, blood, and protoplasmatic groups containing small pale cells, small dark nuclei, and nuclei undergoing degeneration. Attached to the membrana chorii and all about it are innumerable villi in all stages of growth, young and old. The older villi present the same structure and possess the same cells, including the nucleated red blood cells, as are found in the chorionic membrane.

The conclusion gained from a comparison of this specimen with the structure of the villi is the following: The single

layer of cells immediately under the plasmodium of the chorioic membrane, the spheroidal cells under it, the distinct cells found in the outgrowths, the spheroidal cells in the young villi and in the cell groups which form the new villi, and the cell layer of Langhans, are identical and represent the ectodermal trophoblast cells. The villi are simply excrescences of the membrana chorii.

When the blood comes in contact with the trophoblast cells it does not coagulate. It does, however, exert a decided influence on the nuclei. Strahl found, in examining the placenta of Galago, that the blood extravasated from the vessels of the uterus is made use of by the fetus, in that the degenerating products of the extravasated blood cells are taken up and absorbed. These products are found in the form of larger and smaller yellow granules in the gland epithelium, which the latter uses in furnishing an iron-containing gland secretion. In the placenta of other animals the blood extravasated from the vessels of the uterus is taken up and absorbed by the ectoderm cells, and in certain animals the fetus obtains its nutrition because the ectoderm, covering the villus, takes up the substance given off by the uterine epithelium. Merttens observes that Lieberkühn in the placenta of the dog, and Strahl in the placenta of the mole, find ectoderm cells taking up and absorbing red blood cells. Peters states that Tafani, in the placenta of the cat, found chorion ectoderm cells outside of the placenta taking up red blood cells. We find much blood—that is, red blood cells—in the syncytium. It is also found in the vacuoles. Peters believes that the blood takes part in the changes of the trophoblast nuclei and that its elements contribute to the formation of the protoplasmatic masses known as the syncytium. "Not only the blood plasma, but also the nuclei of red and white blood cells, pass into the composition of these masses." It may be seen in Fig. 31b that the red blood cells change into a detritus and combine, so that finally small irregular areas containing small nuclear remnants result. A gradual destruction of red blood cells takes place, and the presence of leucocyte nuclei may be observed. The plasma, when acting suddenly and in large amounts, causes the trophoblast nuclei to shrink and take on a dark stain. When acting slowly it disintegrates them, flattens them, so that in a resulting homogeneous protoplasm nuclei of all forms may be found. It is evident, then, that the protoplasm of the syncytium is the product of the blood cells, the blood

plasma, and the protoplasm of the trophoblast cells. *The nuclei of the syncytium are the changed trophoblast nuclei.*

Wherever blood comes in contact with fetal cells syncytium results, whether in the decidua or in the intervillous space, so that *syncytium really plays the part of an endothelium* and at all periods, especially in the later stages, resembles endothelium so closely that it was naturally mistaken for it in nearly all the earlier investigations.

Many observers have noted a brush-like covering on the syncytium, though I have never found it. This striated surface on the syncytial covering of the chorion, and often observed on the isolated polynuclear protoplasmatic masses, appears to Spee to be a sort of fibrillation on the cell protoplasm. He believes that it originates through the influence of a sap current toward the ovum, because such a brush-like fibrillation is found in those places where a regulated transit of products through cells occurs, as in parts of the excretory ducts of the salivary glands and the kidneys on the side toward the connective tissue, and also perhaps in the osteoblasts in the Howship lacunæ of bone.

CHAPTER X.

THE BLOOD-FORMING FUNCTION OF THE TROPHOBlast.

In another particular, as I believe can be shown, the resemblance between the processes of development in the human being and those in animals is carried out still further—namely, the formation of blood by the fetal placenta.

In sections through the umbilical vesicle of *Tarsius* and *Tupaja*, Hubrecht finds polynuclear cells which he considers to be mother blood cells. Their nuclei result through fragmentation of a large nucleus. The individual nuclei within the mother cell then become surrounded with a special circle of plasma. When the original mantle of the protoplasm of the large cell is lost, the included cells are freed, forming embryonal nucleated red blood cells. During their circulation in the embryonal fetus the nuclear membrane disappears and the nucleus changes to a drop in which the chromatin diminishes, while the protoplasm begins to resemble the former nucleus. The cells become gradually smaller, and the final forms do not result from the protoplasm but from the nuclei of the cells. The blood cells originate, then, from the original nucleus of a mother blood cell. Hubrecht finds the same steps in the placenta of *Tarsius* as in the umbilical vesicle. He finds the same to occur in the trophoblast and in the maternal spongia, and even in the vessels which lie between the glands. In the placenta of *Tupaja* the same steps are noted, but, in addition, the epithelium, as well as the connective tissue of the decidua, is the seat of red blood cells derived from nuclei.

In his ovum *v.H.* Spee describes the mesoderm layer as follows: He finds two different cell forms occurring side by side and intermingled: (1) Branched cells, extending into fine threads which project in all directions and which may well be considered connective-tissue fibres. Where these cells occur singly—for instance, at certain points in the abdominal pedicle and in the villi—the mesoblast appears exceedingly poor in nuclei. Strongly-stained specimens are necessary to see the numerous fine,

wavy, and sharply contoured fibres of the embryonal connective tissue. (2) Between these structures are found larger, strongly prominent spindle cells with oval nuclei, arranged sometimes closely together like bundles, and at other points separated from each other by wide spaces without evident contents. The course of these fibres follows, with noticeable constancy, the curves of the free mesoderm surface—that is, the surface covered with epithelium.

In our opinion many of the second form of cells are possibly ectodermal. In an ovum described by Reichert, on the inner side of the chorion is a substance which he considers to be a mucoid-like deposit and which also extends into the villi. The cavity of the ovum is likewise filled with this substance. Spee considers this to be mesoderm tissue, although Reichert describes the contents of the ovum simply as a mucoid substance without cells. Spee says that the contents of the amnion and of the umbilical vesicle of ova hardened in alcohol never coagulate, but such a condition does take place in the wide mesoderm slit of dead young human ova.

Thus the early embryonal mesoderm tissue of the membrana chorii and of the villi is very poor in cells, and the description of Reichert, characterized by Spee as mesoderm substance, serves to characterize as mesoderm the non-celled material found between the central space and the external layers of cells seen in Fig. 36 and also found in many of the younger villi.

It has been mentioned that Langhans considers the cell layer to be of mesodermal origin, and that Leopold holds the same view because he finds *transitions between the stroma cells and the cell layer*. Fränkel, who believes that the cell layer and the syncytium are probably identical, finds the question difficult of solution *because of the numerous villi whose interior is composed of ectodermal trophoblast cells*. We have called attention to this occurrence as a frequent one, and have likewise observed the resemblance between the cell layer of the membrana chorii and the several layers of cells under it which are embedded, however, in mesoderm substance (Fig. 36). The origin of the villi, the fact that they are formed out of solid trophoblast groups, the fact that only later does mesodermal tissue reduce them to the generally single layer of Langhans, the fact that in this reduction to a single layer many trophoblast cells are left embedded in the stroma of the villi and the membrana chorii, and the numerous transitions between these cells and the cells of

Langhans, prove that they are ectodermal cells situated in a basis of mesoderm substance in which only later mesoderm nuclei appear.

As the villi grow older, these ectoderm cells are crowded by the growing mesoderm tissue, so that some degenerate, while others take part in the following important changes—that is, **THEY FORM RED BLOOD CELLS.**

In the same manner that the external trophoblast cells, which come in contact with the blood, obtain from it a protoplasmatic envelope, while they themselves form the nuclei, in quite the same manner these cells situated in mesoderm are seen to gradually change their form, become darker, become surrounded by a red-staining granular protoplasm of the same character as the mesodermal tissue. They then represent nucleated red blood cells. They are often seen in stages where the nucleus becomes fragmented or divided into other cells. These larger and smaller nucleated reds are found isolated or in groups, often lying in spaces or slits of the mesoderm of the villi and membrana chorii. These spaces are often surrounded by very long, very dark, and spindle-like cells, evidently forming the endothelium of the future capillaries.

We have observed the statement that in animals the mesoderm, which enters the trophoblast groups and forms the stroma of the villi, carries with it the branches of the allantoic vessels. Vessels, however, are not present in the villi of the human ovum until the third week, and the villi are then filled with mesoderm or ectoderm cells, and it would be impossible for extensions of the allantoic vessels to enter into the innumerable extensions of the membrana chorii constituting the villi. The process followed, however, is probably the following:

Mesoderm does not enter as a distinct tissue into the trophoblast elements *after they have been reduced by the maternal blood to syncytium*, but is present between the trophoblast cells *from the very earliest period*, as may be seen in the description of a tubal ovum. When, therefore, a trophoblast cell group comes in contact with maternal blood, the outer cells are changed to syncytium, underneath which results the cell layer of Langhans, *which in the still later periods disappears*. The mesoderm substance, when increasing in amount, dilates the villus and forms the stroma. In the resulting stroma are left the remaining more or less scattered trophoblast cells. From these, nucleated red blood cells are formed, which lie in small slits or

spaces, which spaces become lined with endothelium, thus forming capillaries. These capillaries gradually increase and finally become very numerous, having a tendency to lie close under the epithelial covering of the villi. Whether any of these trophoblast cells take part in the formation of the endothelium, it is impossible to say with certainty. These same processes are observed in the membrana chorii. These capillaries in the villi and in the membrana chorii later unite with the umbilical vessels which make their way through the abdominal pedicle.

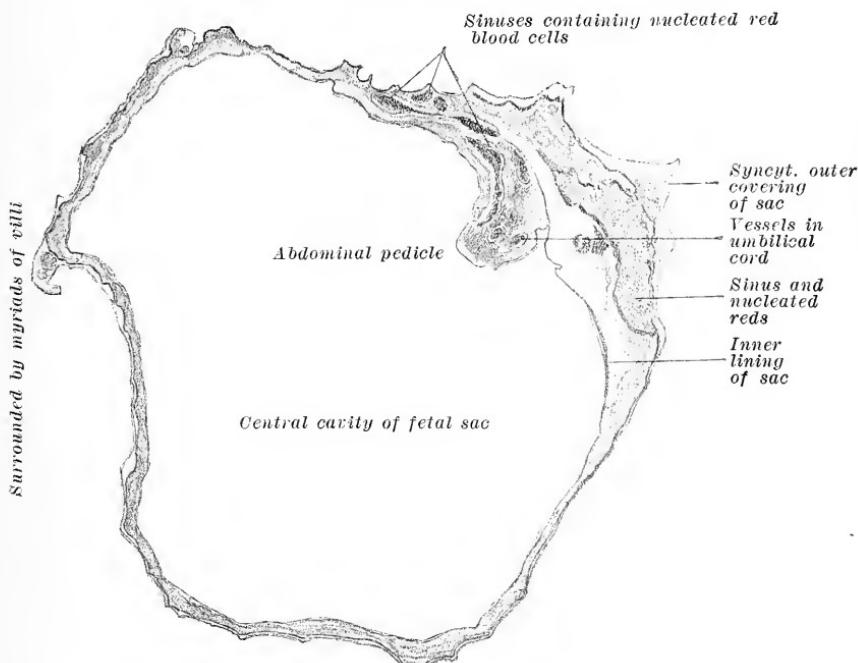


FIG. 38.—Membrana chorii of tubal ovum.

The nucleated red blood cells, in their subsequent circulation through the fetus, become changed, and from the third month on ordinary red blood cells are found. In the membrana chorii of our uterine ovum the same stages of capillary development may be observed, but to a less extent than in some of the villi which already possess numerous and distinct capillaries. In the later stages this process is more marked in the membrana chorii than in the villi, as may be seen from the following description of a tubal gestation with well-preserved fetus:¹

¹The possibility that these nucleated red blood cells came from the fetus was not left out of consideration.

The fetus was lying in the central cavity of the ovum (Fig. 38) attached by an umbilical pedicle which entered the placenta at the site to be mentioned later. The fetus was one centimetre long; the abdominal wall was not yet closed, and the arm and leg formations were just evident as small pinhead knobs. The cavity of the ovum is lined by a membrane composed of flattened cells with distinct nuclei. At various points in the circumference of the membrana chorii, and especially at the point where the abdominal pedicle is attached, are numerous larger or smaller spaces, separated, sometimes, from the maternal blood by only a single or double layer of cells, which at many points are of a distinctly plasmodial character. The membrana chorii has at various points, and especially at the placental site, numerous villi attached to it and numerous small projections of the same structure as villi. These spaces, or sinuses, contain masses of round, red-staining protoplasmatic masses with dark central nuclei, many of which are undergoing division—nucleated red blood cells. It seems possible that many of these sinuses or spaces represent dilatations of the parallel row of cells observed in Fig. 36, wherein were noted, at various points, groups of cells of a trophoblast character. It is probable that those groups too form the nucleated red blood cells observed in these sinuses.

The largest of these sinuses are present near the insertion of the abdominal pedicle, and all of them are separated from the maternal blood of the intervillous space, often by only a single layer. These sinuses may represent, then, areas in the membrana chorii filled with nucleated red blood cells of placental trophoblast origin, which subsequently enter the fetal circulation.

In the pedicle itself a change to the subsequent non-nucleated red blood cells may be observed, for in its substance are fetal vessels and sinuses filled with cells of a different character. They are small, with a thin circumference of protoplasm and a dark nucleus filling out almost the entire cell body. These represent, then, the later stages of the red blood cells.

The resemblance of this process to the one noted by Hubrecht in *Tupaja* and *Tarsius* carries out still further the resemblance of the various processes in the human placenta to those in the placenta of animals. That this theory and explanation, whereby red blood cells are formed of trophoblast ectodermal cells, is a rational one and proven by the examination of our specimens seems to me evident. In addition, it serves to explain the presence of trophoblast cells *in the interior of the villi*—a fact which

has led many, who have considered them mesodermal because situated in the mesodermal stroma, to likewise call the cells of Langhans mesodermal. The formation of nucleated red blood cells from trophoblast cells, on the addition of a protoplasm of mesodermal origin, would serve to make their resemblance to syncytium very great; for the latter is composed of trophoblast nuclei with a protoplasm largely composed of maternal blood plasma. If, as many sections have suggested, the capillaries were lined by endothelium which is also formed of these trophoblast nuclei, the resemblance of this process to the formation of syncytium would be further increased; for the syncytium simply plays the part of endothelium, separating the other fetal cells at all times from the circulating blood.

Uterine (as well as tubal) ova furnish us with the following positive conclusions:

1. *The human ovum possesses an ectodermal growth of cells, the trophoblast, consisting of closely-grouped cells.*
2. *When vascularized by maternal blood, a second external layer, consisting of plasmodial mononuclear and polynuclear elements, results.*
3. *Elements of the maternal blood circulating in the spaces and lacunae of the trophoblast contribute to the protoplasm of the syncytium. Among other elements, the secretion of the uterine or tubal epithelium may likewise contribute to the formation of the syncytial protoplasm. At any rate, much of the protoplasm (but none of the nuclei) is of maternal origin.*
4. *On the villi and the membrana chorii the plasmodial cells form the outer syncytial layer, while the closely-grouped cells beneath it furnish the single layer of Langhans.*
5. *The stroma of the chorionic villi is formed of mesodermal tissue in which are later found capillaries communicating with the umbilical vessels and containing fetal blood. Clear in almost every detail, then, a trophoblast formation, consisting of an inner layer of separated cells and an outer or plasmodial layer such as is found in the placental development of animals, is present in human placentation.*

NOTE.—It is to be noted that the embedding of the guinea-pig's ovum reveals symplasmatic changes in the decidua which make Spee's theory of the maternal origin of the syncytium very attractive. Certain it is that the decidual cells undergo "syncytial changes." Were we to go but one step further and add to the maternal blood symplasmatic decidual structures as an external factor in the production of syncytium, then

the theory of the exclusively fetal trophoblastic origin of this tissue would fall to the ground. The distinction between fetal and maternal cells is often decidedly difficult, and in that fact lies the crux of the problem. If the view we have chosen be correct, it must still be granted that the decidua undergoes primary or secondary changes which make its resemblance to real syncytium striking. Though often and in many specimens led to favor Spee's view by microscopic appearances, yet our choice of views (by no means surely excluding, in whole or in part, the views of Spee) seems well founded.

"Structures not wisely called 'syncytial' may be formed from various forms of tissue, as uterine epithelium, uterine connective tissue, fetal ectoblast, etc. Therefore, especially when we consider the variations in placental formation in various species of animals, it is not correct to speak of 'the syncytium' as if all syncytial formations were alike. Therefore, in the guinea-pig, I did not use this word, but chose the term 'symplasma' to designate those structures (developed from connective tissue) comparable to syncytium" (v. Spee). Van Beneden calls this tissue, developed from ectoblast in animals, plasmodium. Plasmodium is, then, in the human ovum "trophoblast." We believe "syncytium" to be the "plasmodium" of animals. Decidual and other uterine cells undergo the so-called "syncytial changes." It would be advisable to call these "symplastic" cells. The question then is: Is syncytium of symplastic or plasmodial origin? We hold the latter view.

CHAPTER XI.

THE FURTHER DEVELOPMENT OF THE UTERINE PLACENTA.

Just as during the early weeks the trophoblast invades the trophospongia or decidua, so after the formation of villi is the further course of the ectodermal trophoblast and syncytial cells of a destructive character, so far as the decidua is concerned. Fig. 39 presents a fetal sac *in situ* in the sixth or seventh week of

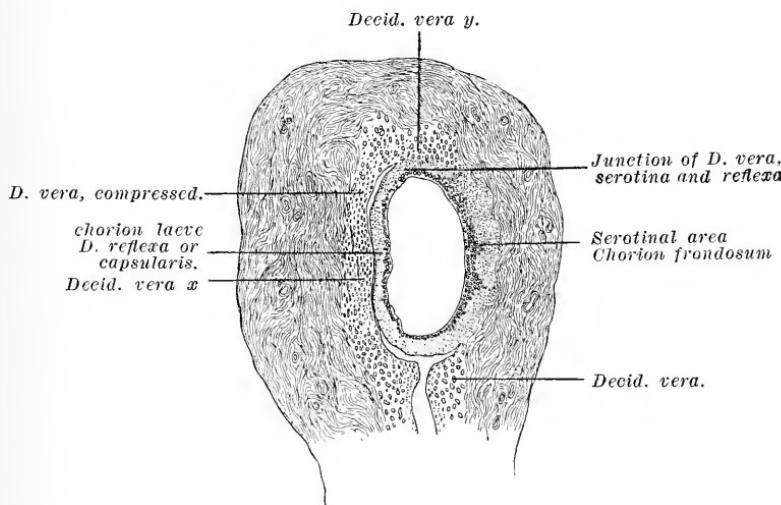


FIG. 39.—Uterus and fetal sac *in situ*—7 weeks—two-thirds the size of the specimen when mounted, and one-half the size of the specimen before hardening. *y*, decidua vera with numerous glands, i.e., well-marked spongiosa; *x*, decidua vera with compacta compressed through pressure of the sac, and evidencing few flattened glands in the spongiosa.

uterine gestation. The specimen from which the drawing was constructed was unfortunately distorted, yet the important relations are very clear.

The capsularis, to which villi are attached, is full of darkly-staining trophoblast cells, between which and through which are blood spaces and blood extravasations. Opposite it, but not yet in contact, is the decidua vera, flattened by pressure

of the growing sac, and composed of pure decidua divided into a superficial layer, the compacta, and a deeper layer of flattened glands, the spongiosa (Fig. 40). Where the decidua vera is not compressed, as at the areas where decidua vera, reflexa, and serotina join (Fig. 39), the decidua retains in its deeper layer the typical character of the spongiosa, evidencing numerous glands of all sizes and lined by preserved epithelium of a "syncytial" character (Fig. 14b).



FIG. 40.—High-power drawing of decidua vera (compressed) of Fig. 39.

The decidua serotina evidences in the deeper layers few flattened glands. The superficial layers are greatly changed. Where the villi come in contact with decidua the syncytium disappears and *the cell groups enter the maternal tissue and mingle with it* (Fig. 41). This process is carried on, then, by all the attached villi also, so that the resulting "decidua" is of the same character as the cell groups. This resulting tissue resembles the decidua, but its nuclei take a deeper stain and it evidences no small-celled infiltration. In it are found cells composed of a

structureless mass enclosing one or more cell nuclei or groups of nuclei. They resemble syncytium and are simply changed fetal

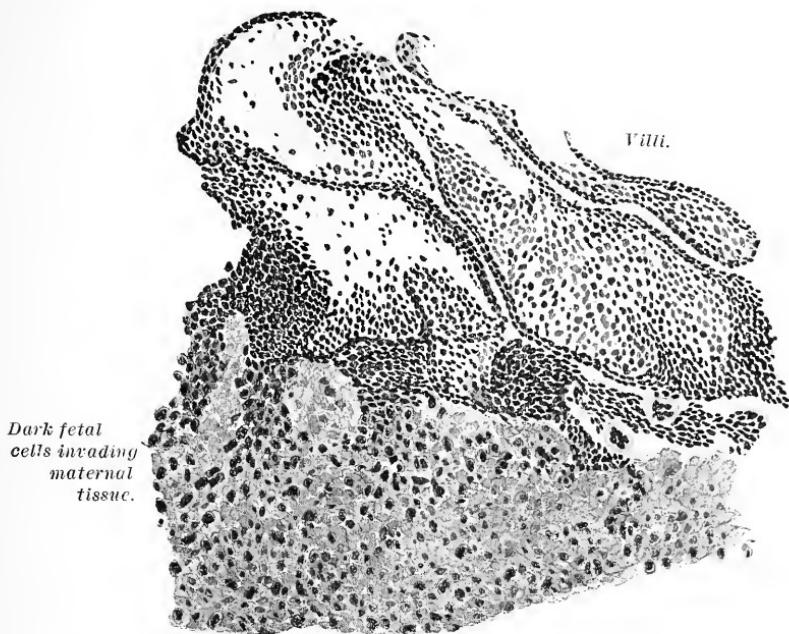


FIG. 41.—Cell groups of villi entering maternal tissue. Taken from the sero-tinal or placental area of Fig. 39.

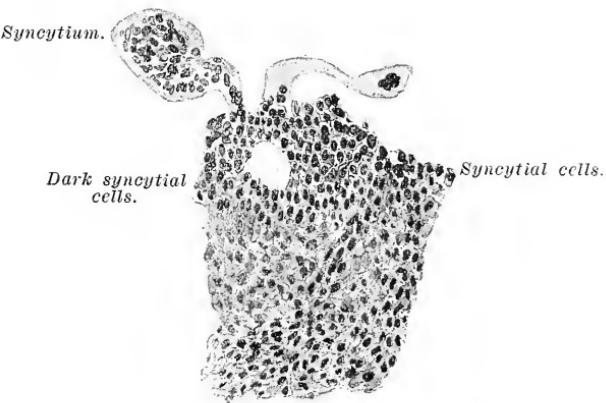


FIG. 42.—Dark syncytial cells infiltrating decidua serotina of Fig. 39.

cells. Without the aid of these syncytial cells, which infiltrate the maternal tissue very thoroughly and in all directions and quite deeply, it is difficult, with the exception of the darker

staining, to mention characteristics which to the beginner would differentiate fetal cells from the decidua. *It is this fact which has made the various questions depending on this differentiation difficult of solution.*

The boundary between fetal and decidua cells shows degenerative changes; the superficial glands are pushed aside and destroyed, while the deeper glands grow, their epithelium swells, unites and forms so-called syncytial changes. These masses fall off and finally degenerate. *The fetal cells at all periods infiltrate the decidua and bring it to destruction.* The advancing ectoblastic and syncytial cells have an erosive action on the vessels which prepares them and the capillaries for bursting. In this way new lacunae and areas of degeneration result, and the blood from these open vessels changes the fetal cells to syncytium and to villi and constantly increases the extent of the intervillous space.

The cell groups of the villi later diminish and disappear. The villi increase in number and are closely grouped. Their stroma consists of star-shaped cells, between which are cells resembling those of Langhans. The rest of the stroma is filled out with numerous capillaries, and finally the syncytium is reduced to a thin substance containing nuclei (Fig. 44).

CHAPTER XII.

THE PLACENTA.

That part of the decidua covering the embedded ovum, plus the scar of Reichert, forms the capsularis, or *decidua reflexa*. That part of the decidua in which the ovum is embedded, and on which the ovum rests in its early stages, is the *serotina*. Here the main changes between the ovum and villi on the one hand, and the decidua and maternal blood on the other hand, take place. The remainder of the decidua is the *vera*.

The villi in the entire periphery of the ovum depend for their growth and development on connection with the maternal decidua and blood. At the end of the first month the villi are evenly distributed over most of the periphery of the ovum. With the growth of the ovum in the next two months there is a stretching of the capsularis, which, however, grows likewise, so that it is not much thinned except at its summit. There is, however, in these two months a constantly slighter connection of the vessels of the capsularis with the maternal vessels of the vera and serotina. In addition the fetal vessels which enter the *membrana chorii* through the adherent band establish closer and more immediate relations with the villi at the serotina, for the abdominal pedicle is at this point.

As a result we distinguish two divisions in the chorion : (1) that in which new villi cease to develop and in which formed villi gradually degenerate and atrophy—the *chorion laeve*; and (2) that in which the villi increase hugely in number and size, the area at the serotina—the *chorion frondosum*.

The chorion frondosum forms the placenta. The growth of the uterus in the first three months is independent of the growth of the ovum, and the uterus undergoes eccentric development. Therefore the area of the serotina increases in extent. The ovum grows of its own accord, and as a result the area of the chorion frondosum *increases in extent* and keeps pace with the growth in extent of the serotina. Septa of decidua extend between villi and groups of villi, due to the irregular invasion of the serotina.

As the ovum grows in the first three months it is covered by

capsularis (Fig. 39). Though the *capsularis* at the end of the third month covers an ovum as large as a goose egg, it is fairly thick except at its summit, and is in contact with the *decidua vera* in its entire circumference. During the fourth month the *capsularis* unites at all points to the *vera* and its nourishment ceases except at its *base*. The area where *capsularis* at its base joins the *serotina* and *vera* is known as the *Border Zone* and constitutes the boundary of the "placental area" in the first three months. The base of the *capsularis* unites last with the *vera*. Septa of *serotinal* *decidua* extend between villi and groups of villi, especially at the point where *capsularis*, *serotina*, and *vera* join, and there lie close to the *membrana chorii*, forming a maternal border or margin to the placenta—*i.e.*, at the border zone the *decidua* is supposed to extend up to and touch the *membrana chorii* as Winkler's Plate.

In the early three months, *up to the formation of the placenta*, even though the uterus grows actively, the growing *chorion frondosum* and the base of the *chorion laeve* extend into *vera* and split it into two layers (?). The lower layer becomes *serotina* and the upper layer is added to the *capsularis*. From the end of the third month on, the uterus enlarges passively through growth of the ovum. It is scarcely possible that the edge of the growing placenta then likewise splits the *vera* into two layers to any extent, but in its growth it will extend, strictly speaking, not upon *vera*, but upon the base of the *capsularis* which has united to *vera*. The villi do not atrophy on every point of the *capsularis*. They may persist on the part of the *capsularis* nearest to the *vera* and *serotina*. When *capsularis* unites to *vera* these villi may grow through the united *capsularis* and *vera*. As a rule the loosest union between *capsularis* and *vera* is at this point and so the loosest union of villi and placenta to *decidua* is at this point.

Three groups are observed in mammalia, as regards the character of the chorion.

(1) The chorion is smooth and is attached to the uterine mucosa, which is rich in vessels. The chorion takes from the large epithelial cells of the uterus nourishment for the embryo.

(2) The chorion has projections or villi. An allantois approaches the chorion, and contained umbilical vessels enter the villi. The chorion and the mucosa are closely connected. In the pig, for instance, numerous projections, rich in vessels, fit into smooth depressions in the mucosa, in which open the uterine

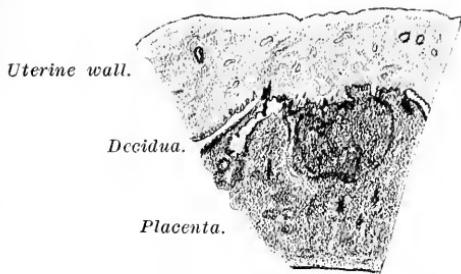


FIG. 43.—Low-pressure drawing of uterus and placenta at full term, showing the close but uneven attachment of the placenta, two-thirds of actual thickness. To the right the villi are on the muscularis. To the left (marked *Decidua*) there is spongiosa. In hardening a separation formed between compacta and spongiosa at this point.

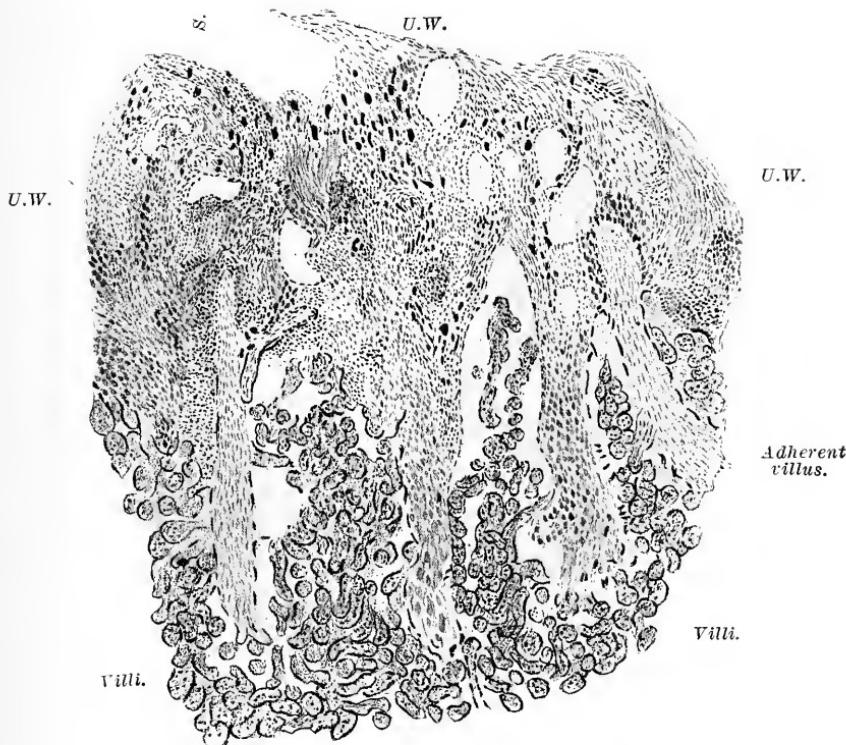


FIG. 44.—Utero-placental junction at full term. *S* and the other spaces are maternal vessels. Villi are projecting into maternal sinuses. *U.W.*, uterine wall infiltrated with fetal cells, especially with the very dark giant syncytial masses. Of the four septa between the villi, the two outer are "adherent villi," the two inner are decidual septa. Some of the sinuses are possibly glands with lost epithelium.

glands. In labor the villi and the uterine mucosa are separated without loss of mucosa substance.

(3) At one or more parts of the chorion a placenta develops composed of *placenta fetalis* and *placenta uterina*. In the sheep, cow, and ruminants a subdivision exists. Numerous small pla-

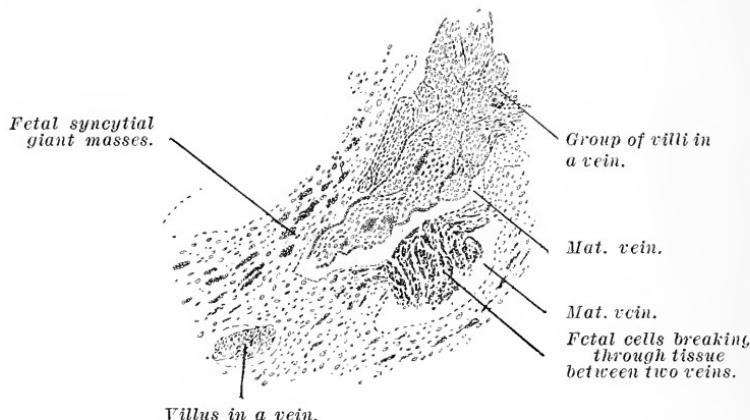


FIG. 45.—High-power drawing of an area in Fig. 44, showing the invasion of maternal tissue by fetal cells even at full term, and also the entrance of villi into the uterine veins through the blood current.

centæ or cotyledones are present. The villi are covered with flat cells and the depressions of mucosa are lined by cylindrical cells. The latter develop a so-called uterine milk for the nourishment of the fetus. All other mammalia with a placenta evidence a *very close connection between the maternal and fetal parts*, and



FIG. 46.—High-power drawing of placental villi of Fig. 44, showing the free syncytial masses and the chorionic syncytium to be identical with the syncytial giant masses of Fig. 45.

in labor a decidua is cast off. Here a syncytium forms the separation between the villi and the maternal blood.

The invading trophoblast and syncytial cells have a decided power of wandering. These wandering cells are often found singly, entering between the muscle and the connective-tissue bundles, in the lymph spaces and often in the blood vessels.

Through pressure they are often reduced to thin, long, spindle-shaped forms, and, when found, the muscle fibres between appear to be changed forms of the latter. In the earlier periods, round or polyhedral cells with very large, round or irregular, long, darkly-staining nuclei are found and have been generally mistaken for decidua cells. Their nuclei are often very large and on degeneration contain vacuoles.¹

In the specimens (Figs. 43, 44) on page 95 is seen a section through a placenta and the uterine wall at full term. The villi are very closely grouped. The uterine wall evidences large sinuses and is *thoroughly infiltrated with fetal cells and many giant syncytial masses¹ of the same character as the syncytial covering of the villi.* In the drawing may be seen four extensions between the uterine wall and the main mass of the placenta. Between the extensions loose and attached villi, as well as free syncytial masses, are present. The two middle extensions belong to the uterine wall, but they are thoroughly infiltrated with fetal cells. On their sides and at their tips villi are attached, sending at these points their cells in the uterine wall.

The two outer extensions are the *so-called adherent villi* firmly embedded in the uterine wall, and their tips are outlined by more closely grouped and more darkly stained ectodermal cells. It is from points like these that the ectodermal cells and the giant cells *invade most deeply the uterine wall*, as may be seen in Fig. 44. The spaces between these extensions represent the mouths of the maternal sinuses which send their blood into the intervillous space.

In the case of normal uterine gestation villi do not project into the arterial openings of the intervillous space. They do, however, project normally not only at the border sinuses, but at all the serotinal points into the veins alone. The villi are of normal structure and often so numerous that they occlude the vein lumina. They are frequently very long and follow the course of the veins for a considerable distance, so that in the uterus villi may be found in direct contact with the uterine muscle or, better, in the veins of the uterine muscle (Fig. 45).

From the very earliest period of trophoblastic development trophoblast cells may be carried into the maternal circulation through the blood lacunæ and the blood capillaries. In the latter stages the trophoblast cells invading the decidua, eroding the

¹Many consider these placental giant cells to be of decidual origin. So does Spee.

vessels, and destroying the decidua furnish still further opportunities for the entrance of fetal cells into the maternal circulation. When the intervillous space is well defined and is limited in its entire circumference by the point of union of the decidua vera and the capsularis (decidua reflexa), a still further opportunity is furnished to syncytial masses, cell groups, and even villi to be carried through the veins into the maternal circulation. In the still later periods and at full term, villi projecting into the maternal veins may be carried off, and the fetal cells which have invaded the decidua and the uterine wall, and which even up to the last days of gestation continue to enter the maternal vessels, may readily enter the maternal blood current. In eclampsia such elements have so frequently been found in the circulation, and especially in the lungs, that they have been considered the cause of this affection. They have, however, been proved to be rather the result than the cause, for Schmorl and others have found them in these locations in perfectly normal cases.

The important fact remains that from the very earliest moment, not only in the primary intervillous space but in the fully-formed intervillous space, as well as through the vessels of the uterine decidua and wall, fetal cells are continually entering the blood of the mother.

CHAPTER XIII.

THE UMBILICAL VESSELS AND CORD.

A. UMBILICAL VESSELS.

The ovum possesses its fluid centre or yolk. It may possess a covering of discus proligerus and albumin. On embedding the trophoblast digests the decidua cells and is surrounded by the so-called *symplesmatic* fluid. The ovum develops and the trophoblast grows. During this time osmosis and the yolk are sources of nourishment.

As the decidua is invaded capillaries are opened, and in the first few days trophoblast is acted on by blood of the mother. It changes trophoblast cells to syncytium and probably blood elements enter into the composition of the cells. Osmosis again serves to explain the growth of the ovum and its contents. On the interior of the ovum we have the embryo, consisting of ectoderm, mesoderm, and a lining of entoderm, which entodermal lining is a part of the entoderm of the yolk sac and abdominal vesicle. The ectoderm of the embryo, consisting of stratified epithelium, is continued into the single-layered amnion reflected over its dorsal aspect. The contents of the yolk sac thus line the entoderm of the embryo. A branch of the entoderm extends into the abdominal adherent band as the allantoic duct (Figs. 30d, 46b).

As the ventral and caudal ends curve anteriorly, and the lateral walls also, they constrict the abdominal vesicle. The connection between extra-abdominal umbilical vesicle and intra-abdominal is through a resulting narrow canal, the ductus omphalo-entericus (Fig. 46 a and b). The duct is called omphalo-entericus because meanwhile mesoderm and entoderm have formed intestine and the duct enters into intestine instead of into a flat entodermal lining of the embryo. All this while arteries and veins have developed on the abdominal vesicle, the arteriæ and venæ omphalo-mesentericæ. The arteries pass from the aorta to the vesicle, the veins from the vesicle to the venous end of the heart tube. In the first month develop two umbilical arteries and two umbilical veins. The arteries are branches of the aorta. They pass along the

future lateral pelvic walls, along the later bladder, along the later anterior abdominal wall to the adherent band of mesoderm. Two umbilical veins develop, one of which atrophies. Their contents at first enter through the duet of Cuvier into the heart.

The two *venae omphalo-mesentericae* pass along the intestine and anastomose at the duodenum. They send branches to the budding liver, the future hepatic veins. These pass into the end area of the *venae omphalo-mesentericae* which empties into the primitive heart. The liver grows and thus needs more blood. Then develop the umbilical veins, which pass through the duets of Cuvier to the venous sinus of the heart, passing over the liver. The right vein atrophies. Some branches of the left umbilical vein anastomose under the liver with the *venae omphalo-mesenteriae*. This anastomosis under the liver soon becomes the important part and finally takes all the placental blood to the liver and through the hepatic veins. Thus the umbilical blood circulates with the yolk blood through the liver and then through the yolk veins to the heart, for the end piece of the yolk veins becomes the cardiac end of the future inferior vena cava. At first the umbilical vessels are small, while the *omphalo-mesenteric* are large. As the yoke is sucked up the *omphalo-mesenteric* vessels atrophy and the umbilical grow. The branches of the latter extend on through the abdominal pedicle, through the mesoderm of the membrana chorii into the villi, and join the capillaries there. At the end of the first month the fetal heart begins to beat, and in the first month circulation through the vessels begins, and FOR THE FIRST TIME there is an exchange between maternal blood and the contents of the chorionic vessels through the syncytium and cells of Langhans. As the liver grows the yolk veins atrophy and the umbilical veins grow larger. When the liver can no longer take care of all the placental blood the *duetus venosus Aurantii* is formed, and so part of the blood goes directly into the inferior vena cava. The original yolk veins form the portal circulation, which becomes active in the latter part of the fetal nine months. After birth the portal circulation, carrying blood from the intestines, pancreas, and spleen, supplies the liver. The portal circulation, as said before, develops from the yolk veins. These changes are due to the change in size and importance of the yolk, placenta, and liver. After birth the umbilical vein becomes the ligamentum teres of the liver; the *duetus venosus* becomes the ligamentum venosum (Hertwig).

B. THE UMBILICAL CORD.

The ductus omphalo-entericus enters the future ileum at a point lying subsequently near the cecum. It is obliterated in the eighth week and disappears. Sometimes it is retained and is known as the diverticulum of Meckel. It may be simply a diverticulum of the ileum. It may extend up to the umbilicus as a patent canal extending from the ileum. It may remain as a band extending between the ileum and the umbilicus.

The umbilical artery of each side forms from its first part the common iliae. The remainder passing along the pelvic lateral wall to the side of the bladder, along the anterior surface of the abdominal wall to the adherent band and umbilicus, becomes the ligamentum vesico-umbilicale laterale. The intra-abdominal



FIG. 46a.—Embryo with abdominal mesodermal pedicle and with umbilical vesicle, showing amnion still close to the embryo, a condition still found in the third week or even later. (Ahlfeld.)

portion of the allantoic duct becomes a part of the bladder and the urachus, which extends from the summit of the bladder along the anterior abdominal wall to the navel.

Of the umbilical veins, one atrophies. Early in its career it joins the venae omphalo-mesentericæ, which, in their later stages, are connected with the liver, and thus it enters the liver and subsequently becomes the ligamentum teres. Coming out of the abdomen of the fetus are:

1. The adherent band of mesoderm.
2. In it the allantoic duct.
3. In it the umbilical vein and umbilical arteries.
4. The atrophied umbilical vesicle and its duct, the omphalo-enteric duct.
5. On the latter the atrophied omphalo-mesenteric vessels.

C. AMNIOTIC.

The amnion, even in the third week, may lie close upon the fetus. In it fluid is secreted which constantly distends it. As the cephalic and caudal ends take on a ventral curve, and as the lateral walls of the germinal plate curve upward and gradually approach, and as the points from which amnion begins thus approach, the amnion comes nearer and closer to the abdominal pedicle and the omphalo-enteric duct. When the amnion is finally distended so that it is in contact with the inner lining of the membrana chorii (6th-10th week), it must envelop the abdominal pedicle and the atrophied umbilical vesicle. The vesicle

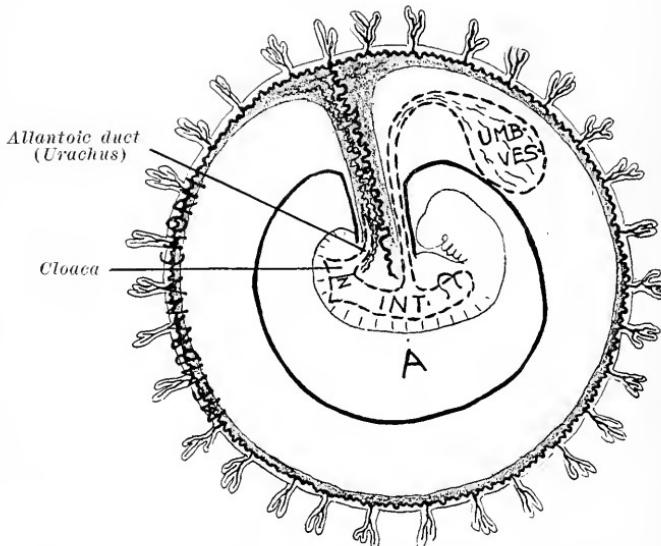


FIG. 46b.—Ovum (schematic), showing extra-embryonal chorion connected with the embryonal area by the mesoblastic umbilical pedicle. In the umbilical pedicle are the allantoic duct and the umbilical vessels. Passing from the umbilical vesicle to the intestine are the omphalo-enteric duct and its obliterated vessels. When the amnion becomes large enough to line the membrana chorii (sixth week or later), it presses these structures together and surrounds the then-formed umbilical cord.

is longer than the abdominal pedicle, so that its tip lies for a distance on the fetal surface of the membrana chorii. The point of attachment of the original adherent band to the membrana chorii is the point of attachment of the abdominal pedicle to the chorion frondosum and the point of attachment of the umbilical cord to the placenta. At the umbilicus the amnion, which thus covers the cord, goes over into the abdominal epidermis. The main substance of the cord consists of Wharton's jelly. A long, thin cord is characteristic of man.

CHAPTER XIV.

GROSS ANATOMY OF THE PLACENTA.

The placenta is a thickly spongy, fairly circular mass measuring 15-20 centimetres in diameter and about 3-4 centimetres in thickness. The surface toward the embryo has attached to it the cord and is covered, or rather lined, with the amnion.

Placenta is the finally developed membrana chorii of the chorion frondosum with its myriads of chorionic villi. Instead of each villus being connected with the membrana chorii, we have connected with the latter large stems on which are numberless branches and sub-branches covered with chorionic villi.

The outer surface of the placenta is divided by furrows into areas or islands called cotyledones. A cotyledo consists of a stem passing out from the membrana chorii. From it pass villi and large branches covered with villi, and from these still smaller branches with villi. The uterine placental area has been invaded by trophoblast cells and villi during a period of nine months, so that compacta and almost all of the spongiosa have been "eaten up." If, from a certain area, we remove compacta and spongiosa, we pass from the region of capillaries toward the region of vessels situated in the muscularis. The uterus, however, has grown actively in the first three months. The vessels of such an area have hypertrophied during this time proportionately with the enlargement and hypertrophy of all the uterine structures. In the next six months the uterus enlarges passively and a further influence is exerted on the openings of the vessels on the uterine surface—*i.e.*, they are stretched and opened in trumpet fashion. As a result, we find on the uterine placental surface larger and smaller sinuses lined more or less with endothelium, which endothelium may extend in trumpet fashion to various degrees.

Except at the margin, practically no glands are found beneath the decidual projections. The remnant of decidua covering the uterine placental area is very thin and is so infiltrated with fetal trophoblast cells as to be practically fetal in character. The same is true of the decidual projections.

The sinuses are situated in hollows or depressions, for the

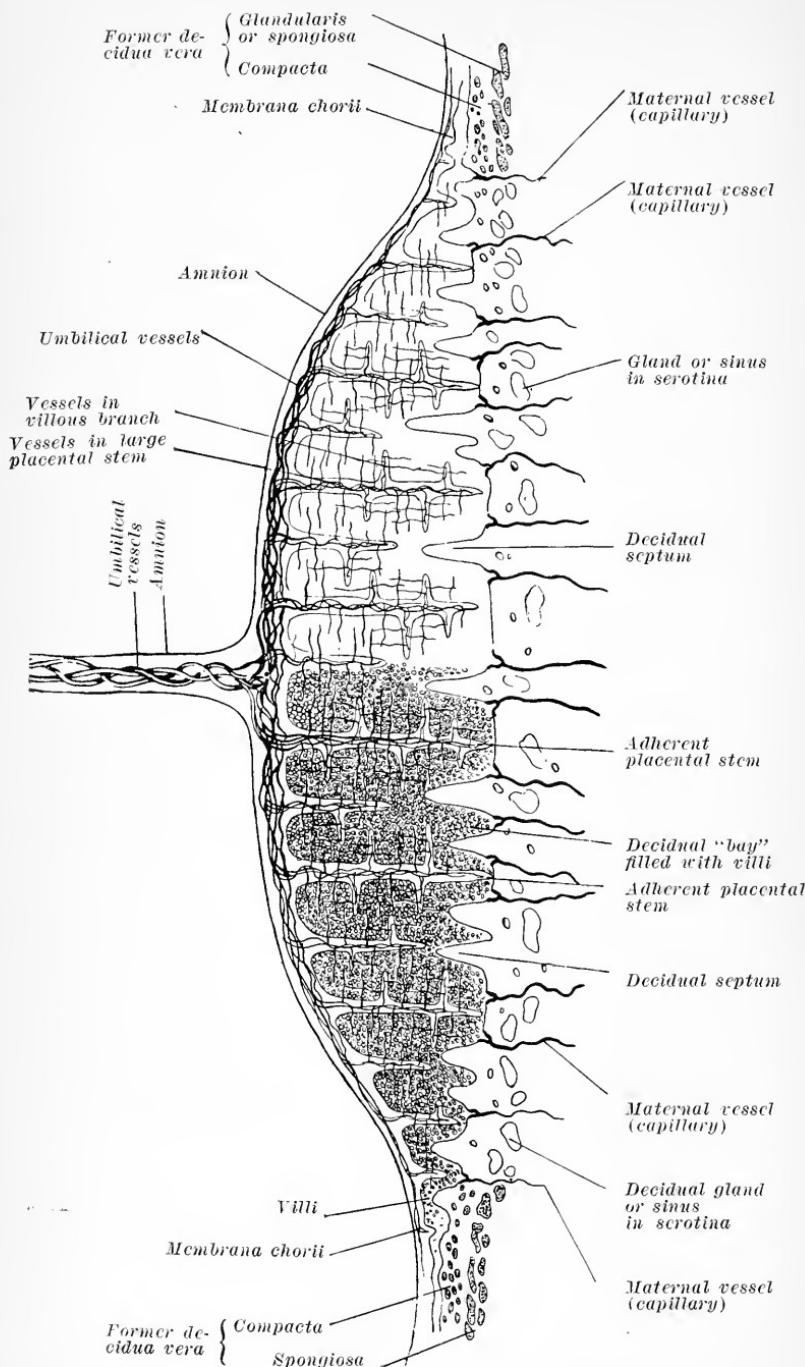


FIG. 46c.—Scheme of placenta. To the right is shown the basic framework serving as the path for the fetal vessels. To the left is shown the almost solid mass formed by the myriads of attached outgrowth or villi, thus increasing hugely the surface bathed by maternal blood. The tips of the large placental stems are attached in the decidual "bays" to the serotina (right). In addition their covering masses of villi are attached to the sides of the "bays" and to the tips of the decidual projections or sepiæ (left). Maternal vessels open into the "bays." The vessels are afferent and efferent, and are really capillary in structure. It is often impossible to distinguish between glands and sinuses.

uterine surface is very irregular. As a result of irregular invasion by trophoblast cells and villi, and as a result of the deeper invasion by the "adherent villi," the uterine surface shows irregular decidual projections. Between these projections and at their base are the "sinuses." Numberless individual villi are loosely attached to the tips and sides of the projections. Masses of villi fill up the hollows or bays. To the bottom of these bays are attached the adherent villi—*i.e.*, the large stems which pass off from the membrana chorii.

The attachment of stems between decidual projections, and the extension of decidual projections between masses of villi, serve to divide off the uterine surface and the placenta into compartments. These compartments are filled solidly with villi. As a result the blood coming from the uterine arteries is limited more or less to the compartment into which it enters. We have the so-called intervillous spaces. The entire placental structure is not washed or infiltrated by all the maternal blood making exit from the uterine placental surface, but by the blood of its compartment.

No space separates external placental surface from uterine surface. The masses of villi composing the placenta have the most external groups close to the openings of the maternal sinuses. Villi and masses of villi extend into the uterine veins. This arrangement, whereby maternal blood passes out between the densely grouped villi and back again through the venous sinuses of compartments, gives the greatest and most plentiful opportunity of surrounding the myriads of villi with maternal blood. *The fetal vessels passing through the cord, through the membrana chorii, through the villous stems, through the villous branches into every villus, give a tremendous opportunity for exchange between maternal blood and fetal blood through the syncytiun.*

The placenta is thus well attached to the uterine wall. In addition the reflexa is attached to the vera at all parts of its periphery. In addition the liquor amnii presses placenta and reflexa closely against serotinal area and vera.

At the edge of the placenta the amnion which lines it passes on and lines the membrana chorii which has not formed placenta, the chorion lœve.

From the edge of the placenta on, the membrana chorii or chorion lœve is attached by means of the reflexa to the decidua vera.

At the edge of the placenta the attachment of reflexa and vera is not very close. This area represents the point of junction of vera, serotina, and reflexa. After union of base of reflexa to vera at this point, the edge of the placenta may extend on upon the reflexa, now united to vera. Beyond this point the attachment of chorion to vera per the reflexa is the same as at other points. The uterine wall at the edge of the placenta has at all times the best-preserved growth of glands, for here the attachment of reflexa and vera is last completed and the pressure is least.

There remains of the decidua only a thin layer at best. In some areas the villi are practically on the muscularis. At other points spongiosa is preserved, but the glands have lost their epithelia and look like vessel sinuses. At other points compacta and spongiosa are fairly well preserved. The invasion of the decidua is very irregular in character (Fig. 43). Generally speaking, little decidua separates the villi from the muscularis. The layer of Nitabuch said to separate fetal and maternal structures, and probably derived from fetal ectoderm, is varying in amount and regularity.

PART II.

THE ESSENTIALS OF TUBAL GESTATION.

CHAPTER I.

PROCESSES ANTEDATING TUBAL GESTATION.

Etiology.—In former years our views concerning the origin of ectopic gestation depended mainly on the discovery of pathological conditions macroscopically evident. Cases were reported with fibroma of the isthmus tubæ or with polyps at the uterine end of the tube. The growth of the ovum in a tubal diverticulum or in an accessory tube was considered to furnish a satisfactory etiology. In some cases the pressure of ovarian or abdominal tumors was supposed to obstruct the onward movement of the ovum. Abel and Freund found in a twisting of the tube and in a failure of development a satisfactory theory for the frequent occurrence of ectopic gestation. Since, in a majority of such cases, peritoneal adhesions are present, these were, and even yet are, considered to so alter the course of the tube's lumen as to prevent the entrance of the ovum into the uterus. Therefore visible inflammations were considered to be the important etiological element.

During my labors a colleague gave me a specimen of extra-uterine gestation combined with a multilocular serous ovarian cyst, with the request that I should find, if possible, a Graafian follicle in the ovary, though he believed none to be present. I considered that this specimen would furnish a proof of external migration of the ovum, and that this migration might stand in an etiological relation to the ectopic gestation. Examination of the ovarian cyst showed no Graafian follicle. Therefore the ovum had come by *external migration* from the other ovary. In Case 3 the same absence of a Graafian follicle was noted.

The experiments of Leopold have shown that the ovum given off by one ovary may enter the tube of the other side. The cases are not so rare in which the tube of one side was closed or absent, and although the corpus luteum verum was found in the ovary of the same side, yet the ovum was found in the uterus. Schröder, Koblanck, and others have found a pregnancy in a rudimentary horn between which and the uterus no epithelial

connection existed. Manierre has collected 39 cases of pregnancy in rudimentary horns. The same is true of those cases in which the corpus luteum verum is on one side and the ovum has developed in the horn of a uterus unicornis of the opposite side. Küstner removed a right-sided extrauterine gestation sac and a left-sided ovarian cyst. Shortly after a uterine pregnancy took place.

External migration occurs frequently in tubal gestation. Although Küstner took note of the frequency of this event in only the last 25 of a series of 100 cases, he found it to have taken place in seven. Prochownik found that external migration had taken place in one case of eight which he had examined closely. Martin found the corpus luteum on the same side as the tubal gestation in thirty-seven cases, on the opposite side in four, and uncertain in thirty-six.

External migration of the ovum has been viewed by Sippel and others as the etiological factor. They believe that the ovum in its migration becomes too large to permit of its passage through the tube lumen. The examinations of Peters, however, show conclusively that no chorionic villi are present until the ovum has been nourished for a considerable time by the decidua in which it is embedded. In addition the Graafian follicle is in the majority of instances found in the ovary of the affected side, so that such an etiology would explain only the smaller number of cases.

This migration, however, calls our attention to the presence of a pathological condition in the mucous membrane of the opposite tube. While it calls our attention to the fact that the other tube is affected, it only proves that it is more affected than the tube in which the ovum is finally embedded, for some cilia must be present in the latter to influence the external migration of the ovum. Various experiments make it seem probable that in the perfectly normal tube no ovum can develop.

Küstner, in his experiments on rabbits, extirpated one ovary and extirpated or tied off the uterine horn of the other side. In these attempts no extrauterine gestation resulted. However, since such a pathological point of development of the ovum does occur in animals, a satisfactory explanation of the failure of these attempts could not be found until the experiments of Mandl and Schmit were published. In their work upon animals they found the following to be the case: After coitus, and after the lapse of time sufficient to permit union between the ovum

and the spermatozoa at the abdominal end of the tube, they tied off the tube at the uterine end. Their results were negative. No tubal gestation resulted. When, however, the uterine horn was tied off a pregnancy in this horn resulted, showing that the ligation was not the disturbing factor and that in all probability ova do not develop on a normal tubal mucosa.

In considering the history of those cases which have been closely noted, it is found that ectopic gestation occurs most frequently in multiparae and that a sterile period of varying length precedes this pathological development. Martin found that 65 multiparae were affected as compared with 20 nulliparae. In a series of 100 cases of Küstner's, only 10 ectopic gestations occurred in nulliparae. The other 87 had borne children and 3 had aborted. In 24 cases it occurred five or more years after the last labor; in 55 cases, from one to five years after; and in 8, in less than twelve months. Veit found that in 52 cases of repeated ectopic gestation a sterile period of two to eleven years preceded the occurrence of this process. Between the two events was a period of six weeks to six years. This sterile period represents the time in which inflammatory changes in the mucosa may occur, either *gonorrhreal*, puerperal, or tubercular. These changes naturally involve the uterine end of the tube more than the abdominal, and in the subsequent course of events, when healing does result, the uterine end improves slowly. What are, then, the pathological changes in the tubal mucosa which stand in an etiological relation to ectopic gestation?

In a series of 8 closely examined cases Prochownik found a gonorrhreal history three times; in one of these cases there was an acute gonorrhreal affection of the pregnant tube. Moskowicz found that of two cases tuberculosis was the etiological factor in one, and that in the other gonococci and staphylococci were present in the pyosalpinx of the non-pregnant tube. Median to the ovum Veit found microscopical changes which represent the results of pus inflammation. In two cases Dührssen found cilia abdominal to the ovum, but none toward the uterine end. Küstner observed very frequently a hemorrhagic tendency of the non-affected tube, showing that tube at least to have been abnormal. I found in three cases distinct changes in the mucosa median to the ovum.

Franz makes *inflammatory changes in the tubes* responsible for the occurrence of ectopic gestation. This is the more probable since inflammatory processes are so frequently found in the

other tube. Franz found such changes in eighty per cent of these cases in which a sterile period of two to seventeen years was noted. In cases where a sterile period of less than two years was observed tubal changes of the other side were present in only 53 per cent. He comes to the conclusion that we must seek the etiology in those afflictions of the tubes which have run their course, and which, having for a long time prevented the moving of the ovum, have permitted a gradual and partial restoration to normal conditions.

While in a certain number of cases no pathological microscopic changes are found in the tubal mucosa, it may be explained by the fact that so-called catarrhal conditions frequently show little microscopical change. Even during or after *gonorrhea* the tube may seem microscopically perfectly normal. Ahlfeld, in an experience of many years at the University of Marburg, met with so few cases of tubal gestation that he considers the relative freedom of his patients from gonorrhea, as compared with those in the larger cities, to be the only explanation.

Various inflammatory influences are etiological factors in that *they destroy the cilia in whole or in part or diminish their functional activity.* Besides, from the experiments made on animals we know that absence or early atrophy of the ovaries influences the muscular development and the functional activity of the uterine wall and the structure of the mucous membrane and the cilia. In cases of functional interference with the secretion of the ovary, or in the atrophy subsequent to labor or as the result of lactation or of constitutional disturbances or of failures of development, *the activity of the cilia is diminished.*

Naturally there must be activity to a certain extent on the part of the cilia, especially at the abdominal end of the tube, for extrauterine gestation occurs most frequently in the isthmus tubæ. Prochownik found an ampullar location in only 3 cases out of 45. Mandl and Schmit found in 69 cases an ampullar situation in only 15. Dührssen found an isthmic location in all of his 29 patients. In the 5 cases which I have examined closely the same is true. Although Mercier has collected 30 cases of interstitial gestation (with rupture), Leopold 10 cases of ovarian gestation (?), and although the growth of an ovum on the fimbriæ, as well as tubo-ovarian pregnancies, occurs, yet the vast majority are found near the uterine end of the tube.

A further proof may be found in the fact that recurrences of tubal gestation take place but rarely in the same tube.

Patellani, in a tabulation of 36 cases, found that first one tube and then the other was the seat of development. Veit in 52 cases found that it recurred only three times in the same side. An additional point of importance is the occurrence of tubal gestation in either tube at the same time, of which Gebhardt mentions 9 cases. Further, Patellani has collected 37 instances of combined uterine and extrauterine gestation—a practical proof of an affection of one tube, and certainly excluding external migration.

I believe that in the so-called sterile period *gonorrhreal*, puerperal, tubercular, and atrophic processes take place. The interval of years between the last labor and the ectopic gestation, the fact that the location is generally in the middle area of the tube, the fact that repeated gestations are observed and rarely in the same tube, the occurrence of an ectopic gestation on both sides at the same time, and the frequency of external migration together with a combination of extra- and intrauterine gestation, point certainly to an affection of one tube and probably, but to a lesser degree, of the other tube. The frequency with which, according to Küstner, a hemorrhagic tendency of the non-affected side occurs, as well as the microscopic discovery of catarrhal conditions, together with the history and the microscopical evidence of the presence of *gonococci*, point distinctly to a tubal affection. The observation of Dührssen, who found cilia abdominal to the placental site and none median to it, and Veit's observation of the presence of inflammation median to the ovum, as well as the theory of congenital and acquired atrophy of the tube, especially subsequent to labor, lead us at the present day to seek in the microscopical changes of the tubal mucosa, *the injury to the cilia*, the etiological factor in tubal gestation.

CHAPTER II.

VARYING VIEWS CONCERNING THE HISTOLOGY OF TUBAL GESTATION.

The Decidua.—In the uterus the ovum descends by a centrifugal process into a well-developed decidua, which is gradually divided into a superficial compact layer and a deeper spongy glandular layer. Concerning the existence of a tubal decidua, however, opinions differ. Webster's view that a compacta and spongiosa are formed receives no support. Wyder found, in one case, decidua in the periphery of the region occupied by the ovum. Mandl describes decidua cells in the placental area, while Veit found decidua cells not only at the placental site but at a distance from it on the opposite wall. In his eight closely examined cases Prochownik describes decidua cells and in one a well-developed decidua. Abel found a well-marked decidua about an ovum, especially in the region of the placenta. His statement is positive, since he compared it microscopically with the decidua cast out from the uterus.

On the other hand, Aschoff found none at the placental site. Kühne describes only a *pseudo-decidua consisting of fibrin, connective tissue, and invading ectoderm cells*. Though slight decidual changes may occur at the placental area, *the cells which have been previously described as decidual are considered by him to be the cells of Langhans, and the same view is shared by Aschoff and Ulesko-Stroganowa*. In most cases fibrin masses, blood, villi, and cells which at least look like decidua cells, but which many consider to be trophoblast cells, are to be found. It is a fact, however, that at the area of greatest pressure the so-called decidua is often thinned to a decided extent.

Veit in turn considers the so-called cells of Langhans to be, in the peripheral areas, decidual cells. Though he finds cells of the layer of Langhans in the maternal vessels, yet they are not connected with like cells about the vessels. There is an absence of like cells between the vessels and the serotinal surface. While granting that the resemblance is great, he considers those interstitial cells present in the peripheral areas about the ovum to be

decidual, and concludes that the connective tissue in the circumference of the intervillous space in tubal gestation undergoes decidual change, only that this layer is very much thinner than in the uterus.

Embedding of the Ovum, and the Reflexa or Capsularis.—A considerable difference of opinion exists as regards the reflexa. Wyder found none in his case and the ovum was only loosely attached. Abel found no capsularis and most of the villi were free and only few were adherent. Werth described sections of a tube in which *the ovum had settled at the height of a tubal fold*. Werth described, in another case, *a capsule containing muscle fibres passing out at the base from the muscular fibres of the tube wall*. The external covering of the capsule consisted of epithelium. According to Kreisch, *the capsule may consist of united folds*. He described the presence of a reflexa in three cases, however, which was not composed of tubal folds. In a specimen examined eight days after conception a reflexa was found by him. The formation of a pseudo-reflexa readily takes place if the ovum is embedded in a thick system of folds. Others believe that in many cases a capsule consisting of longitudinal muscle fibres and mucosa, with no epithelium on the inner side, is of frequent occurrence.

Aschoff and Füth make it evident that the ovum makes its way down into the mucosa, as is the case in the uterus, and that it extends even further deeply into the muscular wall. It is then covered laterally by the inner layers of the muscularis and by the mucosa, which are pushed over it, *forming a capsularis and bounding the intervillous space*. In several closely examined cases the ovum has been found almost under the entire thickness of the tubal wall, lying directly on the vessels of the ligamentum latum. Muscle fibres are absent, however, at the summit of this resulting capsule. In another case the ovum was nearly extratubal, lying between the muscularis of the tube and the muscle fibres of the ligamentum latum. The ovum was, of course, separated from the lumen by the mucosa and by muscle fibres. At the apex of this capsularis there was considerable thinning, but the epithelium upon the mucosa was intact. Such a development would explain the so-called pseudo-intraligamentous cases of Küstner. The capsule has been found in some cases to be lined with the cells of Langhans.

Intervillous Space.—If the capsule be composed of folds, according to Kreisch, they may be so firmly united that an *inter-*

villous space may exist. In Abel's case the space between the chorion frondosum and the decidua was larger than at any other point; adherent villi extended into the decidua, but most of the villi were found free in the space. The vessels of the decidua opened directly into these intervillous spaces, which contained blood. According to Aschoff, the centrifugal descent of the ovum makes the existence of an intervillous space easy. Veit, in a well-preserved non-interrupted case, found a *well-developed intervillous space in which blood circulated*, and traced a vein containing villi into this common intervillous space. Kühne, on the other hand, considers the invasion of the blood vessels by the villi to be pathological, and does not consider the existence of an intervillous space probable, because in eight cases he found no reflexa. Since Von Both collected 84 cases of ectopic gestation with viable fetus, an intervillous space must have been present.

Villi.—Abel, as stated before, finds a decidua vera in the peripheral areas of the ovum; the villi, however, he found *to extend only up to the muscle.* Leopold and others, on the contrary, find the *villi extending deeply into the muscular wall of the tube.* The villi are covered with two layers. At their ends are pillar-like groups of cells growing into the wall, between the muscle fibres, and representing extensions of the trophoblast. The cells of Langhans growing into the muscle and uniting with it form the so-called *Saugeplatte.* It is with this layer that the villi unite through the cell groups found at the ends. These complexes of cells of Langhans and syneytium speak for the fetal nature of the latter, and the pillar-like groups are simply groups of trophoblast not yet reached by the mesoderm.

Abel found most of the villi free and only some adherent to the decidua. As they did not enter into the muscle, the connection was not a close one. Wyder removed an ovulum in its entirety from a tube and later examination showed *no villi extending through the decidua to the muscle.* Kühne, Ulesko-Strogonowa, and others find that the *cells of Langhans extend actively into the submucous tissue and into the muscle bundles,* and believe that these are the cells which are often mistaken for decidual cells. It is upon them, as upon the compacta of the uterus, that the villi grow. At the same time, *they may extend entirely up to the serosa of the tube.*

As just mentioned, the villi may extend up to the serosa, and perforations of the wall covered by thrombi have been frequently found. Kühne describes cells which have entered the serotinal

veins. In the vessel spaces under the pseudo-decidua he finds free cells whose character proves them to be pseudo-decidual elements. He considers these cells to have entered through the vessel walls. Mandl found decidual cells in the vessels and believes that they have passed out of the surrounding tissue into the lumen. Other cells in the same location he views as proliferated endothelium. Cornil found syncytial elements in the vessels and considers them decidual cells which have entered through the vessel walls. Other investigators hold that the cells of Langhans grow directly into the submucous tissue and into the muscle bundles, and that these cells *may penetrate the vessels and open them as in the uterine compacta.* They describe the villi as following the course of these cells of Langhans into the muscularis, up to the serosa and through the serosa, which latter perforations are usually covered by thrombi. Aschoff, Ulesko-Stroganova, Leopold, and others have described *villi as perforating the vessel walls.*

Deportation.—By Veit a quite different theory is given, however, to explain the presence of fetal cells and villi in the muscular wall of the tube. In discussing the relation of the ovum to the uterus, Heinz considers that the fetal tissues grow into the maternal structures and says that the *maternal glands, vessel walls, and tissues are eaten up by the villi.* Cornil considers that the ovum in an extrauterine gestation not only does the same, *but follows a course resembling that of a uterine chorioma.* While Veit may grant the correctness of such a view in pathological conditions, he does not believe it justifiable to consider it the usual course.

In the case of a normal uterine gestation, villi do not project into the arterial openings of the intervillous space. They do, however, project normally, not only in the border sinuses but at all the serotinal points, into the veins alone. The villi are of normal structure and often so numerous that they occlude the vein lumina. They are frequently very long and follow the course of the veins to a considerable distance. In the uterus, villi are thus found in direct contact with the uterine muscle, or, better, in the veins of the uterine muscle. The same is true in tubal gestation. Whether this occurs in all cases or not cannot be positively stated (Veit).

Veit believes that cells in the periphery of the ovum enter at an early period into the peripheral veins. In normal gestation the uterus grows as much as the ovum needs, and the ovum

as much as the uterus permits. Later, as soon as the ovum in the uterus has attained a certain size and the edge of the placenta extends laterally, another process is added to that of general growth. This process is brought about by the villi which project into the serotinal veins. If a villus enters a vein it generally remains in contact with the periphery of the ovum and grows further in its length as well as in its circumference. In its growth it dilates the lumen of the vein so decidedly that its serotinal portion is everted and taken up into the general intervillous space. On either side endothelium then extends for a short distance upon the serotinal surface. In the uterus, normally, only cells of the ovum or of the villi are loosened and pass on into the veins, but the villi themselves, as a rule, remain in contact with the ovum. In uterine gestation the entire endometrium becomes deciduous, and, therefore, decidua cells are found in the periphery of these vessels which contain villi. In the peripheral region of the well-preserved tubal sac Veit found free villi in a vein. Series sections showed this vein to enter into the intervillous space. Therefore the villi were no longer connected with the ovum. This occurrence, whereby syncytium and villi, loosened from all connection with the ovum, are found in the veins, is given by Veit the name deportation.

Veit, then, explains the descriptions of Aschoff, Füth, Cornil, and Ulesko-Stroganova, who find a destruction of the tube wall and an invasion of vessels by the villi, on the theory of deportation through the veins.

Veit does not believe that the fetal cells possess the power of wandering, because he finds the cells of Langhans in a vessel but no such cells about it. Therefore, in his opinion, the cells have not perforated the vessel because none are present in the connective tissue. Veit, however, forgets that, in addition to deportation, a wandering of cells and a perforation of the vessels does occur.

Veit denies that the cells of the ovum wander into the connective tissue. We know that cell groups go gradually over into the cells of Langhans on the villi, while on the other side they are either separated from the decidua by fibrin, or else the cells pass gradually into the decidua with such a resemblance to the latter cells that it is difficult to differentiate them.

Veit does not believe that the cells of Langhans in the periphery of the tube look different from those which are near the villi. He found at the area of the tubal vera a few changed

connective-tissue cells and considers them to be a beginning stage of a decidua vera. In the periphery he saw cells about the vessels differing somewhat in appearance and stain from the cells of Langhans, and, since they are like the above-mentioned vera cells, he considers them decidual. In this connection he overlooks the power of the trophoblast.

Where the cell groups enter the decidua he sees a certain form of cells. At other points where, in series sections, no villi enter the serotina he finds the same cells. Therefore either the cells of Langhans can wander in all directions or else a decidual change is present, and he believes the latter to be the case and states that a decidua exists, but much less than in the uterus.

CONCLUSIONS.—*In the uterus there takes place a centrifugal descent of the ovum on a spot free of epithelium. The compact layer of the decidua forms the enveloping zone. There is an early connection between the maternal blood and the fetal cells in the form of a trophoblast which is a product of the ectoderm. The trophoblast, at points, extends far into the compacta and the cells have a decided power of wandering. The trophoblast is changed to syncytium through the corrosive action of the blood. The opening of the maternal vessels occurs even before villi are formed, and at this and at all subsequent stages fetal cells enter the maternal circulation. The primary enveloping zone becomes the subsequent intervillous space. Neither the uterine epithelium nor the maternal endothelium play any part in the formation of the syncytium. No reflexa is formed, but through the descent of the ovum a capsularis results. The ovum, through its trophoblast cells and villi, invades and destroys the maternal decidua. The fibrin layer of Nitabuch and the thickness of the decidua prevent the extension of the villi too deeply into the wall.*

In the tube we find no division into two layers, as is the normal in the uterus. In fact, no decidua is formed. There is an absence of the thin-walled decidual vessels, and the muscularis, if at all, is only slightly hypertrophied. We are only anticipating the results of our investigations when we say that, *with the exception of the absence of the decidua and an enveloping zone composed of compacta, the processes of gestation in the tube are the same as those in the uterus, modified only, as we would naturally expect, by the absence of the decidua and the thinness of the tube wall.*

The varying differences of opinion are due to five causes:

1. Ova of different states of preservation have given, quite naturally, different pictures.
2. Ova of very different ages have been described and compared without taking this fact into consideration.
3. As in the uterus, but even more so, the differentiation between fetal and maternal cells has been almost overlooked, so that trophoblast cells have been viewed as decidual. The ability of the fetal cells to wander, to invade the tube wall, and to perforate the vessels has been granted by only a few investigators.
4. The fact that there are several forms which the ovum follows in its embedding in the tube has almost entirely escaped attention.
5. While Veit's deportation is an accepted fact, he leaves out of consideration the processes of "cell wandering" and "vessel invasion."

How nearly and how closely in all important details the histology of tubal gestation may resemble the course of uterine gestation, will be seen in the following pages.

CHAPTER III.

EMBEDDING OF THE OVUM AND DEVELOPMENT OF EXTRA-EMBRYONAL STRUCTURES.

Of the cases examined in series sections, these three are taken as types representing the important forms and stages.

I. THE COLUMNAR TYPE OF TUBAL GESTATION.

The following specimen (Fig. 47) presents an oval body, measuring one-third of a centimetre in its greatest diameter, situated on the mucous folds of a tube removed because of the

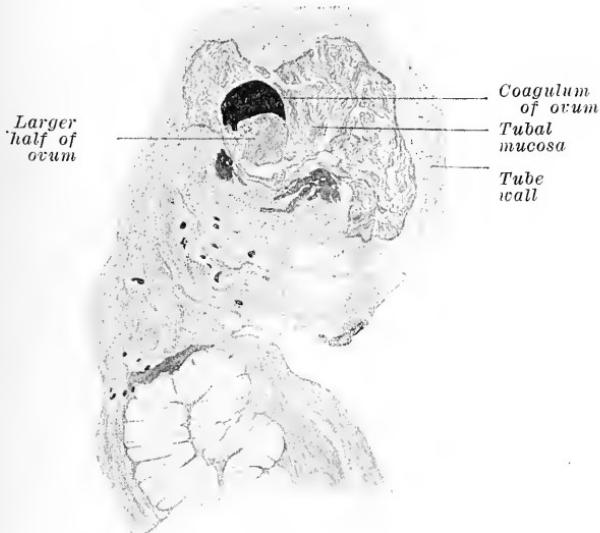


FIG. 47.—Very young ovum situated on the tip of the folds of the tubal mucosa in a case of ectopic gestation accompanied by an old intraperitoneal hemorrhage.

diagnosis of ectopic gestation (on operation the abdomen contained clotted blood). The tube was stained *in toto*, embedded in celloidin, and cut in series sections. Fifty slides from the middle area of the tube contain sections of this oval-shaped structure (Fig. 47a). A section through its greatest diameter shows it to be composed of two unequal halves: a smaller half consisting

of a blood coagulum, and a larger half composed of closely grouped cells, of protoplasmatic cells and masses containing one or more nuclei, and of villi. Villi are also found, but sparsely, about the coagulum (Fig. 48).

Sections through the smaller diameters show the coagulum diminishing in size and extent, with an actual and relative increase in the extent and area composed of closely grouped cells

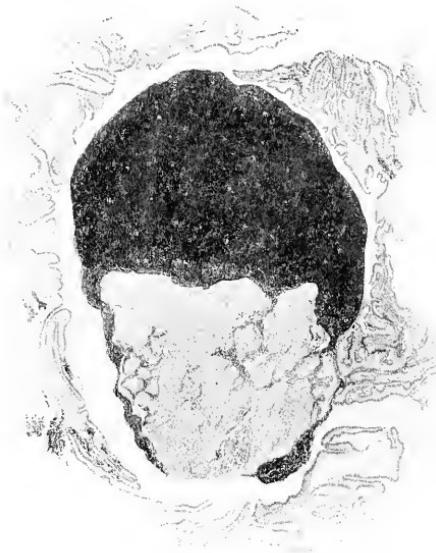


FIG. 47a.—Ovum of Fig. 47, showing division into two halves and an intimate connection with the tubal mucosa on the right. (Section through the greatest diameter.)

and villi, furnishing, then, a centre composed of cells and a periphery in which are villi (Fig. 49).

The more external sections are continually smaller (Fig. 50), and the central cell area diminishes in size, so that the small and most external sections present only villi (Fig. 51). In all the sections the villi are in the periphery of the centrally situated closely grouped cells.

By reconstruction, then, we obtain a solid, oval-shaped body possessing a covering composed of protoplasmatic cells and villi. Within this covering are closely grouped cells, and in the interior, although eccentrically situated, is the coagulum. We are dealing with an ovum (?) in which the amnion, umbilical vesicle, and the central cavity are obliterated by blood, while the trophoblast and villi are preserved. This ovum, as we may view it,

is adherent on one side to folds of mucosa which evidence a proliferation of their epithelial covering. On the epithelial



FIG. 48.—Another section through ovum of Fig. 47, showing in one half trophoblast cells, villi, and very dark syncytial masses.

covering, and between the folds, are rows of round, structureless bodies densely grouped, and the same are found at numerous points covering the ovum, except at the end opposite to the



FIG. 49.—A more peripheral section, showing trophoblast cells and villi and very little of the blood coagulum.

coagulum. In other words, on and between the mucosa folds in contact with the ovum is seen a plasmatic substance distinctly excreted by the epithelium, and in it are also red blood cells and leucocyte nuclei.

The tube wall facing the other end of the ovum presents mucosa folds, whose epithelium is interrupted at various points, while at other points are proliferations of epithelium and round cells. The stroma of many of the folds is absent, while others possess dilated capillaries and vessels, and still others are filled with blood extravasations. The tube wall itself is covered with epithelium, and in the wall are extravasations of blood, and dilated arteries and veins. *At no point in the tube wall or in the mucosa is there any decidual change or any condition representing an entrance of trophoblast cells or of villi.* The ovum is surrounded by a plasmatic substance which I have observed on uterine and cervical polyps covered with epithelium and in active growth. It is not a deposit from the blood, as such, but one actively secreted or produced. This same element is found on the mucosa folds in immediate contact with the ovum. The

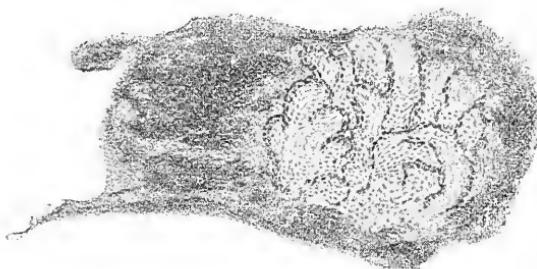


FIG. 50.—A still more peripheral section, showing only villi.

adhesion of the ovum at these points is a firm one. It is well preserved and therefore in active contact with some nutrition-producing tissue. The shape and character of the oval body, together with its other characteristics, prove it beyond doubt to be an ovum. If not entirely surrounded by the tips of the mucosa folds, it was probably situated among them near the tube wall, but not on or in the wall.

The other end, with the numerous trophoblast cells, plasmoidal elements, and villi, was probably surrounded, too, by mucosa folds, and here, too, an intervillous space, containing blood coming from the capillaries of the mucosa, was probably present. Although we shall see that this end was infiltrated by maternal blood, I cannot forbear, in this connection, to mention the possibility that the ovum obtained nutrition from the epithelium of the mucosa at the point of adhesion, which represents to me the *reflexa* or *capsularis* composed of tubal folds.

(In the placenta of many animals it is found that the blood extravasated from the uterine vessels is made use of by the fetus, in that the degenerating products of the extravasated blood cells are taken up and absorbed by the ectoderm cells. According to Strahl, in Galago, a Madagascar lemuride, numerous extravasations are found in the uterine mucous membrane, and their products lie in the connective tissue in the form of larger and smaller yellow granules. The epithelia of the glands near this extravasated blood are more or less filled with granules resembling these. He considers them to be rests of the blood cells which are taken up by the gland epithelium and made use of by the latter in furnishing an iron-containing secretion. The gland epithelia here perform a function which in other placentæ is performed by the fetal ectoderm. In Galago fetal villi are always



FIG. 51.—Most peripheral section, showing a villus only.

in contact with the uterine epithelium and obtain nourishment from it. Between the uterine epithelium and the epithelium of the villi are seen strands which represent the uterine milk taken up by the villi. Thus the fetus gains the greatest portion of its nourishment through the chorionic epithelium.)

The main cell body is composed of cells, without distinct cell boundaries, containing nuclei of varying form and staining darkly. At various points are found paler cells with a distinct nuclear membrane which unite in groups of three or more. More external, in the region of the villi, the cells are more distinct and the nuclear membrane is more clear and nucleoli can be distinctly seen (Fig. 52). The more external these cells are found the larger and the more swollen and the paler do they become, and a homogeneous intercellular substance is present. In the centre of those sections where the cells are so densely grouped (Fig. 49) are minute, darkly-staining granules whose identity at first was not distinct. In the more peripheral areas, however, their char-

acter is evident; for they are only found where red blood cells are present and they are therefore leucocyte nuclei. The nearer we approach the circumference the more do we find blood in the interstices between the cells, at first as isolated cells or groups, and later in larger spaces containing red blood cells. Wherever blood in small amount is present, the cell nuclei become dark and change their form, becoming long spindle-shaped, arranged in single strands or in longer parallel groups. Where, however, blood in larger amounts is present, protoplasmatic masses containing numerous round or spindle nuclei may be observed. Near the

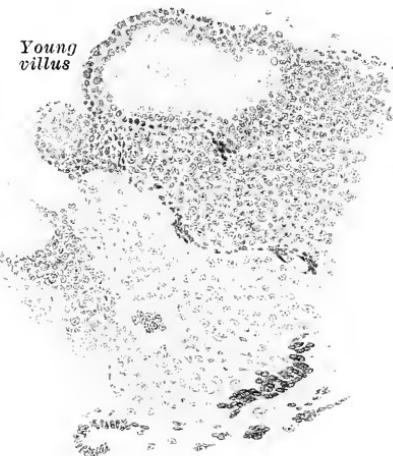


FIG. 52.—An area from Fig. 48, showing trophoblast cells, their transition to the chorionic epithelium in the upper part, and to syncytial groups in the lower part.

circumference the number and extent of the round protoplasmatic groups and the number of the darker or longer spindle-shaped nuclei increase, and the latter seem to invade the sections described from the exterior toward the interior. In the exterior of the sections are villi covered with a double layer of thin syncytium with flat nuclei. The centre is composed of embryonal connective tissue staining blue and containing branching cells (Fig. 52). Other villi, some of which are quite long, have a centre composed of round nuclei with nucleoli which are very closely grouped, and transitions from these cells to syncytium may be distinctly observed, as in Fig. 52. These villi, as well as the others, are found only in the external area or near the coagulum. Knobs of syncytium and so-called syncytial giant cells are pres-

ent about these villi, connected by a pedicle or lying apart. In the central areas no connective tissue can be observed. In the external layers, however, as said before, it is evident in the form of a structureless substance between the cells, giving these areas a pale appearance (Fig. 52). It is not present in the younger villi and is only evident in the fully formed ones.

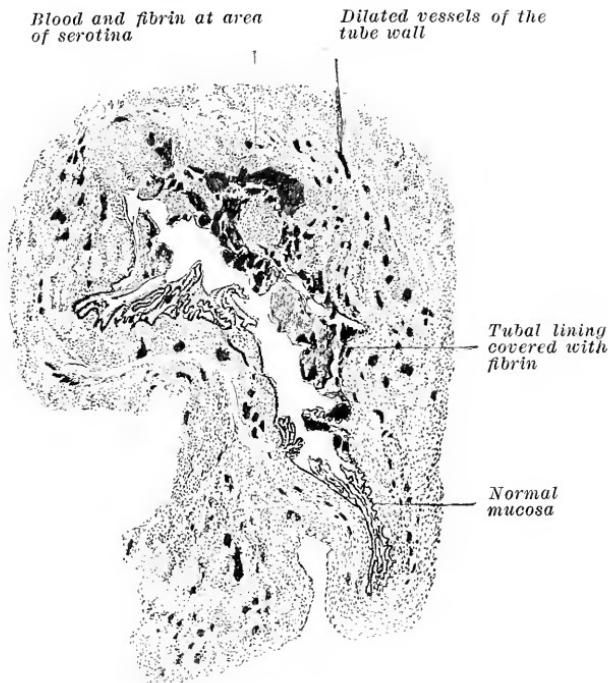


FIG. 53.—Section of tube, showing normal mucosa in the lower and left area, while above and to the right is the area of attachment of the ovum, giving a typical picture of the tube after tubal abortion.

II. THE INTERCOLUMNAR TYPE OF TUBAL GESTATION.

A further stage of development of a tubal ovum is well represented in Fig. 53. The examination, in series sections, of this specimen, known clinically as tubal abortion, divulges numerous interesting features. One half of the circumference of the tube lumen is of a normal character; the other half is in a torn, infiltrated condition which involves not only the mucosa folds but also the submucosa up to the muscularis. Close examination evidences the fact that the ovum was situated on the tube wall, compressing and destroying the mucosa folds at the situation known as the serotina. On either side of this point the mucosa

folds were evidently united about the ovum, forming a pseudo-reflexa. The capillaries and large vessels at the serotinal point are filled with blood (Fig. 53), and many evidence larger or

Muscular fibres of tube wall

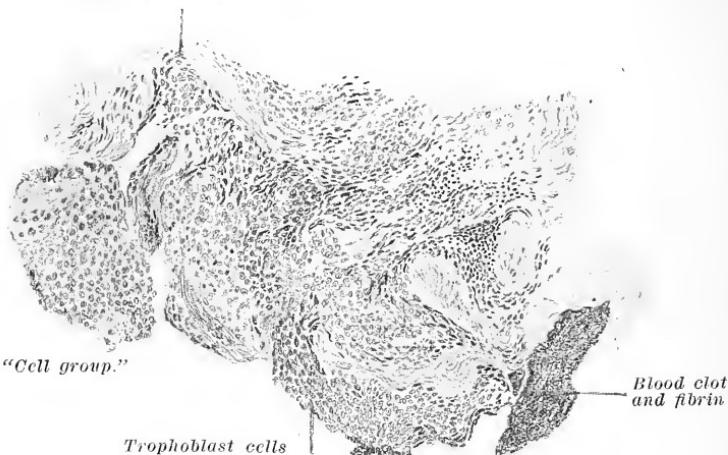


FIG. 54.—Portion of serotinal area of Fig. 53, showing pale trophoblast cells and cell groups and the darker syncytial cells.

smaller injuries and invasions of the wall. At the serotinal portion and for some distance on either side is observed in the submucosa a tissue resembling at first sight decidua.

On close examination we find oval, very closely grouped cells,



FIG. 54a.—Typical area in Fig. 54, commonly mistaken for decidua, showing pale trophoblast cells and dark syncytial cells.

consisting mainly of nuclei and possessing each a nucleolus (Fig. 54). They are extremely closely grouped and between them are found no capillaries or spaces. This tissue rests not only on the free surface of the tube wall, but invades it at many points to a

considerable depth (Fig. 55). The invasion of the tube wall is not an even one, for at some points it forms a continuous deep layer like the decidua, at other points it enters the submucous tissue in irregular branches and projections, and at other points, through lateral infiltration, is separated from the free surface of

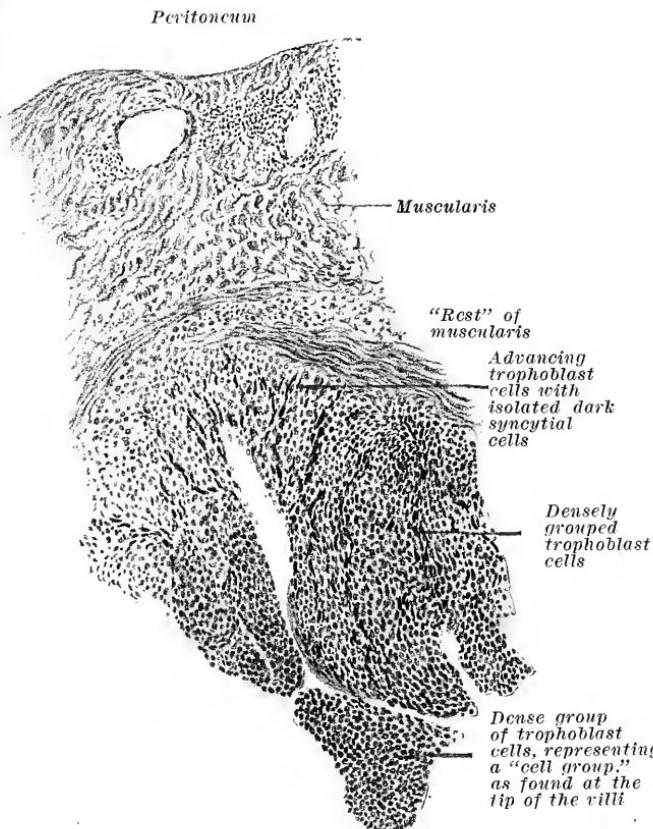


FIG. 55.—Section through tube wall at serotinal area, showing the paler character of the advanced trophoblast cells and the darker character of the densely grouped trophoblast cells near the ovum. Typical field formerly mistaken for decidua, but differentiated by the dark spindle-shaped syncytial cells. This drawing shows that in tubal abortion the "rests" are of the same nature as in uterine abortion.

the tube wall by a considerable space of normal submucous tissue.

These cells are found about the smaller and the larger vessels, invading and infiltrating their muscular walls up to and into the lumen of the vessel.

The character, arrangement, and structure of these cell masses

leave no doubt that they are trophoblast cells. In addition they may be identified by the fact that at almost all points, especially those points near the inner surface of the tube, they are accompanied by isolated, long, spindle-shaped protoplasmatic cells, by groups of polynuclear protoplasmatic masses, and by long, often parallel bands of the same character (Figs. 54*a* and 55*a*). These are the syncytial cells, absolutely identical with the same elements observed in all the previous specimens, especially those seen in the first tubal ovum.

These cells may be further identified as trophoblast cells by the fact that they are found, at the tip of some adherent villi,

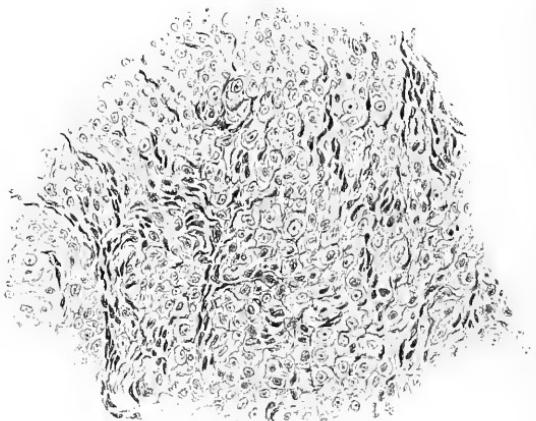


FIG. 55*a*.—Another area from Fig. 55, showing the "advance guard" of infiltrating trophoblast and syncytial cells.

extending directly into the tube wall. These trophoblast cells enter into the submucosa and muscularis, forming the tissue which so many investigators have called decidua. These complexes of trophoblast and syncytial cells, especially the groups found at the tips of the villi, are not yet filled out with mesoderm, but are to form future villi.

At no point do the connective-tissue cells of the tubal folds or of the submucosa evidence any change resembling, in the slightest degree, those changes occurring in the uterine mucosa which result in the formation of decidua cells.

At no point do the epithelial cells of the mucosa evidence any change of a so-called syncytial character.

III. THE CENTRIFUGAL TYPE OF TUBAL GESTATION.

Figure 56 represents a tubal gestation, containing a fetus, in the sixth or seventh week. The tube wall is preserved in its entire circumference, with the exception of two minute areas where the chorionic villi have actively perforated it. Projecting into the dilated lumen of the tube, and almost touching the mucous lining, is the outer covering of the fetal sac, the capsularis. One half of the circumference of the tube is invaded, infiltrated, and

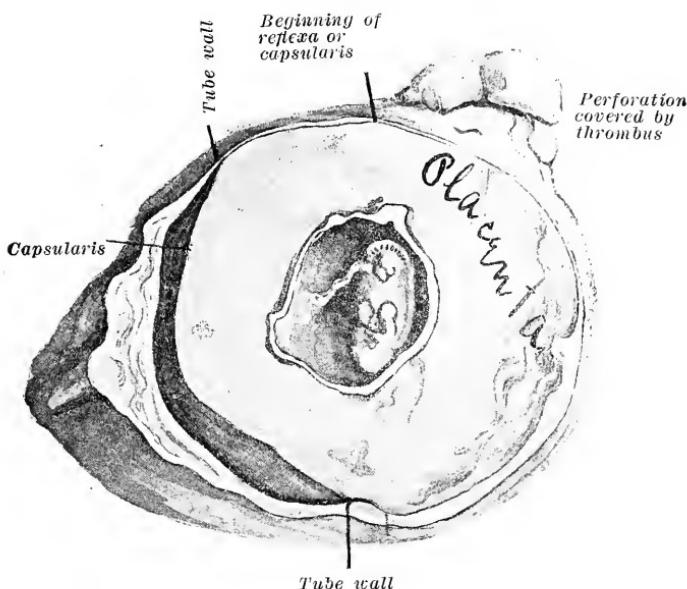


FIG. 56.—Tubal gestation with fetus attached by umbilical cord, showing a perforation by the growing villi closed by a thrombus.

stretched by the other half of the fetal sac wall, that is, the placenta. On both sides, at the junction between the infiltrated tube wall and the preserved but decidedly stretched tube wall which surrounds that half of the fetal sac which projects into the tube lumen, the mucosa of the latter passes over upon the fetal sac and can be followed upon it as a covering of the latter for a certain distance. The summit of the sac is covered by a tissue composed of fibrin, trophoblast cells, leucocytes, etc. The ovum possesses, then, a capsularis composed in part of fetal folds and submucosa. This capsularis is, strictly speaking, no reflexa. It must be called a pseudo-reflexa or capsularis.

The area of the tube circumference entirely filled by villi is

the true placental site, and here is found a real intervillous space bounded on all sides by the point of union of the tube wall with the base of the capsularis. The remainder of the circumference of the fetal sac evidences villi, but the space between the capsularis and the inner lining of the sac is mainly filled with blood and blood crystals. At the real placental site almost no remnants of the tube wall can be found. At this half of the tube wall there are nothing but villi, trophoblast cells, syncytial cells, and a sea of maternal blood (Fig. 56a).

The process involved in this invasion of the tube wall and the

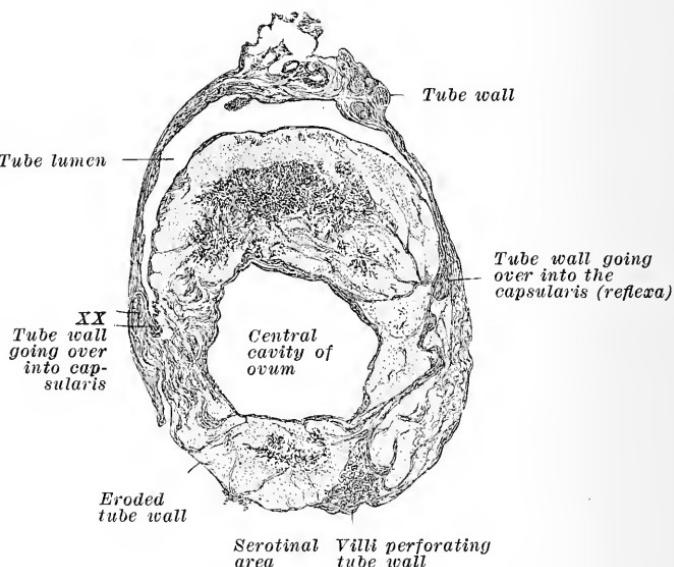


FIG. 56a.—Tubal ovum with perfectly developed capsularis. Tubal wall passes on both sides over into the capsularis. Major portion of placenta is at the lower border, where tubal tissue is eroded and perforated by the villi. This area is the intervillous space. Between the tube lumen and the central cavity of the ovum is the capsularis filled with blood and blood crystals.

process involved in the formation of the villi can, in each section, be extremely well judged on either side at the point of junction of the capsularis with the tube wall (Fig. 57). Here, on either side, there extend into the gradually thinning tissue of the tube villi, cell groups, and syncytial cells which invade the muscularis in what may be called a concentric path. We observe, at the tips of fully formed villi, the typical trophoblast cell groups. We see them extending into and between the muscular fibres, changing into and accompanied by syncytial

cells and masses of every kind and form yet described (Fig. 58). At no point is there the slightest evidence of any decidua or of any decidual reaction. The trophoblast cells invade the vessels of the tube wall, and the villi themselves enter the tube wall and, with the trophoblast groups at their tips, enter the vessel lumina.

At the main placental site are found trophoblast cells with the clearest and most distinct change into various forms of independent syncytial groups and into the syncytial covering of the

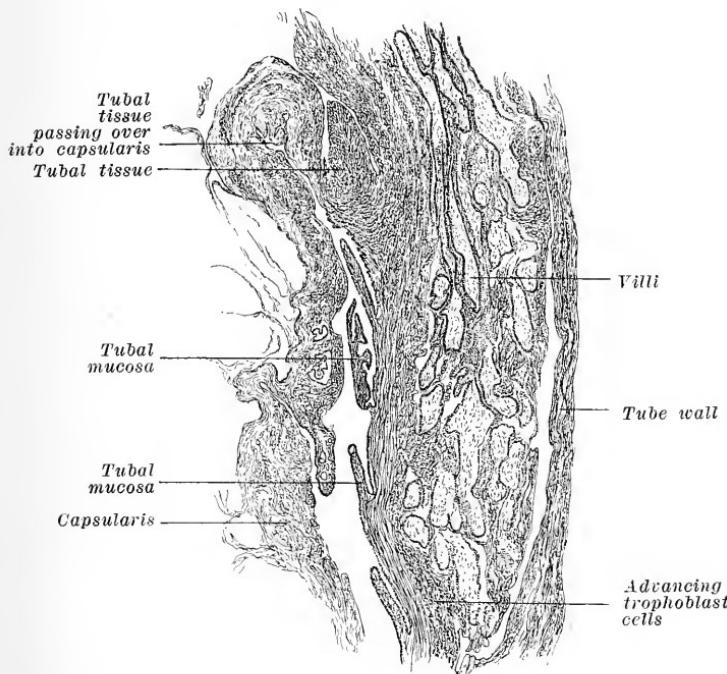


FIG. 57.—Area XX of Fig. 56a magnified, showing the reflexa or capsularis and the infiltration and invasion of the tube wall by the trophoblast and villi.

villi (Fig. 58). They are likewise seen to pass over gradually into the layer of Langhans, which at all points resembles the syncytial layer to such an extent that it might be said that the covering of the villi consists of a double layer of syncytial cells (Fig. 59). The villi have actively perforated the tube wall individually and in groups, so that the intervillous space communicated with the abdominal cavity at these points.

This stage is the exact macroscopic and microscopic counterpart of the stage seen in Fig. 39.

If no interruption take place, the capsularis unites with the mucosa of the enveloping tube wall in the same way that this process is exemplified in the uterus.

The various steps in the succeeding months of development of a tubal gestation may, so far as the development of the placenta, the fetal sac, and the fetus is concerned, be considered as identical with the same processes in the uterus; for over ninety cases of full-term ectopic gestation with viable fetus are recorded.

The various steps in the development of the tubal placenta,

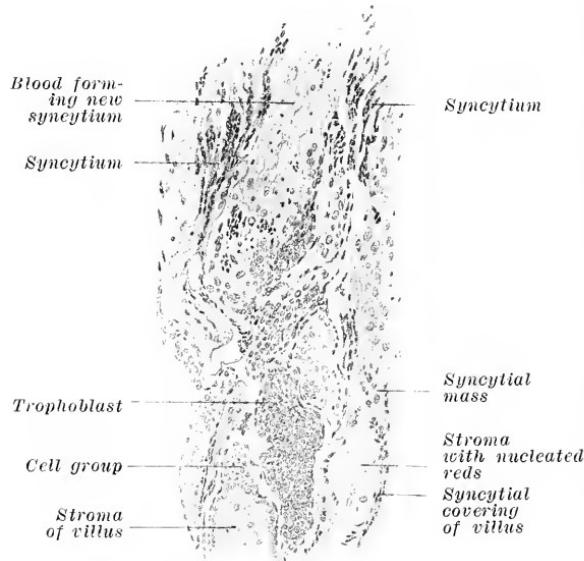


FIG. 58.—Area of advancing trophoblast cells of Fig. 57, showing transition of trophoblast cells to syncytial groups and to the syncytial covering of the villi.

depending as they do mainly on the cells of the ovum itself, the trophoblast cells, are the same as in the uterus. The difference is mainly in the different character of the base or trophospongia. We may draw the following conclusions:

(1) *In tubal gestation no decidua or trophospongia develops. The mucous lining of the uterus is really a lymph tissue; the submucosa of the tube is not.*

(2) *Fetal cells, in tubal gestation, may at any time enter the maternal circulation.*

(3) *As regards the trophoblast, the syncytium, the villi, the formation of blood by the trophoblast cells, and all other par-*

ticulars, the processes depending on and originating in the ovum are the same in both uterus and tube.

Certain investigators have described ova embedded in the tube wall, where a capsularis was present containing muscle fibres passing out from the tube wall into the base of the capsularis. The ovum was found entirely under the entire thickness of the tube wall, lying on the vessels of the ligamentum latum—the so-called pseudo-intraligamentous form.

CONCLUSIONS.

In the tube, the embedding of the ovum and the development of the placenta, then, is found to follow three fairly distinct forms: (1) the columnar, (2) the intercolumnar, (3) the centrifugal.

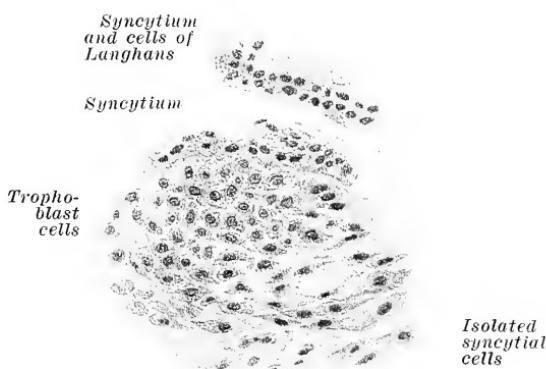


FIG. 59.—Drawing from serotinal area of Fig. 56a, showing trophoblast cells going over into isolated syncytial cells and into syncytial masses.

1. In the columnar type of development (Fig. 47) the ovum is surrounded by mucosa folds only. Here an invasion of the capillaries of the tubal mucosa occurs. Such a columnar situation makes abortion easy and of little danger. Very soon after the entrance of the ovum tubal bleeding may result; the ovum dies and further hemorrhage expels it. The tube may return to a normal state without any evidence of the previous condition, or else a hematosalpinx may be formed if the abdominal end of the tube is closed. The ovum may, theoretically, develop to a much further degree and press the folds against the tubal wall. If development continues the villi may extend into it, and the connection of the ovum and the villi with the surrounding tissue is a loose one, as in the case of Abel and of Wyder and in the specimen Fig. 47.

2. In the intercolumnar form (Fig. 53) the ovum may rest on the wall of the tube. Any tubal fold beneath it will be compressed, but epithelium may be present in a depression. Other folds may form a capsularis, which consists then of mucosa alone; if such a capsularis be firm an intervillous space may develop. The villi at the placental site enter into the wall; here a hemorrhage may result through this invasion of the wall and of the vessels and through an invasion of the capsularis by fetal cells; or, since the capsularis does not undergo decidual change and is therefore less yielding, the capsularis may rupture. If it be torn, or if it be not closely adherent, the intervillous space is opened. Abortion, complete or incomplete—usually incomplete—is the general rule, but rupture might occur. If the abdominal end be closed a hematosalpinx or a tubal mole may represent the final outcome.

3. In the centrifugal form (Fig. 56a) the ovum sinks into the wall of the tube and an invasion of the wall and vessels by the villi may take place even up to the serosa. The capsularis is formed by muscularis and mucosa. It may rupture at its summit. The invasion of the vessels entering the intervillous space may cause hemorrhage. The villi which extend up to the serosa may cause bleeding, though their penetration is so gradual that these points are usually covered with thrombi. Finally a rupture may take place at the placental site through multiple perforations producing an arrosion. The ovum practically eats up the wall. Even though the tubal diameter be large enough to give sufficient room, this occurs. It is not the result of pressure, as may be seen in gestation at the fimbrian end, where rupture also can result. Villi which perforate the serosa may cause a very decided hemorrhage into the peritoneal cavity. When no rupture has occurred and the abdominal end of the tube is closed, only the microscope may divulge the source of such an intraperitoneal bleeding. Such minute perforations may cause collapse through hemorrhage, even though the opening be no larger than the head of a pin. Even after the death of the ovum the villi can grow, and an active tubal mole is found with continued bleeding. If they do not grow, hemorrhage continues, since no contraction can take place, as is the case in the uterus. The centrifugal form furnishes the majority of tubal ruptures. But the vast majority of these so-called tubal ruptures are either erosions or due to erosion by the perforating villi.

CHAPTER IV.

THE USUAL COURSE OF ECTOPIC GESTATION.

The theory that the tube ruptures because the ovum is too big is, as a rule, wrong for cases in the first three months. The various interruptions of ectopic gestation are all the result of hemorrhages primarily minute. The usual ending, clinically, of the gestation begins with bleeding in the tube. The invasion of the vessels of the mucosa and the tube wall and the invasion of the serosa furnish the causes for hemorrhage. The death of the fetus, as in the case of the uterus, brings about changes which result in bleeding. The primary cause is a lack of decidua. In a mucosa previously affected, when many large vessels are changed by the fetal cells and invaded by villi, an increase in tension through contraction of the tube walls furnishes an easy explanation of this hemorrhage. In the uterus the vessels are firmly embedded in the thick decidua and take a twisted course; in the tube the vessels are straight and embedded in loose connective or fetal tissue. Bleeding on the part of the capsule is possible and of frequent occurrence, since it does not undergo decidual change and may be invaded by fetal cells. The contraction of the muscle fibres on either side of the capsularis renders the rupture of this pseudo-reflexa easy because of the absence of decidual changes, and the point of rupture is usually at the summit of the capsularis. If the capsularis be composed of tubal folds the intervillous space is easily involved. If the capsularis be composed of muscularis and mucosa a decided bleeding may result if only the summit of the capsularis be torn.

Rupture of the tube almost always takes place at the placental site, which is the seat of old and new hemorrhages. The hemorrhage and loosening of the ovum which represent the clinical ending of these cases is not the first bleeding, for older ones are usually present. The various processes depend upon the ovum, the condition of the tube before pregnancy, the character of the union of the ovum with the tube, the place of union, and trauma. The reaction of the tube is limited to the area of the ovum; and in this we find the main difference between tubal and uterine

gestation. The uterus undergoes early independent growth, the tube does not. With the development of the ovum the uterus grows hand in hand, while in the tube the ovum makes room for itself and obtains its nourishment by the invasion of the tube walls. It may stretch the circumference of the tube so that its wall, as in the case of Abel, may be reduced to a layer of connective tissue so thin that rupture may result at any point.

Ampullar cases usually end in abortion, generally with hematocele. There is no obstruction, unless decided adhesions are present, and the blood is generally poured out quickly into the pelvic peritoneum or into the sac of Douglas. Such an abortion may be complete or incomplete. Rupture in this situation occurs, but very rarely. The majority of tubal gestations are situated in the isthmus tubæ nearer the uterine end. In those cases we have (1) abortion without rupture, complete or incomplete, with bleeding from the abdominal end of the tube. Generally a hematocele is found at the abdominal end. The tubes are often so curved that it is difficult for the blood to make its way to the fimbriæ, and the oozing is of a slow character. The blood extends rarely more than a very short distance toward the uterine end, because of the numerous short curves present here. (2) We may have single or multiple microscopic perforations of the tube wall by villi, causing even decided hemorrhage without apparent cause. (3) We may have macroscopic perforations or "erosions" of the tube wall, covered or not covered by thrombi, and causing great hemorrhage. (4) We may have abortion with rupture either into the free abdominal cavity with no hematocele at the abdominal end of the tube, or with partial encapsulation, in which event there may be hematocele at the abdominal end if the tube is open. (5) We may have an intraligamentous tear with hematocele at the abdominal end. In these latter cases the placental site is always on the inferior surface of the tube and the ovum has descended centrifugally to the vessels of the ligamentum latum. These are by far the most difficult cases surgically, and may require hysterectomy to remove the mass *in toto*.

CASE I. (Fig. 47).—Twenty-eight years of age, married thirteen years. One labor eleven years ago. Divorced ten years ago, and operated vaginally at Mt. Sinai for pelvic abscess, since which time periodical attacks of pain on the right side every three months. Married again twenty months ago. *One year ago, pain, fainting spells.* Three months before admission, pain. Two months before, only stains instead of menstruation. ONE WEEK

BEFORE, SEVERE CRAMPS, FAINTING SPELLS, following a metrorrhagia of several weeks. Operation showed plenty of old blood in the pelvic cavity. Tube distended, clot at fimbrian end. Other tube normal. Microscope, columnar type, with evidences of a previous old ectopic gestation.

CASE II. (Fig. 53).—Twenty-seven years old; four children, last nine months ago. Nursing. Four weeks of abdominal cramps and uterine hemorrhage. Well for one week and then symptoms returned up to admission, on which day she fainted three times on account of pain.

Operation.—Much blood; right tube distended and clot at fimbrian end. Ovarian cyst. Incomplete tubal abortion. Intercolumnar type, villi and cells of Langhans and syncytium in the wall. Reflexa consisted probably of folds involving one half the circumference of the tube. The other half of the lumen normal.

CASE III. (Fig. 56).—Old blood clots in the abdominal cavity showed the cause of the increasing attacks of pain with intervals of relief to be due to bleedings from perforations of the tube wall which were closed at times by the formation of thrombi.

CASE IV.—Patient 32 years old; two children, last three years ago. Skipped two menstrual periods. Two weeks later examined because of abdominal pain. Examination showed left-sided tumor, elastic, with pulsating vessels on the left side of the vagina. Uterus enlarged. Next day signs of hemorrhage, pale, pulse 120.

Operation showed much blood, dark clots and bleeding from the tubal end. Incomplete tubal abortion, placental mole, slight involvement of the wall. No Graafian follicle in the ovary.

CASE V.—Twenty-five years old; three children, last fifteen months ago. Three months ago first menstruation (probably nursing). Two months ago skipped menstrual period. For four weeks metrorrhagia and cramps lasting fifteen to twenty minutes and recurring as often as three to four times a day.

Operation.—Vaginal celiotomy. Little free blood. Microscope. Ovum, disintegrated by blood and surrounded by villi and tubal folds, found in the tube. Folds also present in the wall at many points. Organized clot adherent at one point. Intercolumnar and partly centrifugal, for muscularis and mucosa extend on either side partly over the mole. Organized tubal mole. No fresh bleeding.

CASE VI.—Thirty-seven years old; married eighteen years; six

children, last three years ago. Last menstruation four months ago. Operated because of abdominal pains. No blood in the peritoneum. Tube and ovary free. Continued bleeding into the tube, which was full of fresh blood. Probably would have broken through the closed abdominal end. Incomplete tubal abortion of the type of hematosalpinx. Placental mole. Decided involvement of tubal wall. Large vessel in the periphery going over gradually into the intervillous space.

These cases point to the decided danger from continued bleedings involved in tubal abortion. The general view is that tubal rupture gives much more pronounced symptoms and a much more decided hemorrhage than tubal abortion. When we consider that incomplete abortion means that villi are left in the tubal wall and that so-called complete tubal abortion means the retention of trophoblast cells, we may readily understand that bleeding may continue for an indefinitely long period. It is a fact, however, that even complete abortion may cause decided symptoms. Mandl reports two cases from the clinic of Schauta, accompanied by pronounced collapse and decided hemorrhage. In the first case no villi were found in the tube wall (see Fig. 53). In the second case, although villi were found in the blood clot in the tube, none were found in the tube wall. Like cases of tubal abortion, with symptoms as severe as are frequently the rule with tubal rupture, have been reported by Klein, Zedel, Piering, and others. It seems to me these histological and clinical evidences are of sufficient weight to destroy the view, prevailing in many minds, that tubal rupture should be treated by extirpation of the tube and that tubal abortion demands only conservative treatment. The proportion of tubal abortion to tubal rupture is probably 8 or 10 to 1. In this connection it is quite sufficient to mention the dangers arising from hematocoele. The injury to the peritoneum, the adhesions which take place, and above all the by no means infrequent occurrence of subsequent purulent degeneration of such an accumulation of blood, are only some of the injurious results avoided by prompt removal.

The possibilities are represented by the processes of abortion, microscopic perforation, macroscopic perforation, rupture, hematosalpinx, and tubal mole. In 99 cases of interrupted tubal gestation in the clinic of Schauta, a hematocoele was found 60 times—55 after abortion, 5 times after rupture. If the bleeding be very slow, the blood forms a capsule (due to peritoneal adhesions) into which the subsequent hemorrhages enter, the so-

called secondary hematocele. If adhesions are present at the abdominal end of the tube they may form a portion of the capsule. The resulting hematocele *after* rapid bleeding furnishes the primary or diffuse form. The secondary hematocele occurs much more frequently than the primary. In the 60 hematoceles found among 99 cases in the clinic of Schauta, only 4 were diffuse. Of the abortions found in the same clinic, 75 were incomplete and 6 were complete.

PART III.

OVARIAN AND PLACENTAL SECRETION.

THE RELATION OF THE CHORIONIC EPITHELIUM TO CHORIO-EPITHELIOMA.

There have been observed and reported over 150 cases of a uterine growth of exceedingly malignant character occurring after abortion and labor, or even after tubal abortion. The clinical symptoms are: (1) Pronounced uterine hemorrhage, recurring even after repeated curettage; (2) very early metastases, especially in the lungs and vagina; and (3) early death through hemorrhage, cachexia, or septic infection.

Macroscopically, these tumors are more or less localized, ulcerating, degenerating, hemorrhagic growths, frequently passing deeply into the uterine wall, or through it with involvement of the peritoneum.

Microscopically, these tumors are characterized by hemorrhagic areas, areas of degeneration, the presence of fibrin, and the involvement and invasion of capillaries and large vessels. They are especially characterized by the presence of (1) pale round and polygonal cells with pale protoplasm and pale nucleus, and (2) of large round and spindle-shaped cells with large dark nuclei and also (3) of large, irregular branches composed of polynuclear protoplasmatic masses.

These atypical growths have been variously described as sarcoma, carcinoma, carcinoma after abortion and labor, and as sarcoma and carcinoma causing abortion.

Sänger, in reviewing these cases, found a decided resemblance in their characteristic elements, and came to the conclusion that the decidua cells were the cause of the growth, giving it then the name of deciduo-sarcom or deciduoma malignum.

As a result of the investigations of Fränkel, and later of Marchand, attention was called to the fact that those cells which so closely resembled decidua cells were really of fetal origin and were, in fact, *the cells of Langhans*, while the spindle-shaped and grouped masses of polynuclear protoplasm were of *syncytial origin*.

From all sides, especially in England and Germany, this view was attacked. It was pointed out how baseless was the view that fetal cells could produce a growth of this malignant character, differing from carcinoma only in the fact that metastases resulted through the blood channels instead of the lymph paths.

This controversy is to-day by no means settled, many holding the view that these tumors are sarcomatous, *originating from the decidua cells*. The giant cells and the protoplasmatic masses are referred, likewise, to changes in the decidua. Others hold that these growths result from the epithelial covering of the villi. That these cells, if they are of fetal origin, should be mistaken for decidua cells is a natural error, for we know that even in the normal processes *a positive distinction is often very difficult*. It is to be noted that many investigators have called the typical trophoblast cells in tubal placentation, too, decidua cells. Still others lean to the view that the stroma of the villi plays its part.

On the other hand, among those who hold that these growths originate from the chorionic covering a division of sentiment exists; for those who consider the syncytium and cells of Langhans to be of uterine origin class these growths as carcinoma and sarcoma of a somewhat atypical character. Those who believe, as we have shown, that the epithelial covering of the villi is of fetal ectodermal origin, and who therefore also class these tumors under the category of carcinoma, are introducing into pathology a new element.

A factor which has served to clear our views on these various disputed points is the knowledge that fifty per cent of these malignant uterine growths, commonly known as decidioma, follow the presence of hydatid mole.

In hydatid mole we find the same elements as in normal placentation, only that these elements are excessive in number and size. Hydatid mole represents a hypertrophic growth of the chorionic covering, accompanied by dropsical swelling of the chorionic stroma. As is well known, the covering of the villi consists of two layers, an outer syncytium, an inner, the cell layer of Langhans. The growth concerns both the syncytium and the cell layer of Langhans. The abnormal element is the occurrence of very large cells with immense nuclei in large number, and a decided growth of the syncytium, accompanied by the formation in the latter of large vacuoles.

Leaving out of consideration those cases malignant because of the diffuse and deep infiltration of the uterine wall by the cystic

villi, by no means are all hydatid moles of a malignant character. An attempt to distinguish between the benign and malignant cases was proposed by Neumann. He observed, in three cases subsequently resulting in the so-called deciduoma, large cell elements in the stroma of numerous villi which he considered to be infiltrating elements of the syncytium. He observed, further, an abnormal infiltration of cell groups through such syncytial elements. Investigation of subsequent cases shows that malignant forms are not always preceded by such changes in the hydatid mole, while others have found these changes and yet no malignant growth occurred.

Even the occurrence of metastases is no proof of malignancy, for Pick reported a case with a metastasis of villi in the vagina and yet the patient recovered. We know that fetal cells are given off from the normal placenta into the maternal circulation. Even the normal placenta, as Pick believes, may give metastases of villi, and these may (1) degenerate or (2) grow slightly or (3) produce the same syncytial growth as is observed in benign hydatid mole. (4) Primary malignant growths may originate, and have originated, from such metastases.

Malignancy, in the case of hydatid mole, is not then to be judged alone by the occurrence of metastases. Those cases which subsequently develop into the so-called deciduoma evidence their malignant character *by the ability of their cells to grow in an unlimited manner*, aided by the character of the tissue which permits or also aids this growth. Various theories have been propounded in explanation of this phenomenon. 1. Through the syncytium there is a constant exchange of products, and after hydatid mole, or on the occurrence of abortion or labor or any process causing the removal or death of the fetus, this exchange ceases. The fetal cells then, if in a favorable surrounding, are supposed to use this nutrition for themselves and increase until an unlimited growth results (theory of Marchand). 2. Ribbert considers the unlimited ability of certain malignant tissues to grow to be due to the separation of their mother cells from their normal connections. 3. As is well known, Cohnheim considered displaced embryonal cells to be the future source of many benign and malignant tumors.

An interesting power or potential retained by displaced cells is that of differentiation. We know that displaced cells, cells removed from their normal relations, are able, after an interval of many years, to grow and produce structures of varying form.

This is best exemplified in the case of dermoid cysts, for their character distinguishes them from all other tumors. The later in the stage of embryonal development these cells are displaced the more simple is the structure of the resulting dermoid; the earlier in the period of embryonal development their displacement occurs the less differentiated are these cells. For that reason embryonal cells displaced in the early weeks produce tumors of complicated character, for their potential as regards differentiation is great. Cells displaced at a later period possess a lesser potential as regards differentiation, while those epithelial and connective-tissue cells displaced very late, as at points where the skin only remains to be united, and at the branchial clefts, produce only the simplest form of dermoid growth. They produce only cells of the same character and structure as the parent cell if the stage of complete differentiation had been already reached. If the displacement occurs before this period, such elements are found in the subsequent growth as the parent cells would have produced had they remained in their normal situation. Such early cells, however, reproduce far greater growths and far more extensive tissues than would have resulted had they not been displaced. This is evidenced by the fact that no loss of any normal tissue or structure results. In dermoid cysts of the ovary, for instance, very large and complicated tumors are found, resulting from the displacement of ectodermal and mesodermal cells by the Wolffian body, and yet the maternal body is otherwise normally developed.

In the genital tract, especially, we find numerous evidences of another cell potential, that is, the ability to first display accelerated growth after a lapse of many years. We find in the uterine wall, under the peritoneum, in the broad ligament, and in the ovary, generally after puberty, epithelial and glandular growths, sometimes of considerable size, resulting from the displacement of cells of the Wolffian body. In the fetus and in the newly-born, hundreds of uteri and appendages have been examined and yet relatively few such displacements of Wolffian-body cells can be found. This means that the displacement concerns simply embryonal cells of this organ, which even at a much later period possess the power to develop the same structures as the parent organ. This growth takes place, as a rule, at and after puberty, and the same is true in the case of dermoid cysts.

We find, then, that the general stimulation of tissue and cell

growth occurring after puberty may influence some embryologically displaced cells in the same manner.

We find, on close investigation, that almost all ovarian and parovarian cysts result from the continued growth of structures which in the embryo were functioning organs, but which in the fetus and in the adult are supposed to undergo regressive changes, namely, the epoöphoron and the paroöphoron, constituting the two divisions of the Wolffian body. Papillomata of the ovary, in all probability, also develop from cells of these supposedly regressive structures. Ovarian cysts also frequently show papillomatous changes. These are, strictly speaking, only huge increases of the characteristics of the primary embryonal organ.

Not infrequently these papillomatous growths are macroscopically of a malignant character, in that they break through the covering of the cyst or through the ovary, grow without restriction, invade the peritoneum, infiltrate the surrounding organs, and produce cachexia.

Such changes may also display the microscopic characteristics which we attribute to carcinoma—that is, a continued growth of the epithelium, a breaking through of the membrana propria, and an infiltration, microscopically, of the tissue surrounding the epithelial cells. In other words, we find a continued unlimited growth of cells, reproducing, even though in a changed relation, the character of the mother cell.

These smaller and larger reproductions of the Wolffian body, these cystic growths originating from the Wolffian-body cells, these papillary and malignant growths originating from the same source, as well as dermoid cysts, furnish us with evidence of the ability of regressive cells and organs, and cells removed from their normal relations, to undergo a more or less unlimited growth, even after lying dormant for many years. These cells, however, are cells of the patient and are open to the same influences as normally situated cells.

In the case of chorio-epithelioma, however, we find fetal cells, often only a few weeks old, possessing naturally no great potential as regards differentiation, but an exceedingly high potential as regards their ability to grow. The energy and potential of these cells may be appreciated from the fact that the earliest case occurred two weeks after the interruption of pregnancy, while the latest occurred nearly four years after hydatid mole.

The most prominent point of a ripening Graafian follicle is poor in blood supply and is called the stigma folliculi. It is here that the opening takes place which furnishes an outlet for the ovum. This opening is probably the result of the reaction or chemical effect produced by the ripe ovum, for, in the newly-born and in children, follicles of the same size and even larger ones exist without bursting—the so-called atresic follicles.

After ovulation the ovum is thrown out into the abdominal cavity, and then, influenced by the wave movement of the ciliated epithelium of the tube, the fimbriæ of the ampulla, and the fimbriæ ovaricæ, finds its way into the uterus. The wave movement of the ciliated epithelium causes a current in the peritoneal plasma which directs the ovum into one or other of the tubes.

A fecundated ovum embeds itself in the lining of the uterus through centrifugal descent. The ovum then causes a reaction in the surrounding tissue and a dilatation of the surrounding lymph spaces, so that a resulting localized edema takes place. In addition a dilatation of the capillaries is produced.

The outer layer of the ovum develops into what is known as the trophoblast, which is a product of the ectoderm, and from it develop the cells of Langhans and the syncytium.

Shortly after the ovum is embedded in the mucosa a connection between the trophoblast and the maternal blood takes place through a rupture of the capillaries. The maternal blood then bathes the ectodermal trophoblast. This opening of the maternal vessels occurs, however, *before the formation of villi*; and the cells of the trophoblast may therefore enter the maternal veins at the very earliest period.

The compact layer of the decidua is the zone which envelops the ovum. The trophoblast at points may extend far into the compacta, for *the cells have a decided power of wandering*. The trophoblast, therefore, invades the maternal tissues even at the earliest period.

A gradual transition of trophoblast cells into syncytial cells, and a gradual change of trophoblast nuclei to syncytial nuclei, take place through the corrosive action of the maternal blood, and elements of the maternal blood aid in forming the syncytial protoplasm. The syncytium does not originate from the maternal endothelium, nor from the uterine epithelium, nor from the decidua cells.

Just as, in the early stages, the trophoblast invades the decidua, so, after the formation of villi, is the future course of the

ectodermal trophoblast and of the syncytial cells of a destructive character so far as the decidua is concerned. The trophoblast and syncytium invade the maternal tissue and mingle with it. They infiltrate the decidua and bring it to destruction. The trophoblast and syncytial cells erode the capillaries and blood vessels, the blood in turn changing fetal cells to syncytium.

The invading trophoblast and syncytial cells have at all times a great power of wandering. They enter, between bundles of muscular and connective tissue, into the lymph spaces and into the blood vessels. At full term the uterine wall is infiltrated with fetal cells of a syncytial character.

From the very earliest moment *fetal cells are continually entering the blood of the mother*, not only in the primary intervillous space but in the fully formed intervillous space, as well as through the vessels of the uterine decidua and wall.

CHORIO-EPITHELIOMA.

Under chorio-epithelioma we distinguish two forms, the typical and atypical. In the typical form (Fig. 60) we find large, round, polyhedral cells with strikingly large, very irregular, lobulated nuclei which stain very deeply and often degenerate, forming vacuoles. The protoplasm is relatively scanty. These cells are capable of great wandering and are found more or less isolated between the muscle and the connective-tissue bundles (Fig. 63), in the lymph spaces and in the vessels. They form the advance guard in the way of infiltration. There are, further, irregular bridges of protoplasm containing scattered or grouped nuclei of various sizes (Fig. 61). Many of these groups of nuclei are the same large, irregular, lobulated nuclei as were observed in the form just mentioned. In addition are found irregular masses of protoplasm containing many small nuclei. The character of the latter is identical with normal syncytium.

The irregular groups of protoplasm containing grouped nuclei of various sizes are undoubtedly of syncytial character, for they result through the blood surrounding and infiltrating the cells of Langhans, and it is very evident that these cells form the aforementioned grape-like nuclei (Fig. 63). The isolated large cells are likewise of syncytial character. They have generally been mistaken for decidua cells. They may be distinguished from the cells of Langhans, for the latter are pale, polyhedral groups of distinctly epithelial character. They are rich in glycogen and therefore often contain vacuoles. The nuclei are large but pale.

The cells of Langhans are better illustrated in the atypical form (Fig. 64) where the syncytial elements are relatively in the background. In fact, no more and no different syncytial cells are present here than in normal gestation. The trophoblast cells lie closely grouped and are surrounded by syncytial elements in

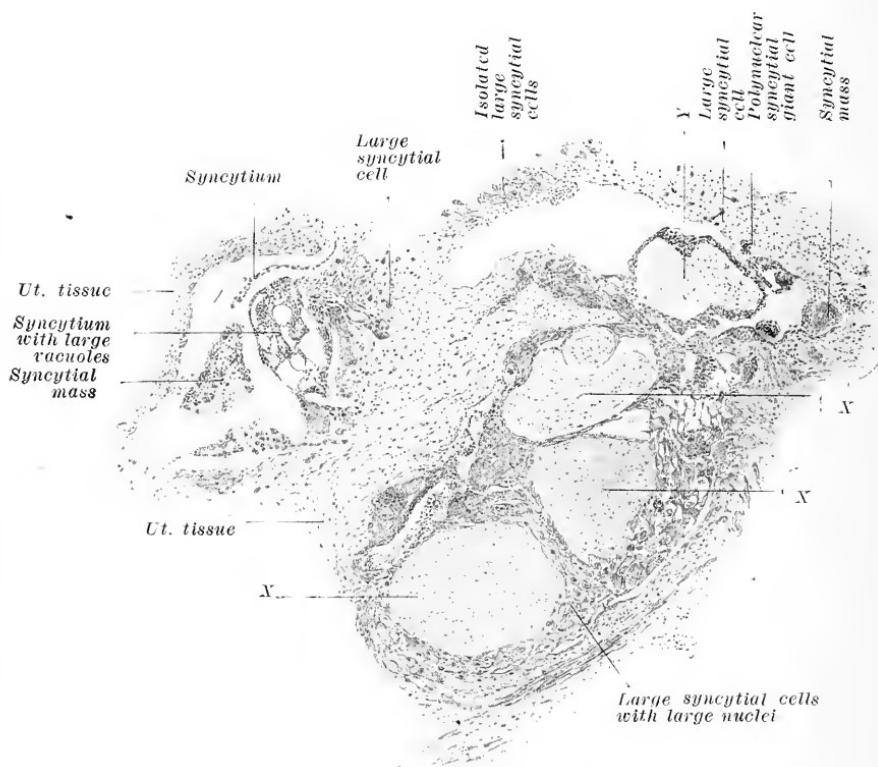


FIG. 60.—Low-power drawing of the typical form of chorio-epithelioma, showing the uterine wall invaded by chorionic elements. *X*, *X*, *X*, three areas of dense connective tissue surrounded by chorionic epithelial elements and resembling chorionic villi. *Y*, connective tissue centre surrounded by polynuclear syncytial mass of considerable thickness, probably a villus.

quite the same manner as in normal gestation, or especially in tubal gestation. They are polygonal cells, concerning which different views have been held. They have been called decidua cells. No vessels of their own, however, are present in these epithelial-like groups, and their character, their structure, and their arrangement so closely resemble the trophoblast cells observed in normal gestation that any other view is not to be con-

sidered. These epithelial-like cells and the syncytial masses of various forms all originate from the trophoblast cells.

In these growths newly formed villi have not yet been found—a proof of the limited power of differentiation possessed by the



FIG. 61.—Upper left-hand corner of Fig. 60 highly magnified, showing the character of the polynuclear syncytial masses. Along the right and lower borders are larger isolated mononuclear syncytial cells.

trophoblast cells alone. It may be said, therefore, that two forms of this tumor exist, the first typical, the second atypical. The former cases are so characteristic that they cannot be mistaken. The latter have been so frequently called carcinoma by eminent au-

thorities that our belief that many of these are overlooked and incorrectly diagnosed is certainly true.

A study of the histology of so-called deciduomata, and a comparison of their structure with the structure of normal placental elements, prove these tumors to be of fetal origin. The cells from which they develop are the cells which cover the chorionic villi. Since these are epithelial in character, these tumors, belonging as they do to the most malignant forms, should be called chorio-epithelioma.

We have, then, in the chorio-epitheliomata a reproduction of

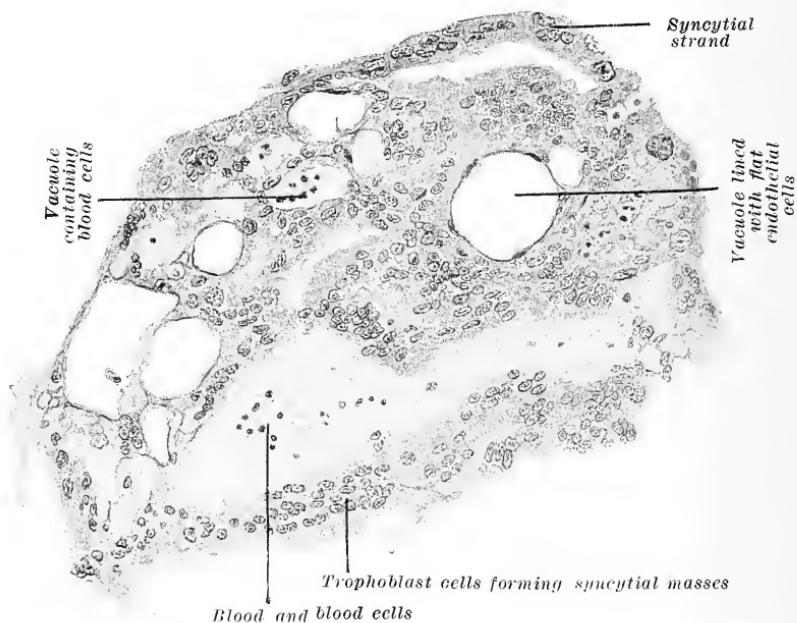


FIG. 62.—Highly magnified area of Fig. 61, showing finer characteristics of syncytial masses. Change of trophoblast cells *en masse* into polynuclear, vacuolar structures.

the same constituent elements as are found in normal placentation and as are observed in benign and malignant cases of hydatid mole. These cells exert the same influence and effect on the maternal tissues as do the fetal cells in a normal uninterrupted pregnancy. They invade, as do the normal trophoblast cells, the maternal decidua and destroy it. They infiltrate and erode the walls of the vessels. They invade and infiltrate deeply, too, the uterine wall. They advance either as distinct Langhans or trophoblast cells or

as syncytial cells, or else they undergo in their advance a change from the former to the latter, especially when in contact with maternal blood, as in the case of placentation either uterine or tubal. Their invasion of the maternal vessels and capillaries gives them, from their earliest existence as malignant cells, the opportunity of invading the maternal circulation with a resulting early formation of metastases. Their ability to erode the vessels causes profuse and constant bleeding. Their ability to destroy the maternal tissue as they advance produces larger and smaller areas of degeneration and necrosis accompanied by the presence of much fibrin. These cells preserve their ability to grow when

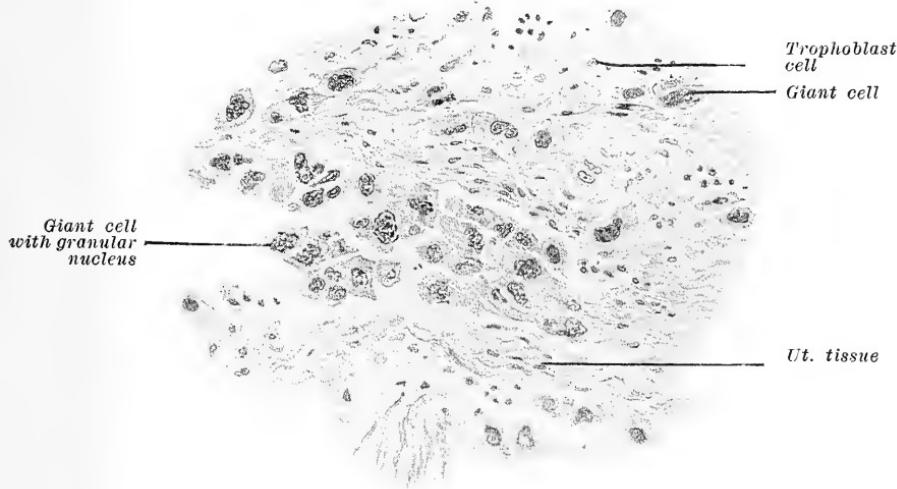


FIG. 63.—Highly magnified area of Fig. 60, showing character of isolated mononuclear giant syncytial cells and the infiltration by them of the uterine tissue and lymph spaces.

they reach their new locations, with the result that they produce in the various organs, but most frequently in the vagina, malignant nodules of the same character as the parent growth. In fact, these secondary nodules have in some cases been observed before the character of the uterine symptoms called attention to the presence of malignant conditions in the uterus.

The fetal cells producing a chorioma are situated in the most favorable surrounding. They have been performing practically malignant functions in that they have destroyed, even during normal placentation, maternal tissues, and have invaded maternal vessels, and have been carried off into the maternal circulation.

When connected as part and parcel of an ovum, when feeding and nourishing the fetus with the products of the maternal blood which have passed through them, they are, so to speak, under control of the parent organism the ovum, yet when released from this connection they continue an independent growth of their own. It is quite probable that in hydatid mole the edematous swelling of the chorionic stroma is due to interference with the proper exchange between the fetus and the mother, due to a more or less increased and independent growth on the part of those cells whose function it is, normally, to aid and permit of this exchange. It is likewise probable that the growth of the chorionic

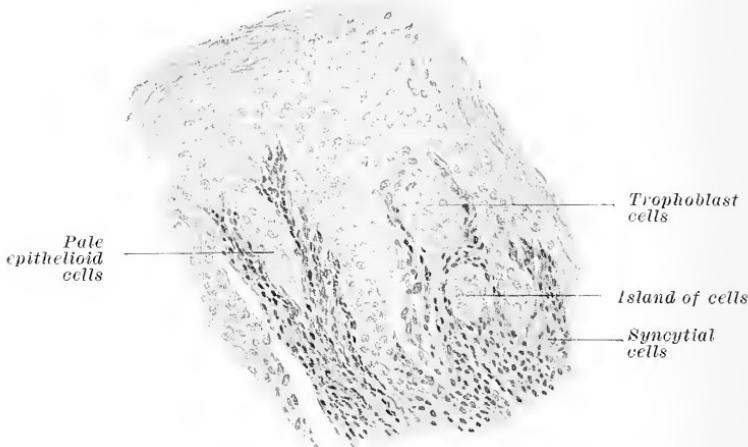


FIG. 64.—High-power drawing of atypical chorio-epithelioma greatly resembling carcinoma.

cells in chorio-epithelioma takes place during the pregnancy and is rather the cause than the result of abortion.

We have observed in the development and change of trophoblast cells to syncytium that the closely grouped cells, when vascularized, change to plasmoidal or syncytial cells. That the blood of the mother furnishes the greater portion of the protoplasm of these syncytial cells has been clearly shown. Therefore their production and growth, even in normal conditions, depends upon their taking up from the mother essential elements, while the trophoblast cells themselves furnish the nuclei. Therefore the growth of so pathological a tumor as a chorio-epithelioma is not absolutely a reproduction of fetal cells, but is in a more or less direct manner a maternal production also.

The invasion and destruction of maternal tissues in normal gestation occurs within certain fixed limits, and the fetal cells entering the maternal circulation undergo no future growth. What preserves this balance? What limits and controls the potential of the parasitic fetal cells? In hydatid mole, and especially in chorio-epithelioma, the fetal cells are no longer held in check and they possess the power of unlimited growth. What has upset the normal balance?

When the fecundated ovum enters the uterus, it destroys the surface epithelium under it and descends actively into the decidua. It produces a decided reaction in its immediate circumference, so that even in its earliest stages it evidences a chemical power. When the maternal blood makes its exit from the capillaries it ought to coagulate, but does not. It circulates against the fetal cells which have the power to prevent coagulation. The trophoblast and syncytial cells are bathed by maternal blood and enter the circulation; therefore the ovum has a certain *enzyme action* and the fetal cells may be said to furnish or represent a placental secretion.

On the other hand, the blood contains elements which exert a corrosive action on the trophoblast cells, changing them to syncytium. The resulting syncytial cells then cover the villi; they play the part of endothelium (which they then greatly resemble) and protect the cells of Langhans and the stroma from the corrosive influence of the blood. That the individual cells in chorio-epithelioma have the power to grow without limit, and that the cells entering the circulation have the energy to produce malignant metastases, shows that *the decidua and the blood no longer have the power to limit and control their growth.*

In reviewing the anatomical and physiological characteristics of the female sex before and during pregnancy, and bearing in mind the normal changes of pregnancy (such as the cessation of menstruation, the act of labor, etc.), but especially the pathological states occurring almost entirely or exclusively in connection with pregnancy (such as osteomalacia, eclampsia, and chorio-epithelioma), we are forced to the conclusion which, while partly theoretical, is nevertheless logical.

Ovarian secretion has a great trophic influence upon the uterus. It stimulates the growth of the round cells of the stroma into decidua cells. The lining of the uterus is truly a lymphoid tissue, and it is the great development of the decidua and its secretion which prevents the involvement and macroscopic perforation of

the uterine wall by the placental villi. Although it may be said that ovarian secretion stimulates the growth of the fetal cells and that certain elements in the blood hold their growth in check, it is probably the ovarian secretion in the maternal blood which aids the decidua in holding the placental development within normal limits, and which renders the trophoblast cells and the syncytial cells entering the circulation innocuous.

In normal placentation the human organism furnishes us with a process parallel to that occurring in certain bacterial infections, that is, the production of two opposing toxins or ferments: (1) a blood element, probably the ovarian secretion, and (2) the placental secretion.

In the normal woman the ovary is responsible for the periodical loss of blood known as menstruation. This process is due to a secretion furnished by the ovaries, for on their removal this process ceases and the reduction of oxygen exchange amounts to twenty per cent. This secretion stimulates various functions of the body, and at regular periods an outlet for this secretion occurs. Every menstruation represents, in addition, the birth of a non-fecundated ovum, that is, a labor *en miniature*.

When, however, fecundation and development of the ovum take place, the ovum and its enzymes nullify the menstrual stimulation of the ovarian secretion. The trophoblast cells invade the maternal decidua which is stimulated by the ovarian secretion, and likewise enter the blood of the mother. A normal gestation is accompanied by the stimulating effects of the retained ovarian secretion and these two enzymes are then opposed in their action. No menstruation occurs, for the placental secretion has nullified the action of the usual forces.

At the end of nine months, when the ovarian secretion is sufficient in amount or character to overcome the neutralizing action of the enzymes of the ovum, labor occurs, that is, the same process as is observed in a minuter degree in menstruation, for menstruation, as said before, is a labor *en miniature*.

Remembering the constitutional action of ovarian secretion, it may be said that if, shortly before, during, or after labor, there is an overwhelming superiority of the ovarian secretion over the placental or an opposite mal-relation between ovarian and placental secretion, the constitutional involvement known as eclampsia results.

Following the analogy further, it may be said that chorioepithelioma is due to the fact that the resistance to the fetal en-

zymes and fetal cells offered by the blood and the ovarian secretion is insufficient to hold the growth of the fetal cells in check. Chorio-epithelioma occurring generally after abortion or hydatid mole is certainly the cause rather than the result of the abortion. Chorio-epithelioma represents a more advanced stage than that of hydatid mole, but both of these conditions follow the normal processes in their course and growth. The only difference is the power of unlimited growth possessed by the chorionic cells in these pathological conditions. The difference in the resistance offered by the patient points to a constitutional element as an important factor in the etiology of chorio-epithelioma.

COLUMBIA UNIVERSITY LIBRARIES



0041065280

BORROWER N

RG586

Bandler

Uterine and Tubal Gestation

322

Copy 1

